MICROLAB STAR Line

Operator's Manual

CE

Important Notice

Table of Contents

1. Information about Safety and Security

HAMILTON's MICROLAB STAR, MICROLAB STARlet, and MICROLAB STARplus (in the following summarized as ML STAR) is a pipetting instrument. This Operator's Manual is designed to get the most out of your ML STAR.

ATTENTION

You shall carefully read through the entire manual before beginning to operate your instrument. This first section must be read with particular attention. It contains important information about safe use of the ML STAR.

1.1 About this Manual

This manual refers to the VENUS four Software Version 4.5 for the ML STAR.

NOTE

Refer to the contents of the Data Storage Device (see Section 8.1.33 VENUS Software).

This manual helps users operate the ML STAR correctly and safely. To achieve this goal, the manual describes the different components of the ML STAR and their functionality. The manual describes both the hardware and software of the ML STAR in depth, enabling the user to operate the instrument.

After introducing you to the various parts of the ML STAR, the manual will describe step by step how to operate the instrument. After you have read through this manual, you should be capable of operating the ML STAR.

Warnings and ***Notes*** are part of this manual to emphasize important and critical instructions. Symbols used in this manual for these warnings and notes are:

ATTENTION

Any special problems, warning or important information will be accompanied by this symbol. Read these items carefully and follow the instructions given.

NOTE

Information is given to the programmer that is useful but not essential to the task at hand.

[...] Push buttons and their corresponding description.

"**...**" Description for all kinds of entry fields, control fields, check boxes, lists, etc.

___ Underlined texts are references to Manuals, Figures, Sections, etc.

1.2 Additional ML STAR Manuals

For the programmer of the system, the VENUS Software Programmer's Manual describes all the features of the Microlab STAR VENUS Software. Sample methods of typical applications guide you through the programming. A detailed software reference for the ML STAR can be found in the Help Function of the VENUS Software.

NOTE

Refer to the contents of the Data Storage Device (see Section 8.1.34 ML STAR Manuals).

The Total Aspiration and Dispense Monitoring (TADM) is an additional safety tool for the pipetting process. The TADM feature allows optimization of the entire pipetting process by reading out the pressure curves and comparing them to a tolerance band in real-time. The description of its functions and how to work with TADM is described in the TADM Manual.

VENUS Dynamic Scheduler is a software tool for organizing and controlling the workflows of a laboratory equipped with HAMILTON instruments as well as other manufacturer's instruments. The use of this tool is described in the VENUS Dynamic Scheduler User Manual.

VENUS Data Base Plus is a software tool that allows using additional functions on the VENUS Database. It allows tracking over multiple runs and the use of an SQL Server via the network. The use of this tool is described in the Help Function of the VENUS Database Plus.

Further manuals for specific applications, standard solutions, workstations, features, etc. come together with the dedicated ML STAR instrument. Please refer to these specific manuals for the respective application.

1.3 ML STAR VENUS Software License

NOTE

HAMILTON Bonaduz AG and HAMILTON Company confirm that, there are no license payments foreseen, when using the following HAMILTON ML STAR software.

- ML STAR VENUS Software

1.4 Intended Use

The ML STAR is a robotic pipetting workstation which is classified as a general laboratory instrument and not as an *in vitro* Diagnostic device. It is intended to automate routine pipetting tasks and the required transports of labware such as microplates and disposable tips used in the pipetting assay.

1.5 Intended User Group Profiles and User Environment

1.5.1 User Group Profile

Title of user group	Demographic data (Age range, Gender, specific physical attributes)	Typical tasks (supported by the medical device)	Expected qualification, expected job experience
Primary user: • Laboratory Operator	• Age: approx. 20-65 Years • Gender: All • Specific physical attributes: Education in good laboratory practice (GLP)	Operating the pipetting instrument and user maintenance	• Knowledge of all reference documents • Knowledge of good laboratory practice • Knowledge of the ML STAR Software • Knowledge of the ML STAR • Trained for operation
Secondary user: Field Service Engineer	• Age: approx. 20-65 Years • Gender: All • Specific physical attributes: Education in service techniques	Service tasks, Preventive maintenance, Maintenance & Verification	• Completed training as a Service Engineer • Good English skills

1.5.2 User Environment

Title of user group	Organizational / social environment during task completion	Physical environment during task completion	Typical equipment used (in conjunction with the medical device)
Primary user: Laboratory Operator	Education in good laboratory practice (GLP) and the Laboratory Operator of the ML STAR must have attended an official training course.	General Laboratory	Labware (Sample tubes, Reagent container, etc.) Liquids Cleaning agent Personal protection clothing (Lab gloves, …)
Secondary user: Field Service Engineer	HAMILTON Trained Field Service Engineer, education how to operate and repair / maintain the ML STAR Customer-oriented, flexible and communicative	Different environments depending on customer site	Tools, Service manual, adjustment equipment, spare parts, etc.

1.6 Operation

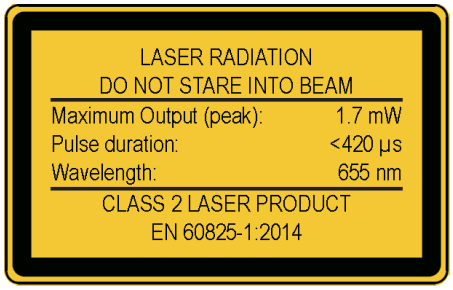
The operator of the ML STAR must have attended an official training course. The procedures contained within this manual have been tested by the manufacturer and are deemed to be fully functional. Any deviation from the procedures given here could lead to erroneous results or malfunction.

During instrument operation, stand clear of all moving parts and the working deck of the instrument. In general, never lean over or into the instrument while it is in operation.

ATTENTION

LASER BEAM

The Barcode Scanner on the optional Tube Twister Channels of the ML STAR has a Class II Laser Diode. Do not stare into the beam.



Optional Tube Twister Channels

If the ML STAR is used in a manner not specified by HAMILTON, the protection provided by the equipment may be impaired.

NOTE

A breakdown of the power supply during a run may cause loss of data. If data loss is unacceptable, use an independent power supply or an Uninterruptible Power Supply (UPS).

The ML STAR is operated by means of control software. Specific methods, programmed by specially trained method programmers according to assay descriptions serve as job instructions for the control software. The operator shall not modify methods except in consultation with the persons responsible for the programming and/or the assay supplier. Improper method changes can lead to false results.

In order to ensure error-free operation of the instrument, the content of the maintenance and verification procedures needs to be understood. In addition, the mandatory verification process must be carried out within the required timeframe and the verification process must be documented.

After the lifetime of the instrument has terminated, please contact HAMILTON for disposal regulations (see Section 8.4.6.1 Recycling of a ML STAR Line Instrument). Give due regard to the parts labeled with the biohazard warning symbol in Section 1.10.6 Hazards.

1.7 Malfunction / Troubleshooting

ATTENTION

For ML STAR malfunction and for troubleshooting, contact your local HAMILTON Representative.

1.8 Training

Training courses will be held by your HAMILTON representative. Please feel free to contact your HAMILTON representative to arrange for an operator training.

1.9 Legal Regulations

All local, state and federal laws which prescribe the use, application, and/or the handling of dangerous materials in connection with the instrument must be strictly followed.

1.10 Safety Precautions and Hazards

The following section describes the main safety considerations, electrical and biological, in operating this product, and the main hazards involved.

ATTENTION

- Only authorized personnel may have access to the instrument.
- The laboratory or operator of the instrument is responsible for ensuring that the environmental conditions for safe operation of the instrument are met at all times.
- The laboratory or operator is responsible for any improper handling. It must be ensured that untrained or unauthorized personnel cannot manipulate or interfere with the instrument while it is in use.

ATTENTION

Read the following safety notices carefully before using the ML STAR.

1.10.1 General Precautions

1.10.1.1 Instrument Installation and Relocation

The ML STAR must be installed or relocated by a HAMILTON Trained Field Service Engineer. Installations of instrument options and accessories are also done by a HAMILTON Trained Field Service Engineer.

ATTENTION

ELECTROSTATIC DISCHARGE

The ML STAR conforms to European norms regarding interference immunity. However, if the ML STAR is subjected to electromagnetic fields or if static electricity is discharged directly onto the ML STAR, its Liquid Level Detection ability may be negatively affected. It is therefore recommended that the ML STAR is kept away from other equipment that emits too high electromagnetic fields (EMC) in the laboratory, and that static electricity is minimized in its immediate environment.

NOTE

During operation, the ML STAR must be shielded from direct sunlight and intense artificial light. The instrument shall be positioned in the laboratory in a way permitting personnel to access the front and sides of the instrument in order to operate, maintain, open and close the protective covers. Accordingly, to calculate how much space is needed, consider the dimensions of the instrument (see Section 7.1 Basic ML STAR Specifications) and sufficient space for a person to move and work comfortably.

ATTENTION

Never lift a fully installed instrument to transport it from one place to another. It must be re-installed in the new work location by a HAMILTON Trained Field Service Engineer.

ATTENTION

The instrument weighs more than 150 kg. Necessary precautions shall be taken when transporting the instrument.

1.10.1.2 Maintenance, Service and Repair

Only HAMILTON Trained Field Service Engineers are authorized to perform mechanical maintenance on the ML STAR.

For repair or shipment, all mechanical parts must be put in their rest positions. An ML STAR sent away for repair must also be decontaminated (see Section 6 ML STAR Instrument Decontamination) if it was in a laboratory environment with infectious or hazardous materials. The ML STAR must be repacked in the original shipping crate only by a HAMILTON Trained Field Service Engineer (contact your local HAMILTON representative). There must be no containers or tips on the ML STAR during transportation.

The HAMILTON Trained Field Service Engineer and the laboratory share the responsibility for the Installation Qualification (IQ) and the Operation Qualification (OQ), i.e. Verification and Training. The Process Qualification (PQ) is the sole responsibility of the laboratory.

Only original HAMILTON ML STAR specific parts and tools may be used with the ML STAR, e.g. Carriers, Racks, CO-RE Tips, and Tip Waste Chute or Tip Waste Bag. Commercially available Liquid Containers, such as Microplates and Tubes, may of course be used.

1.10.1.3 Qualities of HAMILTON Solutions

The table below shows the Qualities of HAMILTON Solutions which depicts the responsibilities of both HAMILTON and the Customer.

Quality of HAMILTON Solutions				
	DQ	IQ (Installation Qualification)	OQ (Operation Qualification)	PQ (Process Qualification)
Responsible	HAMILTON: System Customer: Assay	HAMILTON	HAMILTON/Customer	Customer
			Verification of the customer's specifications Training	

1.10.1.4 Operating the Instrument

When using the ML STAR, Good Laboratory Practices (GLP) must be observed. Suitable protective clothing, safety glasses and protective gloves must be worn, particularly when dealing with a malfunction of the instrument where the risk of contamination from spilled liquids exists (see Section 1.10.6.2 Safety Measures).

ATTENTION

When using flammable or explosive fluids or vapors, necessary precautions must be taken. There is a high risk of damage to health and to the instrument. The instrument is not explosion-proof

When using aggressive or corrosive liquids, ensure that filter tips are being used. Do not re-use tips. Replace stop disks and O-rings more frequently; make sure to perform Preventive Maintenance to the instrument in shorter intervals since the system will need a higher level of support and maintenance. To avoid corrosion on the deck, use an exhaust suction device to remove corrosive fumes away from the deck.

ATTENTION

ELECTROSTATIC DISCHARGE

Electrostatic discharge can cause damage to the instrument and can influence labware behavior and stability, therefore avoid any electrostatic charge onto labware and disposable tips during handling and manual loading of these to or from the ML STAR.

ATTENTION

During instrument operation, stand clear of all moving parts and the working deck of the instrument. Never lean over or interfere with the instrument while it is in operation! Never reach into a running instrument with your hand(s)!

If not followed as mentioned above, there can be injuries or contamination!

ATTENTION

If the ML STAR is used in a manner not specified by HAMILTON, the protection provided by the equipment may be impaired.

ATTENTION

Any persons operating the ML STAR and the PC running the VENUS Software must have attended an official training course. Any departure from the procedures given here could lead to erroneous results or a ML STAR malfunction.

1.10.1.5 Switching on the ML STAR

The power switch is located on the left front frame. When switched OFF, the power switch is toggled to the down position and the light within the switch is not illuminated. To switch ON the power switch, toggle the switch to the up position. The switch internal light will be illuminated.

Power Switch OFF

Power Switch ON

1.10.1.6 Tip Waste Bag / Tip Waste Chute

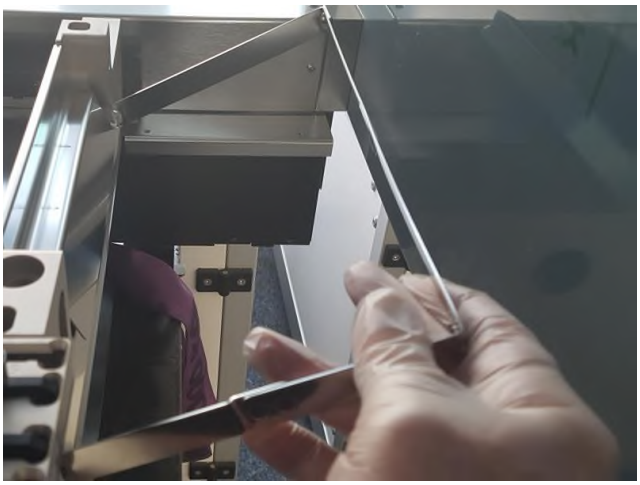
Tip Waste Bag / Tip Waste Chute:

- The Tip Waste Bag collects used tips for disposal
- The Tip Waste Chute funnels used tips into a large waste container in the cabinet below the instrument
- The Tip Waste Bag / Tip Waste Chute is suspended from the Tip Waste Block
- The Tip Waste Block and Tip Waste Bag / Tip Waste Chute are to be managed as biohazardous
- The Tip Waste is always to be regarded as contaminated

1.10.1.7 Installing the Tip Waste Bag / Tip Waste Chute

1. Remove the Waste Frame:
    a. Lift the right side of the Waste Frame.

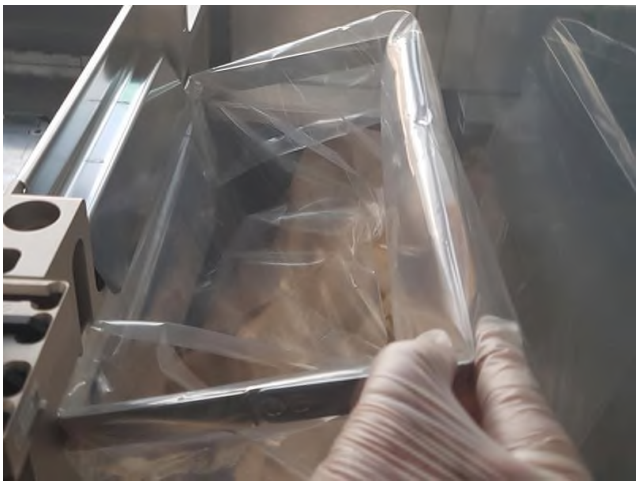
    b. Lower the left side of the Waste Frame from under the ledge of the Waste Block.



    c. Waste Frame removed.

2. Insert a Tip Waste Bag or Waste Chute.

3. Place the Tip Waste Bag and Waste Frame into the correct position under the ledge of the Waste Block.



4. Place the Waste Frame in a horizontal position.

ATTENTION

BIOHAZARD

The tip waste and the waste bag / waste chute are always to be regarded as contaminated.

1.10.1.8 Method Programming

ATTENTION

Never disable any safety measure.

Programming and validation of new methods is the responsibility of every customer.

Pay close attention to all instructions and parameters given in test kit package inserts when programming test methods for the ML STAR.

Perform test runs first with water and then with the final liquids, prior to routine use. Test all liquid classes which will be used. A newly programmed test method must first be run on the ML STAR with the final liquids, prior to validation of the method and routine use. The method programmer must supervise this run.

ATTENTION

Before using any newly created or modified method for routine test purposes, a comparison study between the method previously used and the new one must be carried out by the laboratory supervisor to ensure that the processing and data evaluation of both methods produce equal results.

NOTE

When working with samples, which will be used in particularly sensitive tests, take into account the evaporation and condensation that may occur while the method is running. During method programming, there should be no delays between aspirate and dispense of samples.

ATTENTION

When pipetting without Total Aspiration and Dispense Monitoring (TADM), incorrect pipetting caused by blocked tips, mechanical defects of the pipetting channel or tips that do not conform to the instrument or to the pipetting requirements are not detected by the system. Pipetting without TADM is only to be used for specific types of pipetting such as blood group determination and must be checked visually.

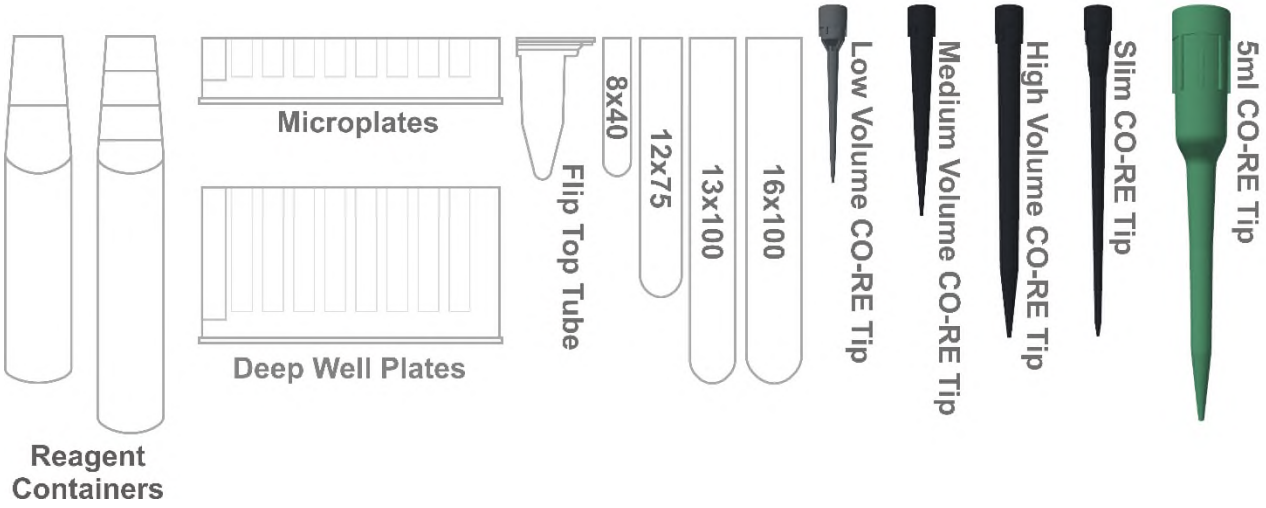
ATTENTION

BIOHAZARD

Always use filter tips for pipetting without TADM to prevent pipetting channels from being contaminated.

If sampling aggressive liquids, use filter tips. Also use filter tips for tasks which are sensitive to cross-contamination (aerosols).

The inner diameter of sample tubes, reagent vessels, etc. must be greater than the diameter of the utilized disposable tip at its maximum immersion depth. This measure will avoid any collisions between labware such as sample tubes, reagent vessels, etc., disposable tips and the pipetting channel. For additional information and proper tip selection, refer to Section 2.5 Tips and Needles.



ATTENTION

Liquid Level Detection needs to be explicitly tested when working with foamy liquids. Foam may affect the accuracy of liquid level detection.

- *The liquid must be conductive so that dual LLD can be utilized.*
- *The tolerance between the first and last allowable detection must be defined empirically.*
- *Then the immersion depth value has to be defined large enough to ensure that the tip is located below the foamy layer before aspiration. This requires some test runs as well. The starting point of the immersion depth is where both LLD's are triggered.*

1.10.1.9 Loading

Do not exchange positions of sample or reagent tubes or switch microplates after they have been identified by the barcode reader. This could result in incorrect test data or instrument crash.

Microplates must be placed on the carrier such that well A1 is in the position defined in the deck layout of the method.

When pouring liquid into the containers, ensure that there is no foam on the surface of the liquid. Note that foam may cause pipetting problems.

Do not overfill reagent containers, tubes or other liquid containers.

Do not mix tip size and type (e.g. with or without filter, or different volumes) in the same tip rack. Make sure to match the tip type to be used with the particular method. Take care when using tips

which cannot be distinguished by the tip recognition feature (refer to Section 2.1.7 Tip Recognition and Section 2.5 Tips and Needles).

Do not fill up partially consumed tip racks with tips from another rack. Tips must be loaded in the tip racks as they are provided in the original packaging. They are individually labeled with a barcode for identification.

1.10.1.10 Work Routine

Periodic maintenance (daily maintenance, weekly maintenance, six-monthly maintenance) is a mandatory part of the work routine and is guided by the user software.

ATTENTION

User access to the cabinets underneath the instruments during a run may interfere with automatic loading and unloading of carriers (the carriers protrude).

If the system is paused, do not wait too long before resuming the run. Loss of liquid from a full tip may result in invalid data.

Discard used tips and do not reuse them. Do not empty the tip waste during a run.

Do not try to open the front cover during a run. It may cause an abort of the run.

Do not leave tips on the pipetting channels for a long period of time (for example, overnight). This may cause damage to the CO-RE O-rings. A daily maintenance procedure will eject the tips.

1.10.1.11 Aseptic Applications

If used for aseptic processing, the ML STAR has to be installed in a grade A clean room with laminar airflow from the cleanroom ceiling according EU-GMP Annex 1. The directed airflow from the ceiling to the bottom ensures the air barrier function of machine housings especially designed for the protection of the open processed pharmaceutical product. During operation, moving machine parts will guide the airflow in a different pattern through the housing. Very important is that the preliminary air flow direction stays in operation during the complete processing in order to prevent contamination due to backflow of air into the instrument housing.

The STAR liquid handling robot system fulfils the criteria for a successful media fill simulation for aseptic processing defined in the currently valid EU-GMP guideline Annex 1.

The performance of the clean room regarding microbiological contamination fulfills the defined requirements of the EU-GMP guideline Annex 1 for grade A areas.

The STAR fulfills the requirements of air cleanliness for aseptic areas grade A defined in EU-GMP Annex 1 and ISO Class 4 according ISO 14644-1.

This means that the ML STAR can work aseptically in a sterile environment and does not present a risk of contamination for the environment or for the process.

1.10.2 Biohazard Precautions

ATTENTION

BIOHAZARD

If the ML STAR becomes contaminated with biohazardous or chemical materials, it shall be cleaned in accordance with the maintenance procedures (see Section 4 Maintenance and Section 6 ML STAR Instrument Decontamination). Observe and carry out the maintenance procedures given. Failure to do so may impair the reliability and correct functioning of the ML STAR.

If working with biohazardous samples, observe and carry out the maintenance procedures, paying particular attention to cleaning and decontamination. Wear gloves when handling the X-Arm and pipetting channels, the carriers, racks, containers and tips.

Avoid touching tips discarded into the laboratory-supplied waste container. Any surfaces on which liquid is spilled must be decontaminated.

Do not use disinfecting materials, which contain hypochlorite (Javel water, Chlorox) or other bleaching fluids.

If working with biohazardous or chemical materials, the user must not touch them. The ML STAR will drop its used tips into a Tip Waste Chute or Bag, which shall be emptied during the daily maintenance or as soon as it is full.

When using an exhaust suction device to remove toxic, aggressive or corrosive gases, make sure to place the device close to the source of gas or vapor in order to prevent contact of corrosives with ML STAR components, especially the pipetting heads.

1.10.3 Computer Precautions

Use the necessary precaution to guard against software viruses. Use only manufacturer's original installation 'data storage device' sets for the operating system. Use only the original HAMILTON VENUS Software supplied on a data storage device. Any manipulation of the VENUS Software data files or other information determining or affecting ML STAR functions can result in erroneous test results or instrument failure. Only the VENUS Software may be used to control the ML STAR.

For reasons of data safety and integrity, use of an Uninterruptible Power Supply (UPS) is recommended, since a loss of power may cause data to be lost or corrupted.

To avoid computer breakdowns, configure a hard disk of sufficient space in the computer. Ensure that there is always enough storage capacity on your hard drive. Delete log files from time to time. Generated data within the Log Files Directory, e.g. traces, TADM data and pipetting files, must be backed up onto the laboratory's host device and deleted from the control PC's hard disk at weekly intervals. Also see Section 1.10.4 End-User Computing Policy.

1.10.4 End-User Computing Policy

1.10.4.1 Scope

This policy applies to end-user computing devices that connect to a HAMILTON Automated System.

All customer employees, members, users, and third-parties utilizing computer devices connecting to a HAMILTON System, assume the responsibility for the security and privacy of information contained within.

1.10.4.2 Policy

End-User Computing Device Policy:

- With finalization of the technical installation of the HAMILTON system, the full responsibility and reliability for the computing device is transferred to the end-user of the system. That applies regardless of whether the PC was delivered by HAMILTON or by the end-user.
- Hence HAMILTON is not responsible for:
  - Administration of the PC, e.g. user rights, user levels, privacy.
  - Data safety, data integrity.
  - PC maintenance.
  - Appropriate security measures, e.g. against threads via the network / internet.
  - Data backup and recovery.
  - Disaster Recovery
  - Operating System updates and security patches.
  - Cyber security
  - Computer security
- If HAMILTON delivers a PC as part of the instrument, a Warranty Agreement will be included by the PC manufacturer. That warranty includes hardware only and excludes explicitly any support for the topics mentioned above.

1.10.5 Mechanical or Electrical Safety Precautions

Before removing a mechanical or an electrical component, the ML STAR must be switched off and disconnected from the main electricity supply and from the PC.

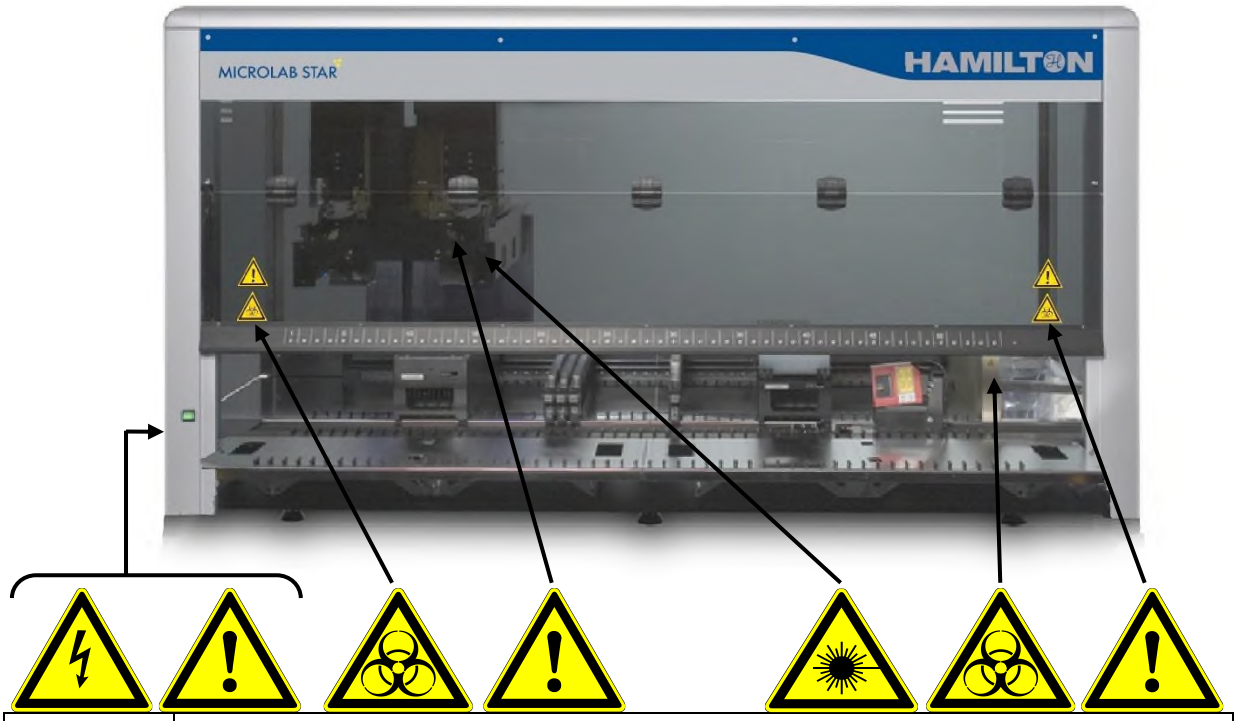
Any installation and de-installation of an electrically powered ML STAR component must be performed by a HAMILTON Trained Field Service Engineer.

1.10.6 Hazards

The safety of personnel and equipment can only be ensured if the safety instructions and the safety-related warnings are strictly observed and followed.

The following section describes the main safety considerations in operating your HAMILTON product and the main hazards involved.

Location and explanation of warning and attention labels:



	**Moving Parts** Be aware of moving parts (arm, pipetting Heads, Gripper, etc.) inside transparent cover. *During instrument operation, stand clear of all moving parts and the working deck of the instrument. Never lean over or interfere with the instrument while it is in operation! Never reach into a running instrument with your hand(s)!* *If not followed as mentioned above, there can be injuries or contamination!* *The run will be aborted if the front cover is opened.*
	**ATTENTION** **BIOHAZARD** Deck may contain biohazardous or chemically contaminated materials.

	**Power Connection** Connect only to earth-grounded outlet.		**ATTENTION** **BIOHAZARD** Waste may contain biohazardous or chemically contaminated materials**.**
	**Connection to PC** Use only the appropriate shielded cables.		**Moving Parts** Moving arm inside transparent cover. Aborts the run if the cover is opened.
	**USB Connection** Having a total cable distance of more than 5m, signals loss can occur.		**ATTENTION** **HOT SURFACE** Avoid contact with the HHS. Surfaces are hot and may cause personal injury if touched.
	**ATTENTION** **BIOHAZARD** Deck may contain biohazardous or chemically contaminated materials.		**Pipetting Arm** Do not move Pipetting Arm by hand.
	**ATTENTION** **LASER BEAM (Tube Twister)** Do not stare into beam of class 2 laser of the barcode reader.		
Laser device designated for use solely as a component of the overall automated pipetting instrument and therefore is not within the scope of FDA Section 1040.11 for complete laser products. Laser devices are only to be installed, repaired, or replaced by a HAMILTON Trained Field Service Engineer.			

1.10.6.1 Substance Related Hazards

ATTENTION

BIOHAZARD

Chemical, biological and radioactive hazards can be associated with the substances used or the samples processed with the instrument. The same applies to waste disposal. The handling of substances and the disposal of waste may be subject to local, state or federal law or regulations with regard to health, environment or safety.

- *Always be aware of possible hazards associated with these substances*
- *Use appropriate protective clothing, goggles and gloves*
- *Strictly observe the corresponding provisions*

ATTENTION

Caustic substances can cause burns and eye injury.

- *Always be aware of possible hazards associated with these substances*
- *Avoid exposure to caustic substances*
- *Use appropriate protective clothing, goggles and gloves*

ATTENTION

FIRE HAZARD

Use caution when using flammable or explosive fluids or vapors. There is a high risk of damage to health and to the instrument. The instrument is not explosion-proof nor for use in Ex zones.

1.10.6.2 Safety Measures

- **Access to Power Switch/Power Cord**

  Ensure there is enough room to access the power switch and power cord.

- **Working Area**

  The safety concept assumes that the doors are always closed and locked during normal operation.

- **Surrounding Area**

  Make sure the ventilation outlets of the instrument are not impaired by obstacles placed in the surrounding area.

ATTENTION

Do not use mobile phone or similar devices less than 2 meters from the instrument while the instrument is in operation. This could cause a possible malfunction of the instrument.

ATTENTION

Rapid temperature changes or direct sunlight may affect certain functions such as barcode reading and pipetting accuracy.

- *Do not open windows next to the instrument*
- *Do not expose the instrument to direct sunlight or intense artificial light during operation*

Use appropriate:

Protective clothing

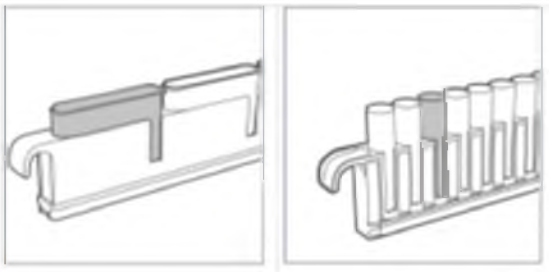
Goggles

and Gloves.

1.10.6.3 Loading Reagents and Samples

Loading Reagents and Samples:

- The 1T Reagent Racks and 1T Sample Racks can tip over during sample and reagent loading and unloading



ATTENTION

Secure 1T Reagent Racks and 1T Sample Racks to prevent the risk of spillage of reagents or samples.

1.10.6.4 Loading Tray

Loading Tray:

- The instrument loading tray is a part of the instrument and must be treated with care

ATTENTION

Risk of damage to equipment. Leaning onto the instrument loading tray will lead to damage of the loading tray.

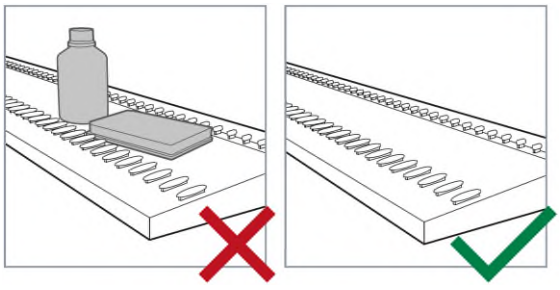
- *Do not lean onto the loading tray.*

ATTENTION

If you are making manipulation below the instrument (e.g. to store or retrieve consumables from under the workbench), when lifting, take care not to hit your head on the Loading Tray.

Foreign object damage:

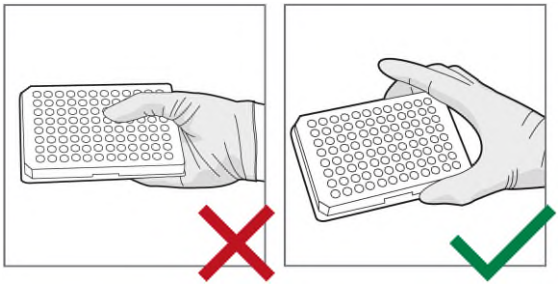
- Make sure that the loading tray has been cleared from foreign objects before running the instrument
- Foreign objects may obstruct the instrument and lead to damage to the equipment



1.10.6.5 Holding Labware

Holding Labware:

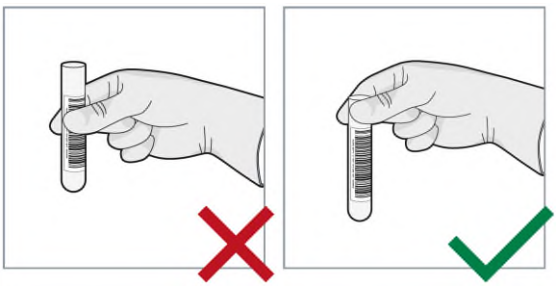
- Hold microplates by its sides



1.10.6.6 Holding Tubes

Holding Tubes:

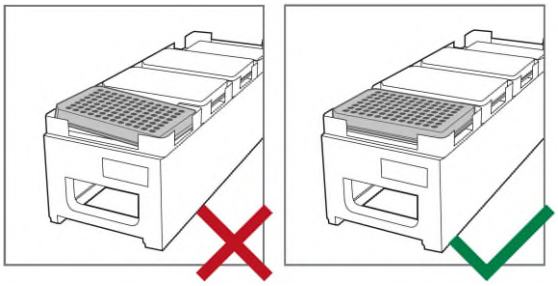
- Avoid holding tubes on barcodes
- Hold the tubes as shown



1.10.6.7 Placing Labware

Placing labware onto carriers:

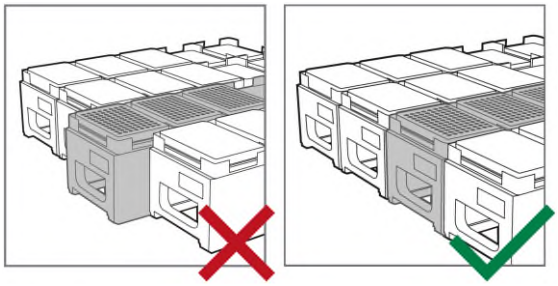
- Make sure labware is placed onto the carrier correctly
- Incorrectly placed labware will lead to collision or malfunction



1.10.6.8 Loading Plate Carriers

Loading Plate Carriers:

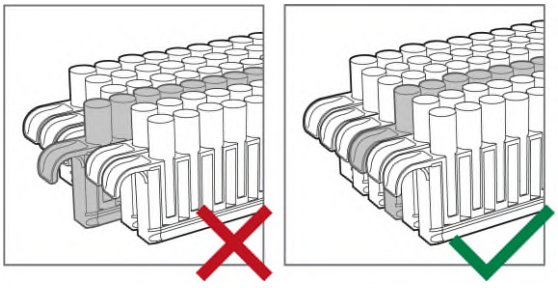
- Make sure the carriers have been pushed to the mechanical stop of the instrument
- Incorrectly positioned carriers will lead to malfunctions or collision



1.10.6.9 Loading Tube Carriers

Loading Tube Carriers:

- Make sure the tube carriers have been pushed to the mechanical stop of the instrument.
- Incorrectly positioned tube carriers will lead to malfunctions or collision.

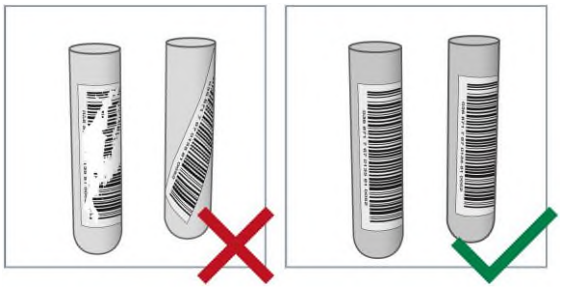


1.10.6.10 Barcodes Placement

Barcode quality:

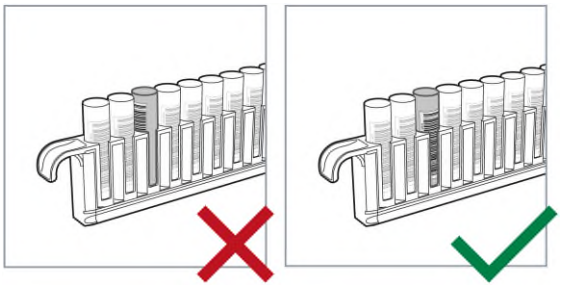
- Make sure that all barcodes are readable as well as correctly applied
- Unreadable, scratched, smudged or incorrectly positioned barcodes on tubes will not be processed

Refer to the barcode types and quality requirements in Section 7.27 Autoload Option: Barcode and Reader Specifications.



Barcode orientation:

- Make sure all tube barcodes have the correct orientation
- Incorrectly placed tube barcodes will not be readable

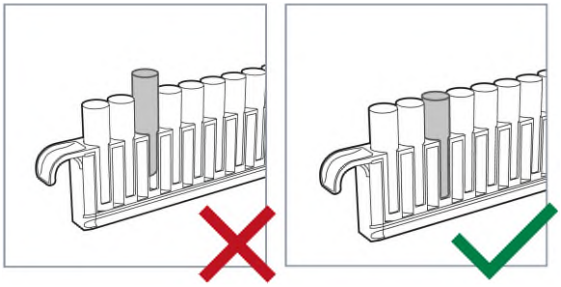


1.10.6.11 Tube Placement

Tube position:

When placing tubes into tube carriers:

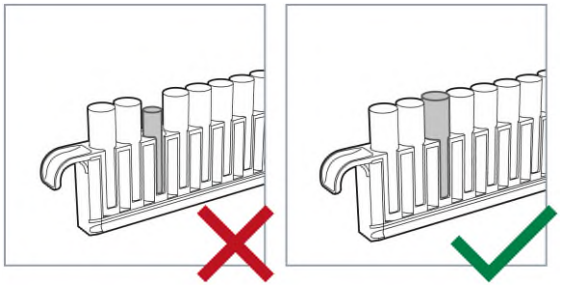
- Make sure the tubes are pushed all the way down into the tube carrier
- Incorrectly placed tubes may lead to malfunctions



Tube height:

When placing tubes into tube carriers:

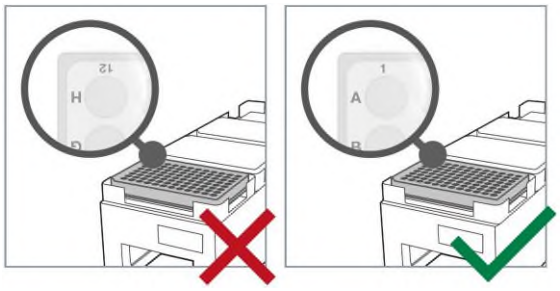
- Make sure that all tubes placed into the tube carrier are the same height
- Different tube heights will lead to incorrect calculation of sample volumes and lead to malfunctions



1.10.6.12 Labware Orientation

Labware orientation on carriers:

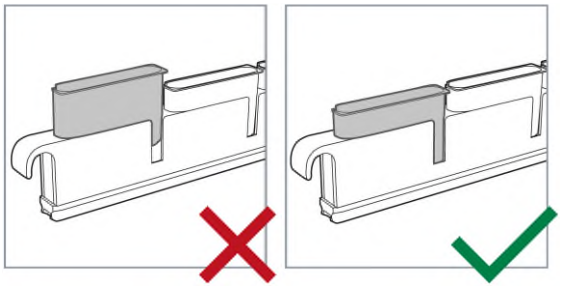
- Make sure plates have the correct orientation on the carrier
- The well position A1 has to be in the upper left



1.10.6.13 Trough Position

Placing troughs into trough carriers:

- Make sure the troughs are pushed all the way down into the carrier
- Incorrectly placed troughs will lead to spillage and/or malfunctions



1.10.7 Reporting Serious Incidents

Any serious incident or near incident that has occurred in relation to the ML STAR shall be reported to HAMILTON. Contacting HAMILTON either by contacting a local HAMILTON representative or by utilizing HAMILTON's webpage, which can be found on the last page of this Operators Manual.

2. Description of the ML STAR

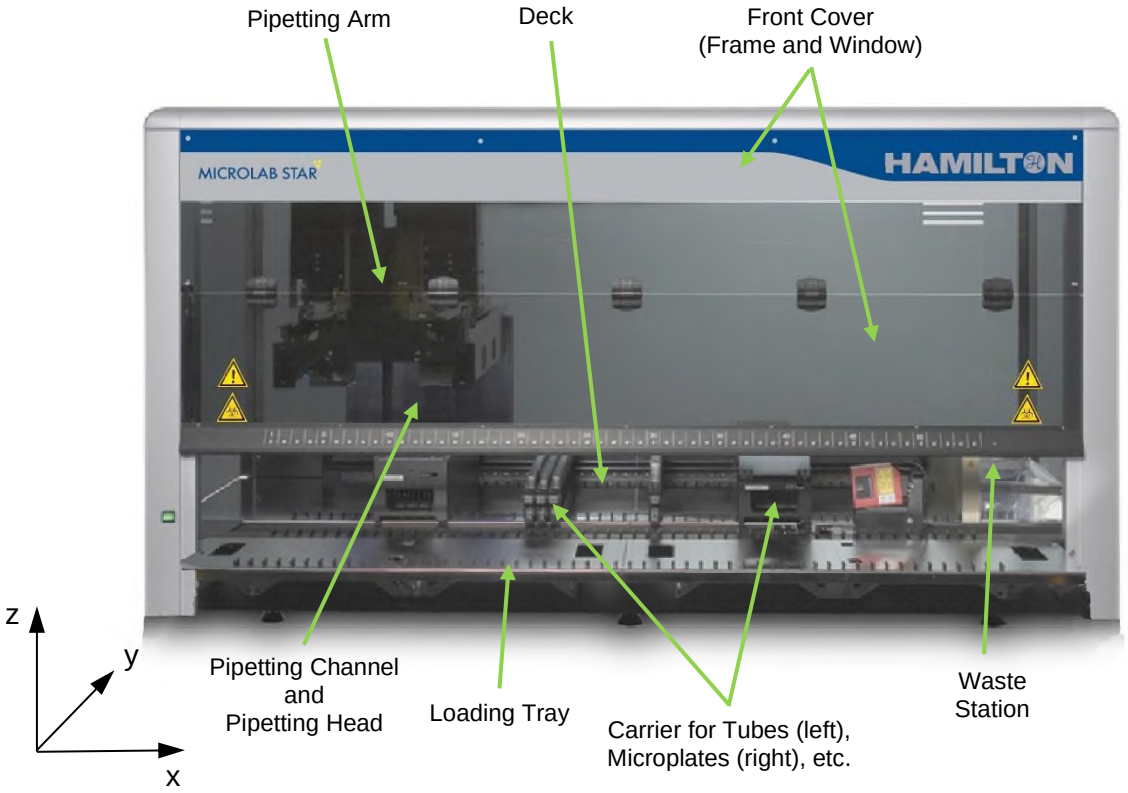
The ML STAR is a robotic pipetting instrument used for pipetting liquid samples in automated processes.

Pipetting means transfer of small quantities of liquid from one container to another. A pipetting operation is achieved by aspirating (drawing) liquid from a source container, then transferring and dispensing (dropping) it into a target container.

The Components and Modules described in this section are either factory installed and / or installed by a HAMILTON Trained Field Service Engineer.

2.1 Platforms

The ML STAR instrument work surface, called a "deck", for placing loadable carriers is available in three different sizes. These carriers hold reagent containers, such as tubes, microplates and other kinds of labware.



The ML STAR

The instrument deck is divided into equal tracks (T) for loading carriers in pre-determined positions. This eliminates the need for precise measurement of positions. The deck has partitions of 22.5 mm, which is equivalent to 1-T (track). The labware carriers are adapted to these partitions (e.g. 1-T carriers for sample tubes or 6-T carriers for microplates or CO-RE tips, etc). An additional partition provides space for the tip waste container.

The instrument is fully covered by acrylic glass covers. The front cover consists of a hinged transparent window made of acrylic glass. This window is equipped with a magnetic switch that is monitored during a run. Opening the cover will abort the running method.

A Touch Screen Monitor is available and can be attached to the right side of the ML STAR instrument. It is the perfect interface to start methods, enter data and to make decisions during runtime. For ergonomic reasons, it is recommended to use a desktop computer with monitor, keyboard and mouse for extended programming work.

The compact version of the ML STAR family is called MICROLAB STARlet. The ML STARlet is the version of instrument with the smallest deck divided into 30 equal tracks (T) for loading carriers.

For detailed information, see Section 7.1 Basic ML STAR Specifications.

ML STARlet

The deck of the MICROLAB STAR is divided into 54 equal tracks (T) for loading carriers.

For detailed information, see Section 7.1 Basic ML STAR Specifications.

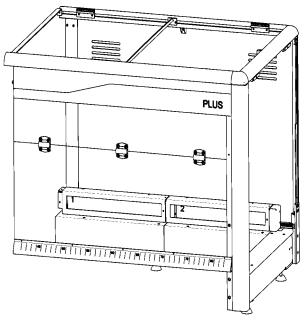
ML STAR

The large version of the ML STAR is called MICROLAB STARplus. If equipped with a full deck extension, the ML STARplus has a deck divided into 71 equal tracks for loading carriers.

For detailed information, see Section 7.1 Basic ML STAR Specifications.

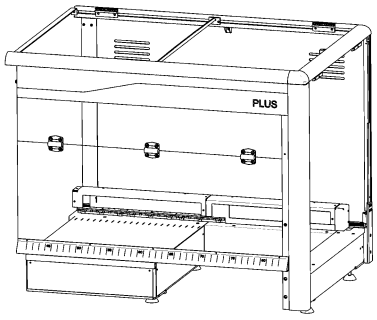
ML STARplus

The ML STARplus is the compact ML STARlet extended on site by additional workspace. The workspace can be filled by additional tracks. The other possibility is to integrate 3rd party devices, such as readers, washers, centrifuges, etc. The extension part of the ML STARplus is available in 3 versions:



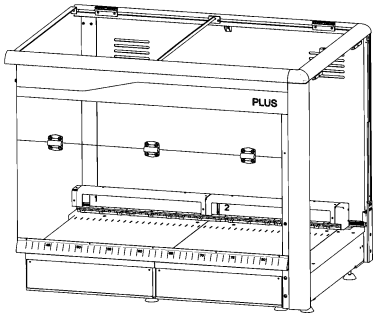
Without deck extension:

- *ML STARplus has 30 tracks*
- *Total of 25 SBS positions*



With left half deck extension:

- *ML STARplus has 50 tracks*
- *Total of 40 SBS positions*

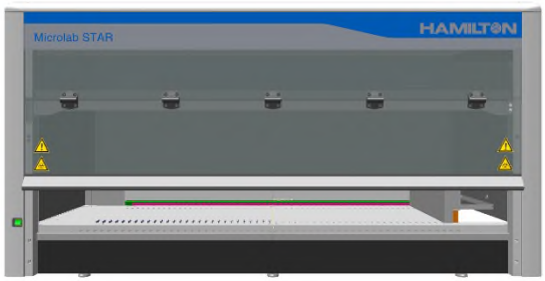


With complete deck extension:

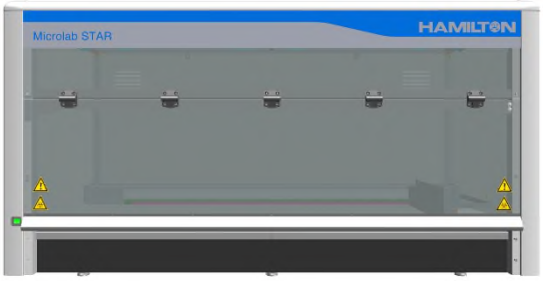
- *ML STARplus has 71 tracks*
- *Total of 55 SBS positions*

2.1.1 ML STAR Manual Load

The ML STAR Manual Load allows loading of racks and carriers directly onto the deck. The ML STAR Manual Load has the option for a standard front cover or full front cover.



ML STAR Manual Load with Standard Cover



ML STAR Manual Load with Full Front Cover

2.1.2 Autoload Option

The Autoload enables the automatic loading of carriers onto the ML STAR IVD-R Instrument deck:

- Moves in the x-direction
- Shunts carriers on and off the deck
- Reads barcodes of carriers, tubes, and microplates

NOTE

The Barcode Scanner on the Autoload of the ML STAR IVD-R has a Class I Laser Diode.

Barcode Scanner

NOTE

Do not put your fingers or hand between the Barcode Reader and Sledge, as there is the possibility of bruising.

The following Carrier and Labware can be handled by the Autoload:

- Carrier Types:
  Tip Carrier, Plate Carrier, Sample Tube Carrier, Reagent Carrier, Stacker Carrier. See Section 8.1 Appendix A: Ordering Information for information regarding carrier types.
- Sample Tubes:
  See Section 8.1 Appendix A: Ordering Information for information regarding carrier types holding sample tubes.
- Reagent Containers:
  See Section 8.1 Appendix A: Ordering Information for information regarding carrier types holding reagent containers.
- CO-RE Tips:
  See Section 8.1 Appendix A: Ordering Information for information regarding carrier types holding CO-RE Tips.

There is a presence sensor that identifies if tubes are present on a sample carrier.

Carrier identification by barcodes, and reading of barcodes on plates and tubes, is only possible in combination with the Autoload.

The following barcode symbologies can be read by the Autoload:

ISBT Standard	Code 128 (Subset B and C)
Code 39	Codabar
Code 2 of 5 interleaved	UPC A/E
JAN/EAN 8	

Barcodes must be black bars on a white background. HAMILTON recommends using the barcode type Code 128 (Subset B and C).

NOTE

In addition, barcodes must have a minimum readability (i.e., good contrast, size, correct orientation and distance between bars) to be fully functional. Ensure the correct barcode orientation for tubes and plates (see specifications).

For details of barcodes, see the specifications given in Section 7.27 Autoload Option: Barcodes and Reader Specifications.

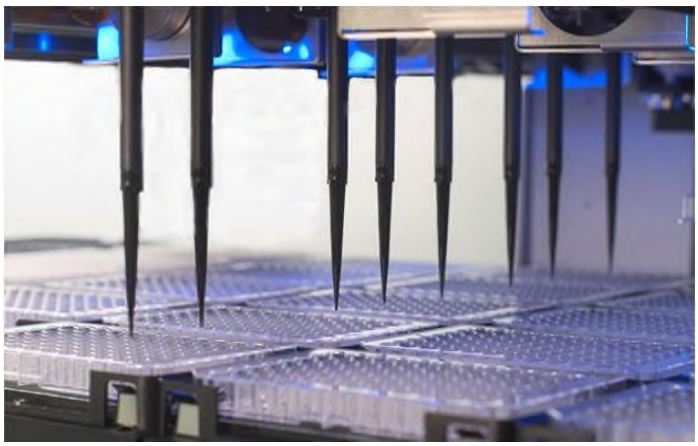
2.1.3 Pipetting Channels

The ML STAR can be equipped with up to 16 1000 µl or up to 8 5 ml pipetting channels, or with a mix of both. The total number of Pipetting Channels in a mixed configuration is Pipetting Arm dependent.

2.1.3.1 1000 µl Pipetting Channels

The ML STAR is available with a maximum of 16 1000 µl pipetting channels working in parallel for simultaneous transfer of liquids. Each 1000 µl pipetting channel moves independently on the Y-Axis, as well as on the Z-Axis. Each 1000 µl pipetting channel uses its own high-precision motors and electronics to reach any position on the deck without the need of teaching.

The 1000 µl pipetting channels with a nominal volume of 1000 µl, support pipetting with disposable CO-RE tips or with needles.



The minimum distance between 2 1000 µl pipetting channels on the Modular X-Arm is 9 mm (for Thin Arm 18 mm).

The 1000 µl pipetting channels have a set 'traverse height' of 245 mm above the origin or 145 mm between the top of the disposable tip and the deck of the instrument. This means, when a pipetting channel is to move from one location on the deck to another, it automatically does so at that particular height. This is a safety precaution, so that pipetting channels will not collide with any items that may be on the deck.

An instrument equipped with 8 1000 µl pipetting channels has full 'random access space', which is the area where any of the pipetting channels is able to reach.

The 1000 µl pipetting channels support pipetting with disposable tips or with needles.

The following table gives an overview of disposable tips which can be used with the 1000 µl pipetting channels. Other combinations may be possible, but require special attention.

NOTE

If a type of tip is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

1000 µl Pipetting Channel	Tips for Working Volume and Purpose					
Plate Type	10 µl CO-RE Tip	50 µl CO-RE Tip	300 µl CO-RE Tip	1000 µl CO-RE Tip	250 µl Robotic Piercing Tip	300 µl Slim Tip
 96-Well						
 96-Well Archive						
 384-Well Archive						
 1536-Well						

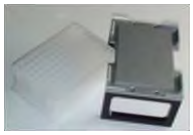
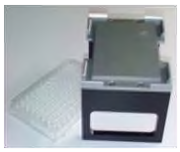
Instruments with 4, 8, or 16 1000 µl pipetting channels are best operated with carriers holding microplates and tip racks in landscape orientation, whereas carriers with portrait orientation for microplates and tips are best suited for an instrument with 12 1000 µl pipetting channels.

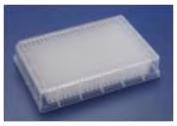
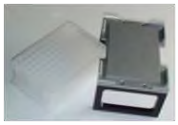
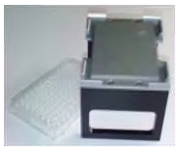
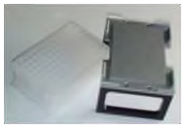
The following table gives an overview of carriers and modules which can be used with the 1000 µl pipetting channels. Other combinations may be possible, but require special attention.

NOTE

If a type of carrier or module is not shown, it cannot be used for that purpose.

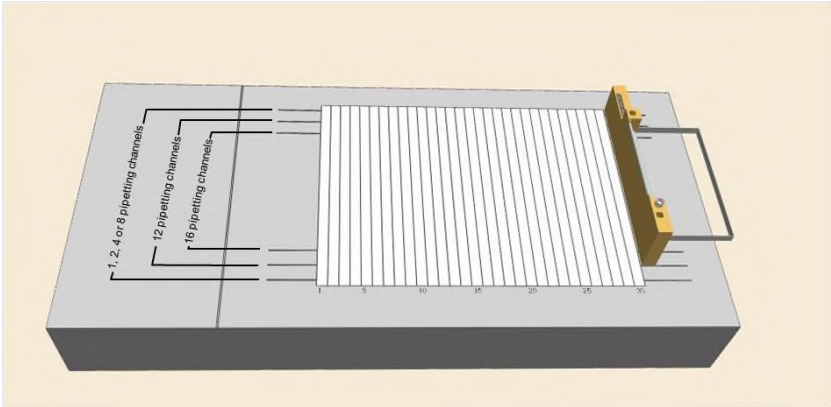
See Section 8.1 Appendix A: Ordering Information for a complete listing.

1000 µl Pipetting Channel	Carriers and Modules			
 96-Well	 Plate Carrier for 96/384 Well Plates or  Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	 Multiflex MTP Module or  Multiflex DWP Module
 96-Well Archive	 Plate carrier for Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested		 Multiflex DWP Module
 384-Well	 Plate Carrier for 96/384 Well Plates or  Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	 Multiflex MTP Module or  Multiflex DWP Module

1000 µl Pipetting Channel	Carriers and Modules			
 384-Well Archive	 Plate Carrier for Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested		
 1536-Well		 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	

An instrument equipped with 1 to 8 1000 µl pipetting channels on the modular arm has the full "random access space", which is the area where any of the 1000 µl pipetting channels are able to reach. For a 12 or 16 1000 µl pipetting channel equipped instrument, the "random access space", is reduced.

As shown in the screenshot below, the "random access space" of the different numbers of 1000 µl pipetting channels is indicated by lines at the left and right side of the deck layout of the VENUS Software. To guarantee random access to sample tube carriers, only the inner tube positions shall be used.



NOTE

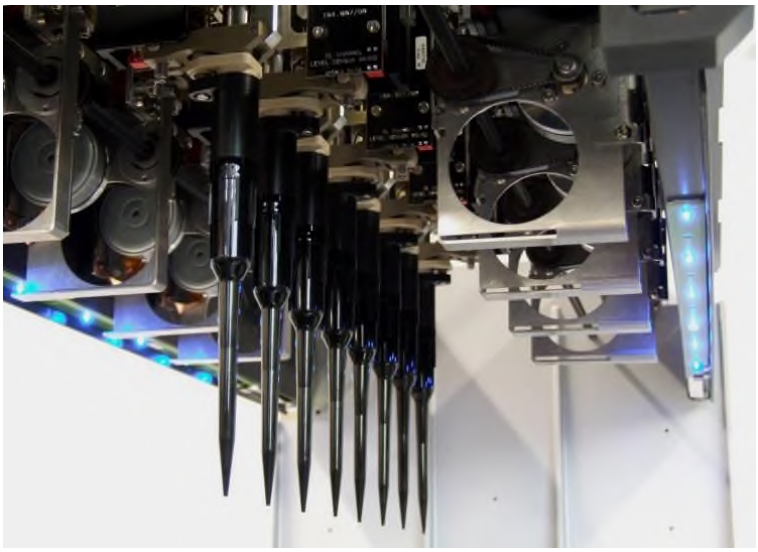
If pipetting positions outside the access space of the instrument are used, the system reports an error. However, a strictly batch-type process can eliminate these problems.

Low-volume tips do not reach the deck, even at the lowest z-position of the pipetting channel.

2.1.3.2 5 ml Pipetting Channels

The ML STAR is available with a maximum of 8 5 ml pipetting channels working in parallel for simultaneous transfer of liquids. Each 5 ml pipetting channel moves independently on the Y-Axis, as well as on the Z-Axis. Each 5 ml pipetting channel uses its own high-precision motors and electronics to reach any position on the deck without the need of teaching.

The 5 ml pipetting channels with a nominal volume of 5 ml, support pipetting with disposable CO-RE tips.



The minimum distance between 2 5 ml pipetting channels on the Modular X-Arm is 18 mm (for Thin Arm 36 mm).

The 5 ml pipetting channels have a set 'traverse height' of 245 mm above the origin, or 145 mm between the top of the disposable tip and the deck of the instrument. This means, when a channel is to move from one location on the deck to another, it automatically does so at that particular height. This is a safety precaution, so that channels will not collide with any items that may be on the deck.

The 5 ml pipetting channels support pipetting with disposable tips.

The following table gives an overview of disposable tips which can be used with the 5 ml pipetting channels. Other combinations may be possible, but require special attention.

NOTE

If a type of tip is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

5 ml Pipetting Channel	Tips for Working Volume and Purpose	
Plate Type	5 ml Tip with Filter	5 ml Tip without Filter
 96-Well	 CO-RE Tip with Filter, 4000 µl	 CO-RE Tip without Filter, 5000 µl
 96-Well Archive	 CO-RE Tip with Filter, 4000 µl	 CO-RE Tip without Filter, 5000 µl

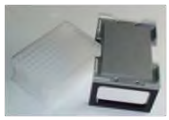
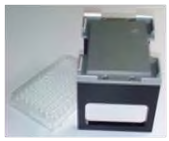
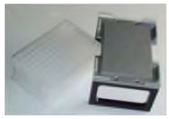
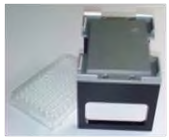
Instruments with 4 or 8 5 ml pipetting channels are best operated with carriers holding microplates and tip racks in landscape orientation, whereas carriers with portrait orientation for micro-plates and tips are best suited for an instrument with 6 5 ml pipetting channels.

The following table gives an overview of carriers and modules which can be used with the 5 ml pipetting channels. Other combinations may be possible, but require special attention.

NOTE

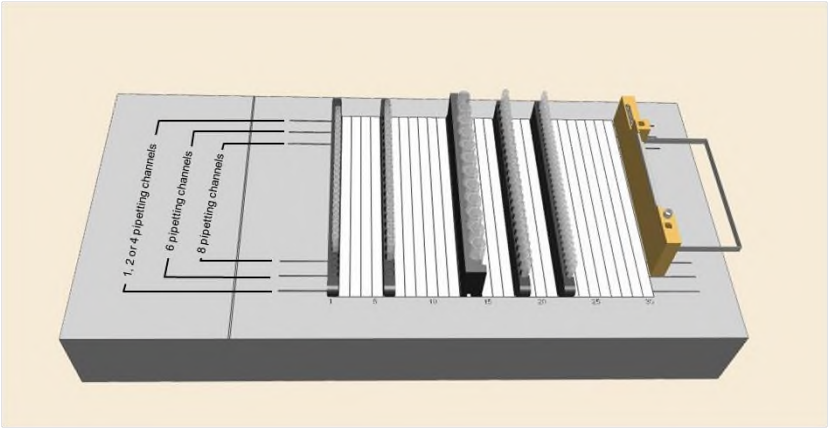
If a type of carrier or module is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

5 ml Pipetting Channel	Carriers and Modules			
 96-Well	 Plate Carrier for 96/384 Well Plates or  Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	 Multiflex MTP Module or  Multiflex DWP Module
 96-Well Archive	 Plate Carrier for Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested		 Multiflex DWP Module

An instrument equipped with 1 to 4 5 ml pipetting channels on a modular arm has the largest "random access space" which is the area, where any of the 5 ml pipetting channels is able to reach. For a 6 or 8 5 ml pipetting channel equipped instrument, the "random access space" is reduced.

The "random access space" of the different numbers of 5 ml pipetting channels is indicated by lines at the left and right side of the deck layout of the VENUS Software, as shown in the screenshot below. To guarantee random access to sample tube carriers, only the inner tube positions shall be used.

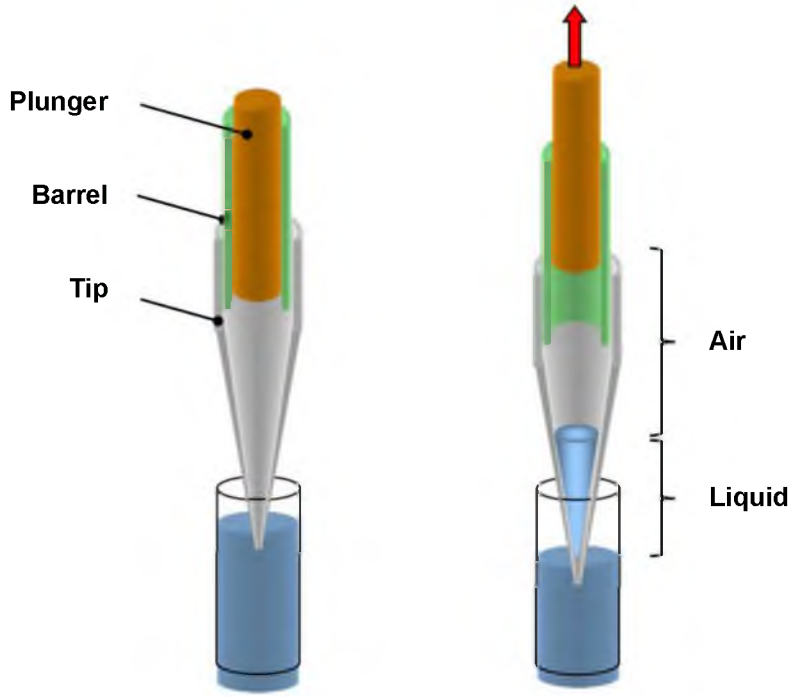


NOTE

If pipetting positions outside the access space of the instrument are used, the system reports an error. However, a strictly batch-type process can eliminate these problems.

2.1.4 The Air Displacement Pipetting Principle

The ML STAR Line instruments are based on the **air displacement pipetting** principle, comparable to the functionality of handheld pipettes. Air displacement means that the liquid is aspirated into and dispensed from a disposable tip or needle by the movement of a plunger. Between the plunger and the liquid surface is air. No system liquid of any kind is involved in the ML STAR.



The air displacement pipetting principle

NOTE

Pipetting with the ML STAR is identical to pipetting with a handheld plunger pipette. The pipetting head´s barrel and plunger are not intended to be cleaned. We recommend using filter tips when working with infectious agents such as viruses, etc.

2.1.5 Liquid Level Detection: LLD

HAMILTON's single pipetting channels (1000 µl and 5 ml) offer two modes of Liquid Level Detection (LLD):

Capacitance- and pressure-based. The capacitive LLD (cLLD) detects conductive liquids. The pressure-based LLD (pLLD) detects all liquids including non-conductive solvents. If required for the application, capacitance and pressure based LLD may be combined as Dual LLD (e.g. foamy samples).

The liquid level of the container to be aspirated from can be detected. Normally, capacitive LLD is used for conductive liquids. The sensitivity of the capacitive LLD that is to be used depends on the vessel size, volume and the conductivity (or polarity) of the liquid to be detected.

For non-conductive liquids, or in case of an insufficient coupling between container bottom and carrier, pressure LLD is used. Pressure LLD only works with new and empty tips for the aspiration of liquids. The pLLD is available on the individual pipetting channels only.

In case of detecting under demanding circumstances (e.g. foaming liquids), the capacitive and the pressure Liquid Level Detection can be used at the same time.

NOTE

We recommend always using a defined sensitivity, such as 'low', 'medium', 'high' or 'very high'. If the value in the labware is 'Off' it causes bad results.

When selecting 'very high' all of the time, liquid may be detected in a wrong position, cLLD can, for example, be triggered by vapor of volatile liquids or by foam.

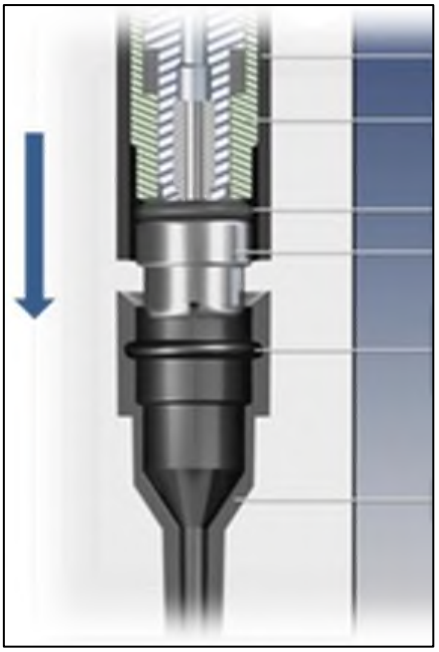
ATTENTION

Liquid Level Detection needs to be explicitly tested when working with foamy liquids. Foam may affect the accuracy of liquid level detection.

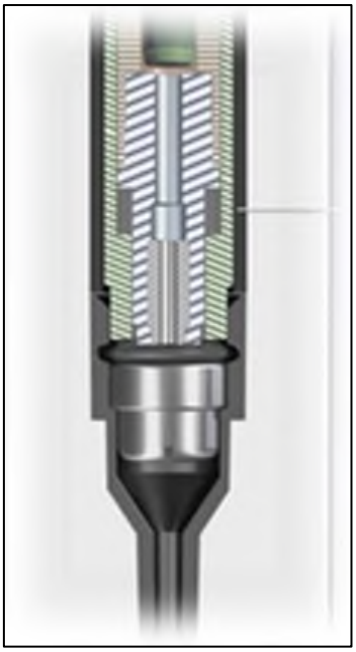
- *The liquid must be conductive so that dual LLD can be utilized.*
- *The tolerance between the first and last allowable detection must be defined empirically.*
- *Then the immersion depth value has to be defined large enough to ensure that the tip is located below the foamy layer before aspiration. This requires some test runs as well. The starting point of the immersion depth is where both LLD's are triggered.*

2.1.6 Tip Pick-up with the CO-RE Technology

The first task for the ML STAR pipetting head is to pick up a HAMILTON disposable tip or a reusable steel needle. Due to the unique CO-RE (Compression-induced O-Ring Expansion) technology, precise tip attachment and positioning is achieved. The system requires no vertical force for tip attachment or tip ejection, thus eliminating mechanical stress and improving overall system reliability along with pipetting speed and capability.



1. Tip Ejection Sleeve
2. Pressure Sleeve
3. O-Ring (unsqueezed)
4. Stop Disk
5. Groove for the O-Ring Expansion
6. Disposable Tip



NOTE

The principle of the CO-RE technology has the following advantages:

- *Enables coupling of disposable tips or washable needles within the same run*
- *Allows use of different sizes of tips on the same pipetting head in the same run*
- *Picking up a gripper and other tools is possible with a pipetting channel*
- *Prevents aerosols upon tip ejection*

ATTENTION

BIOHAZARD

Only HAMILTON CO-RE tips and needles shall be used for coupling to the pipetting heads of the ML STAR. Other tips may cause contaminated or damaged pipetting channels and may lead to erroneous results.

CO-RE technology is used on all HAMILTON pipetting channels, including CO-RE 96 and 384 Probe Head.

2.1.7 Tip Recognition

The tips used in a pipetting procedure must be matched with the suitable pipetting channel or pipetting head in order to prevent damage to the device. Therefore the ML STAR instruments offer two ways to automatically recognize the tip type used. All tip racks have color-coded barcode labels for automated recognition. The color helps distinguishing the tip volume by eye. The text on the barcode label enables a distinct identification by eye.

Tip volume	Barcode label	Color code
10 µl		light orange
50 µl		light red
300 µl		yellow
1000 µl		white
5000 µl		green

2.1.7.1 Tip Recognition using the Autoload Function

The Autoload (see Section 2.1.2 Autoload Option) includes a barcode reader which identifies the barcodes on the tip racks. Barcodes of tip racks are read automatically when loading tips and the risk of mixing up tip types is excluded. This method is suitable for all tip types, regardless of the pipetting tools to be used.

Tip carrier with five bar-coded tip racks loaded

2.1.7.2 Tip Recognition using the VENUS Tip Type Recognition Feature

The VENUS Software offers a tip type recognition feature for 1000 µl pipetting channels, based on the different tip geometries and independent from barcode (please refer to the VENUS Software Programmer´s Manual). It is available for both, disposable tips and needles and it is activated during installation by the HAMILTON Trained Field Service Engineer. This feature increases the instrument´s safety when no Autoload is used and when different tip types are used.

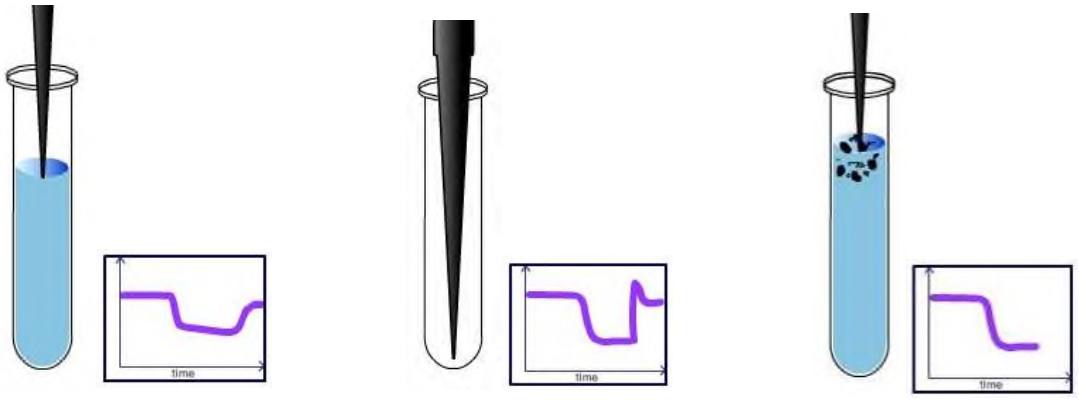
For distinguishing disposable CO-RE tips, 50 µl and CO-RE tips, 300 µl, a special library is needed. Please consult a local HAMILTON representative.

Slim tips 300 µl cannot be distinguished from CO-RE tips 1000 µl by the tip type recognition feature. We recommend using the Autoload for tip loading in this case and in any case to implement a visual check to be sure.

2.1.8 Monitored Air Displacement: MAD

The ML STAR is equipped with an aspiration monitoring feature. During the aspiration process, the pressure within the pipetting channel is measured in real time. Analyzing the shape of the p(t) curve, the system can distinguish the following situations:

- A correct aspiration takes place
- Air is aspirated into the tip (because, for example, the container has not been filled properly)
- A clot blocks the tip



Normal **Empty** **Clogged**

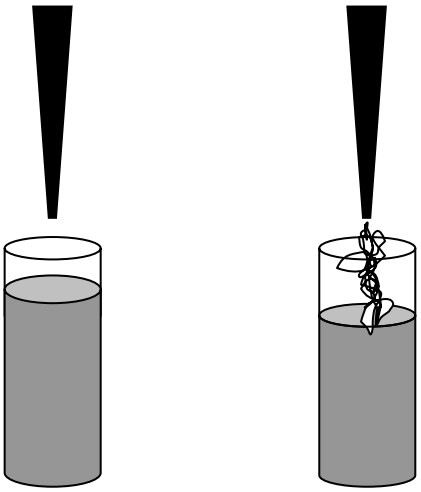
Aspiration monitoring based on pressure.

The MAD function (Monitored Air Displacement) is available on individual pipetting channels only.

The diagram shows the functioning of Monitored Air Displacement based on pressure.

2.1.9 Capacitance-Based Clot Detection

In addition to pressure-based clot detection, the ML STAR is equipped with capacitance-based clot detection. This detection approach works in the case of aspiration with capacitance Liquid Level Detection switched on. The system measures the conductive signal when the tip leaves the liquid after aspiration. Due to the air gap between the tip and the liquid, the capacitance signal will vanish once a given height is reached. If a clot is present, it bridges the distance and the signal will remain, resulting in an error message. This clot detection is independent of pressure-based monitoring.



2.1.10 Anti-Droplet Control: ADC

Does the application require pipetting of volatile solvents? The Anti-Droplet Control (ADC) enables doing this with highest process safety. In conventional pipettes the high evaporation pressure of volatile solvents causes immediate dripping from the tip. The ML STAR uses Monitored Air Displacement Technology and can therefore detect pressure changes following aspiration and compensate for them in real time.

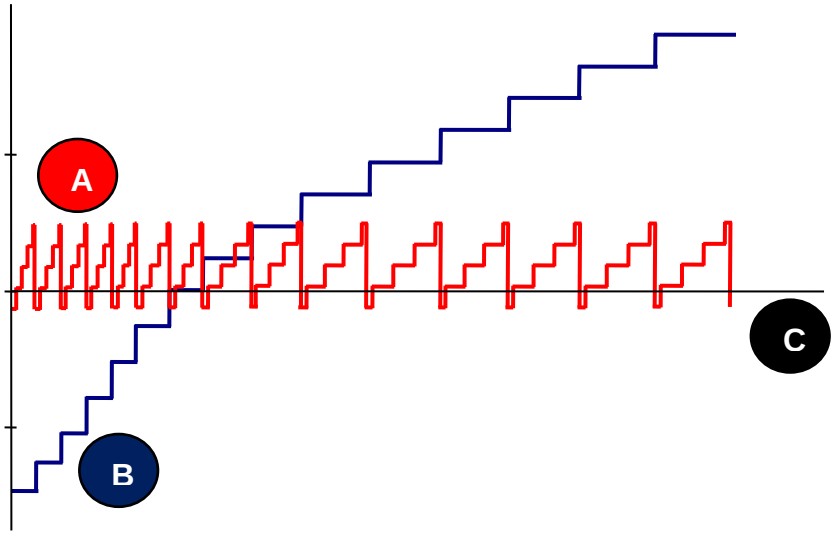
The principle is shown in the illustration on the next page: as the evaporation causes a pressure increase (red) the pipetting unit detects the changes and compensates for them with plunger movements (blue). The liquid remains in the tip.

The Anti-Droplet Control (ADC) has to be activated in the 'HSLML_STARLib' command library of the VENUS Software. It is available on the individual pipetting channels only.

ATTENTION

Use ADC only when pipetting liquids that ask for this mode.

There are two reasons that make a continuous use of ADC not practical. First, one might run into a position in which the plunger is no more able to handle the upcoming pressure difference and will therefore trigger an error. The second reason is that there might be a pressure interference which cannot be compensated by the plunger while the instrument is not pipetting.

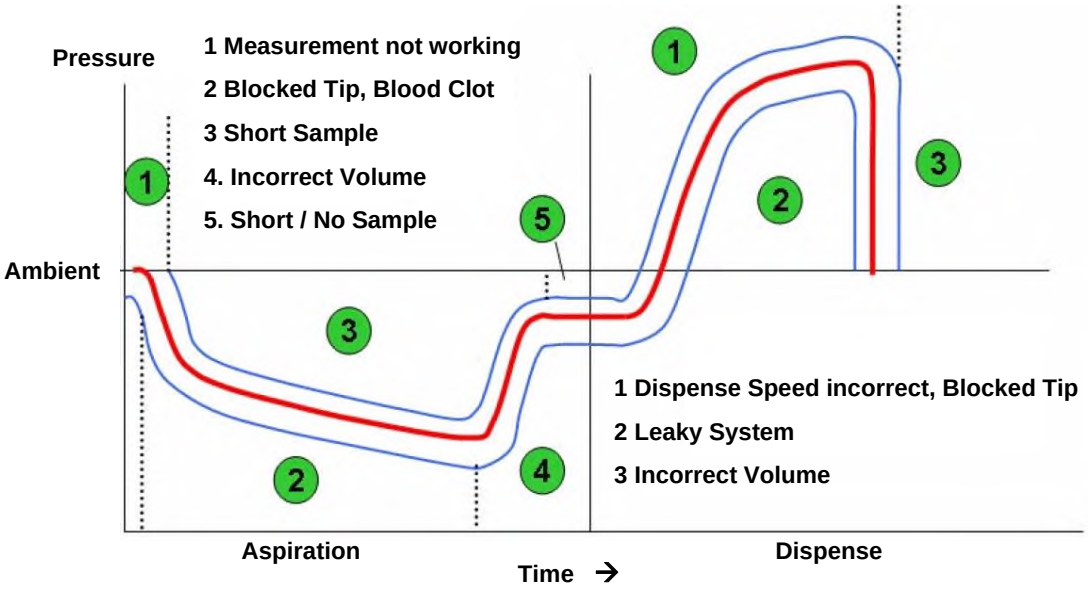


A. Pressure (red) **B. Plunger movement (blue)** **C. Time**

2.1.11 Total Aspiration and Dispense Monitoring: TADM

During crucial sample transfers, the ML STAR can monitor aspiration and dispense steps in real time. TADM verifies that a sample has been transferred by using a traceable digital audit trail. The function of TADM is a software option and therefore not available by default.

TADM is available for individual pipetting channels and with the CO-RE 96 Probe Head 1000 µl TADM.



2.1.12 Availability of Technologies on the Pipetting Tools

The following table provides an overview of individual technologies available with the various pipetting tools.

Technology / Pipetting Tool	CO-RE	cLLD	pLLD[1)]	MAD	TADM	ADC
1000 µl Pipetting Channel	✓	✓	✓	✓	✓[2)]	✓
5 ml Pipetting Channel	✓	✓	✓	✓	✓[2)]	✓
CO-RE 96 Probe Head	✓	✓[3)]	-	-	-	-
CO-RE 96 Probe Head TADM	✓	✓[3)]	-	-	✓[2)]	-
CO-RE 384 Probe Head	✓	✓[4)]	-	-	-	-

[1)] Available only for use with new disposables; not available for needles

[2)] Available with VENUS ONE TADM FEATURE (PN 911099)

[3)] Available on channels A1 and H12

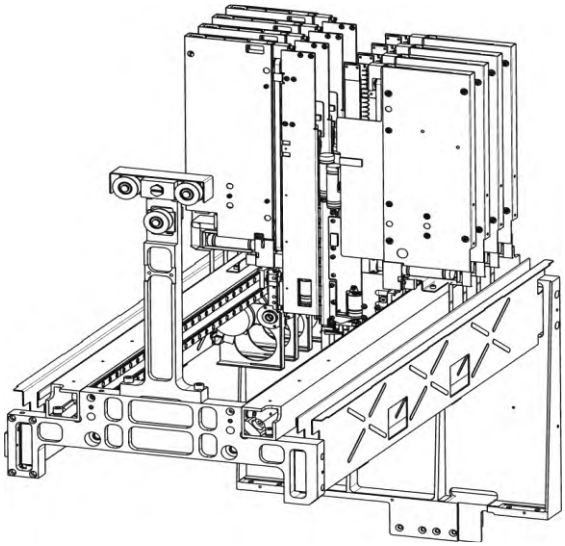
[4)] Available on channels A5 and P24

2.2 Pipetting Arm Configurations

The ML STAR offers a selection of a variety of arms, depending on the pipetting units and plate handling modules chosen. The pipetting arm moves in the x-direction. Whenever higher throughput is required, it is possible to have two arms on the system working in parallel. For instance, while one arm is reserved for the pipetting tasks, the other can transfer plates on the deck or to/from a peripheral device. There are further channel-based tools which can be mounted on the respective arms. Some example configurations are given below.

2.2.1 Modular Pipetting Arm

The Modular Pipetting Arm typically contains a set of pipetting channels, which work independently. It can be equipped with up to 16 1000 µl pipetting heads, up to 8 5 ml pipetting heads, 1 plate handling tool (iSWAP), 1 tube handling tool and 1 imaging channel.



Modular Pipetting Arm

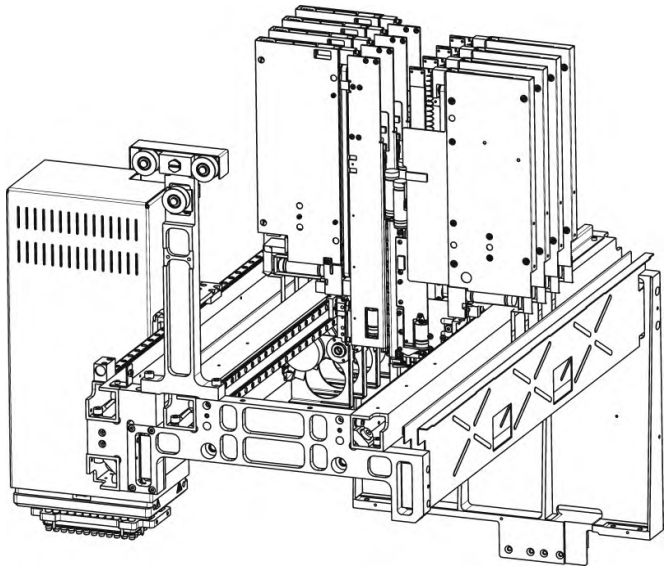
A possible configuration which includes all the tools is; 6 1000 µl pipetting heads, 2 5 ml pipetting heads, 1 tube handler, 1 imaging channel, with or without 1 plate handling tool (iSWAP).

The minimum distance between 2 – 1000 µl pipetting channels on this arm is 9 mm.

The minimum distance between 2 – 5 ml pipetting channels on this arm is 18 mm.

2.2.2 Modular Pipetting Arm MPH

The Modular Pipetting Arm MPH may include: 1 CO-RE Multi Probe Head (CO-RE 96 Probe Head, a CO-RE 96 Probe Head TADM or a CO-RE 384 Probe Head) as an option paired with up to 12 1000 µl pipetting heads, up to 8 5 ml pipetting channels, 1 plate handling tool (iSWAP), 1 tube handling tool and 1 imaging channel.



Modular Pipetting Arm MPH

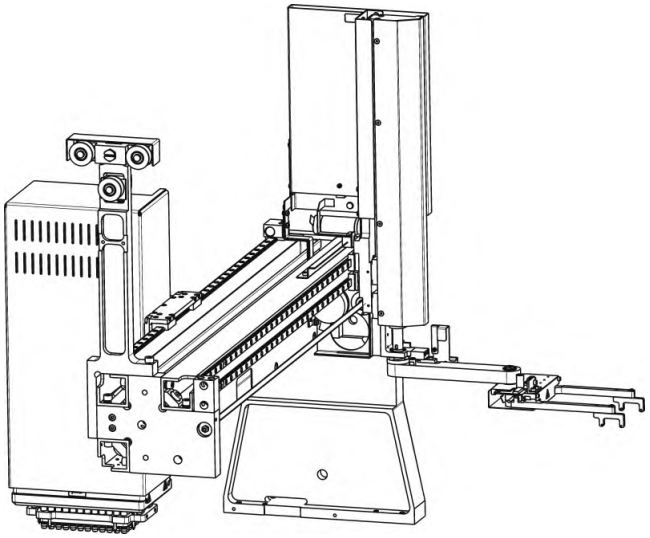
A possible configuration which includes all the tools is; 1 CO-RE Multi-Probe Head (CO-RE 96 Probe Head, CO-RE 96 Probe Head TADM or CO-RE 384 Probe Head), 6 1000 µl pipetting heads, 2 5 ml pipetting heads, 1 tube handler, 1 imaging channel, with or without 1 plate handling tool (iSWAP).

The minimum distance between 2 – 1000 µl pipetting channels on the modular pipetting arm MPH is 9 mm.

The minimum distance between 2 – 5 ml pipetting channels on the modular pipetting arm MPH is 18 mm.

2.2.3 Pipetting Arm MPH/iSWAP

The Pipetting Arm MPH/iSWAP may include: 1 CO-RE Multi Probe Head (CO-RE 96 Probe Head, a CO-RE 96 Probe Head TADM or a CO-RE 384 Probe Head) on the left. The right side of this arm can be equipped with up to 8 1000 µl pipetting heads, up to 4 5 ml pipetting heads, 1 plate handling tool (iSWAP), 1 tube handling tool and 1 imaging channel.



Pipetting Arm MPH/iSWAP

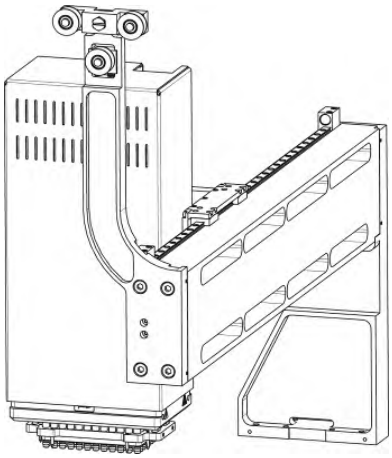
A possible configuration which includes all the tools is: 1 CO-RE Multi Probe Head (CO-RE 96 Probe Head, CO-RE 96 Probe Head TADM or CO-RE 384 Probe Head), 2 1000 µl pipetting head, 1 5 ml pipetting head, 1 tube handler, and 1 imaging channel, with or without 1 plate handling tool (iSWAP).

The minimum distance between 2 – 1000 µl pipetting channels on the pipetting arm MPH/iSWAP is 18 mm.

The minimum distance between 2 – 5 ml pipetting channels on the pipetting arm MPH/iSWAP is 36 mm.

2.2.4 Pipetting Arm MPH

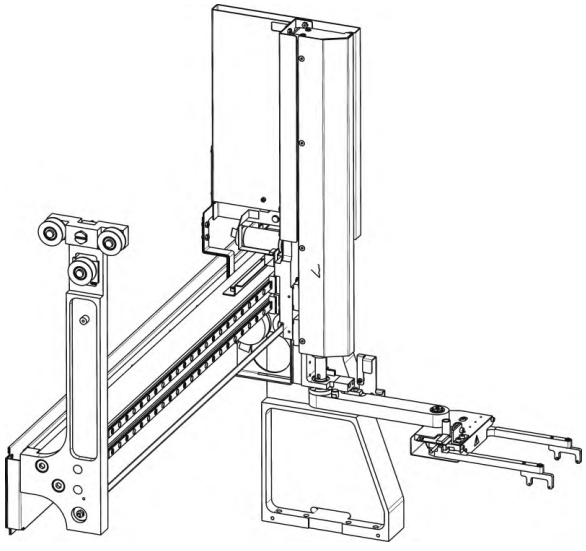
The Pipetting Arm MPH is equipped with 1 CO-RE Multi Probe Head (CO-RE 96 Probe Head, CO-RE 96 Probe Head TADM or CO-RE 384 Probe Head).



Pipetting Arm MPH

2.2.5 iSWAP Arm

The iSWAP Arm is typically fitted with the iSWAP for dual-arm configurations. This arm can be equipped with up to 8 1000 µl pipetting heads, up to 4 5 ml pipetting heads, 1 plate handling tool (iSWAP), 1 tube handling tool and 1 imaging channel.



iSWAP Arm

A possible configuration which includes all the tools is: 2 1000 µl pipetting heads, 1 5 ml pipetting head, 1 tube handler, 1 imaging channel, with or without 1 plate handling tool (iSWAP).

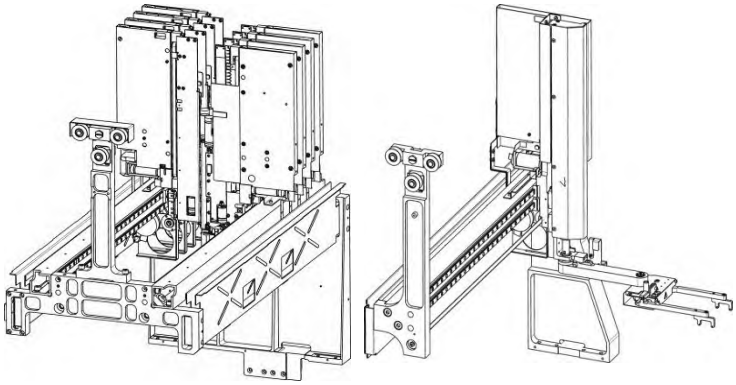
The minimum distance between 2 – 1000 µl pipetting channels on the iSWAP arm is 18 mm.

The minimum distance between 2 – 5 ml pipetting channels on the iSWAP arm is 36 mm.

2.2.6 Dual-Arm Configurations

When higher throughput is required, it is possible to equip the ML STAR with two arms working in parallel. The preferred platform for dual-arm configurations is the ML STARplus. The following examples show some typical dual arm configurations. Several other combinations are possible.

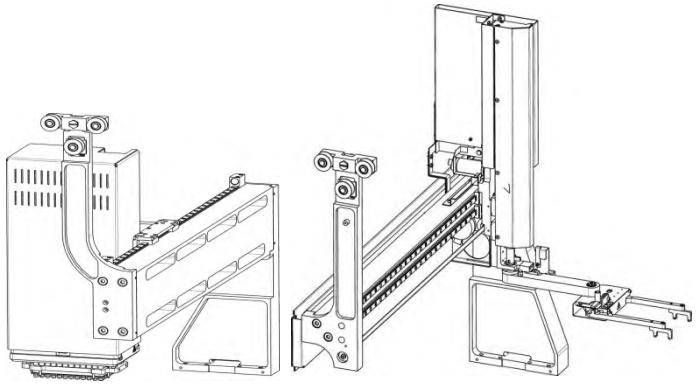
2.2.6.1 Dual Arm Assay Workstation 8+iSWAP



Modular Pipetting Arm with 8 1000 µl Pipetting Channels combined with the iSWAP Arm

This workstation, for instance, makes sense if the sample preparation task (processed by 8 1000 µl pipetting channels) is isolated from the assay tasks (such as incubation tasks, plate washing tasks, analyzing tasks, etc). The iSWAP is used to transfer the processed microplate to/from the 3rd party equipment such as a reader, incubator, plate washer, etc.

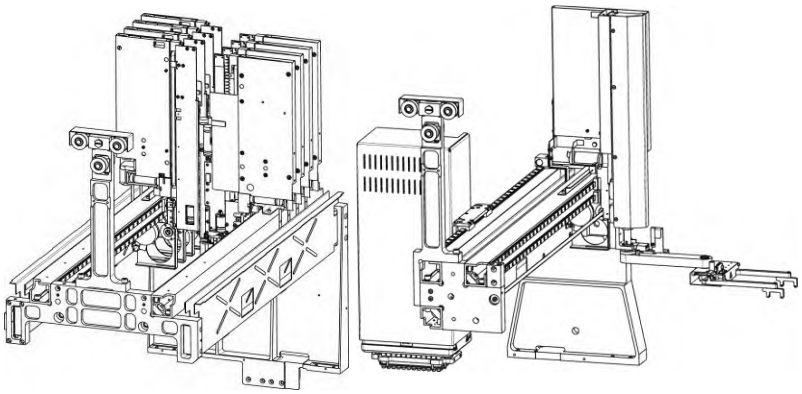
2.2.6.2 Dual Arm Assay Workstation, 96+iSWAP



Pipetting Arm 96 with the CO-RE 96 Probe Head combined with the iSWAP Arm

This workstation, for instance, makes sense if the pipetting tasks (plate copy, plate reformation, serial dilution) take part in the format of microplates only, and the assay needs no further pipetting steps. In addition, plate transfer to and from other devices is required.

2.2.6.3 Dual Arm Assay Workstation 8+96/iSWAP



Modular Pipetting Arm with 8 1000 µl Pipetting Channels combined with a 96/iSWAP Arm

This workstation, for instance, makes sense if the sample preparation task (processed by 8 1000 µl pipetting channels) is isolated from the assay tasks (as incubation tasks, plate washing tasks, analyzing tasks, etc). The assay task needs pipetting in the pattern of the CO-RE 96 Probe Head (one volume per plate, row or column). The iSWAP is used to transfer the processed microplate to and from 3rd party equipment, such as a reader, incubator, plate washer, etc.

2.3 Options and Upgrades

NOTE

This list of options and upgrades does not claim to be complete. Please contact your HAMILTON representative for more information.

Options are defined as components or configurations that are part of the instrument. Predefined options are as follows: Manual load or Autoload, the type and quantity of pipetting arms, the quantity of pipetting channels from 1 to maximum 16 1000 µl pipetting channels or from 1 to maximum 8 5 ml pipetting channels, 1 CO-RE Multi Probe Head, (the CO-RE 96 Probe Head, CO-RE 96 Probe Head TADM, CO-RE 384 Probe Head), 1 plate handling tool (iSWAP), 1 tube handling tool and 1 imaging channel.

Accessories include assemblies such as wash stations, temperature controlled carrier, vacuum system, labware carrier, etc (see Section 2.4 Accessories).

Pipetting channels, a CO-RE Multi Probe Head and accessories may be ordered as an option for new installation, or added as upgrades to an existing instrument.

The instrument's configuration is set within the configuration editor of the VENUS Software by a HAMILTON Trained Field Service Engineer.

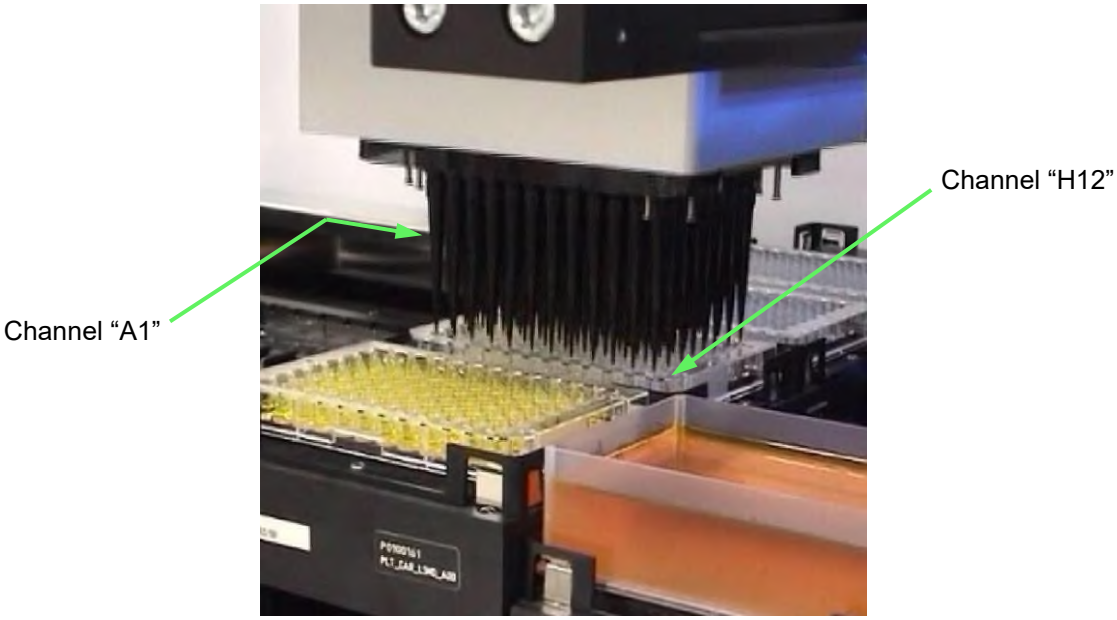
NOTE

An option may require a library, labware definitions etc. for use with the VENUS software. Please consult your local HAMILTON representative.

2.3.1 CO-RE 96 Probe Head

The CO-RE 96 Probe Head is a high-throughput pipettor built with the same CO-RE technology as the ML STAR pipetting channel. The CO-RE technology guarantees fast and accurate pick-up and release of disposable tips.

cLLD (capacitive Liquid Level Detection) is available on the two special channels A1 and H12.



View from the Right to Left Side of the Instrument

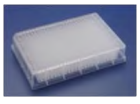
The CO-RE 96 Probe Head is available with 96x 1000 µl pipetting channels. The 96 pipetting channels always work simultaneously with the same volume.

The CO-RE 96 Probe Head supports pipetting with low-volume (10 µl), intermediate volume (50 µl), standard (300 µl), high volume (1000 µl), piercing and slim disposable tips. The following table gives an overview of disposable tips which can be used with the CO-RE 96 Probe Head. Other combinations may be possible, but require special attention.

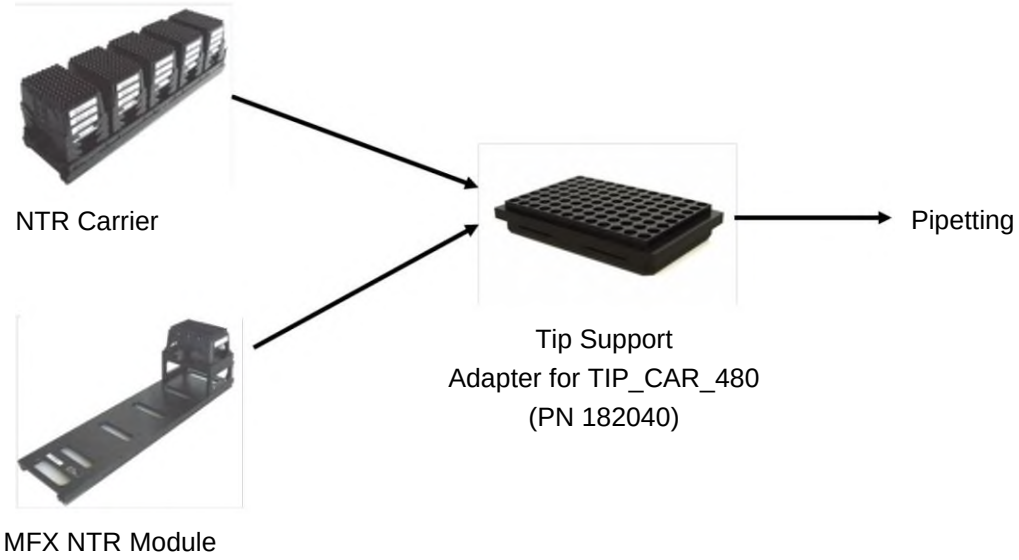
NOTE

If a type of tip is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM	**Tips for Working Volume and Purpose**					
Plate Type	10 µl CO-RE Tip	50 µl CO-RE Tip	300 µl CO-RE Tip	1000 µl CO-RE Tip	250 µl Robotic Piercing Tip	300 µl Slim Tip
 96-Well						
 96-Well Archive						
 384-Well						
 384-Well Archive						

The use of a tip support makes picking up one or more columns, rows, a single quadrant or even a single tip feasible from a Nested Tip Rack (NTR) with the CO-RE 96 Probe Head.



NOTE

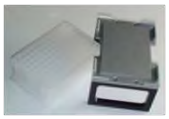
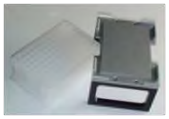
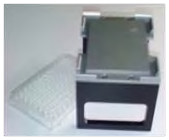
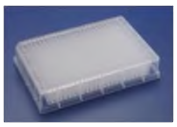
For shifted tip pickup, make sure to use a tip support (PN 182040) as an intermediate storage position for tips. It allows picking up of a single tip/row/column using the CO-RE 96 Probe Head.

The following table gives an overview of carriers and modules which can be used with the CO-RE 96 Probe Head. Other combinations may be possible, but require special attention.

NOTE

If a type of carrier or module is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM	Carriers and Modules			
 96-Well	 Plate Carrier for 96/384 Well Plates or  Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	 Multiflex MTP Module or  Multiflex DWP Module
 96-Well Archive	 Plate Carrier for Deep Well Plates	 MFX MTP Nested		 Multiflex DWP Module
 384-Well		 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	
 384-Well Archive		 MFX MTP Nested		

2.3.2 CO-RE 96 Probe Head TADM

HAMILTON´s proprietary TADM feature is available with the CO-RE 96 Probe Head TADM. All other features of this head are identical to those of the CO-RE 96 Probe Head. The CO-RE 96 Probe Head TADM is compatible with MPH ready ML STAR instruments and runs with VENUS TADM feature as well as with VENUS four.

The CO-RE 96 Probe Head TADM supports pipetting with low-volume (10µl), intermediate volume (50µl), standard (300µl), high volume (1000µl), piercing (limited) and slim disposable tips. The CO-RE 96 Probe Head TADM could be used with piercing tips but does not work in all labware, thus makes pipetting with piercing limited.

The CO-RE 96 Probe Head TADM does not have enough force in the z-drive to pierce a 96 rack format occupied by tubes with caps with standard tips. It is only possible to pierce a 96-well microplate with a thin aluminum foil, therefore piercing tips are used to pierce a 96-well microplate with the aluminum foil or tubes with caps.

2.3.3 CO-RE 384 Probe Head

The CO-RE 384 Probe Head is a parallel pipettor. It uses the CO-RE technology as in the individual pipetting channels and the CO-RE 96 Probe Head, offering accurate and gentle tip pickup. The positioning accuracy allows pipetting into 96, 384 and 1536 well microplates. The 384 pipetting channels always work simultaneously with the same volume. The pipetting range is between 0.1µl and 50µl.

NOTE

Special 50µl tips are designed for pipetting with the CO-RE 384 Probe Head.

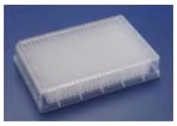
cLLD (capacitive Liquid Level Detection) is available on two special channels (A5 and P24).

The following table gives an overview of disposable tips which can be used with the CO-RE 384 Probe Head. Other combinations may be possible, but require special attention.

NOTE

If a type of tip is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

CO-RE 384 Probe Head	Tips for Working Volume	
Plate Type	50 µl Tip	300 µl Tip
 96-well	 384 Probe Head CO-RE Tips 50 µl	 Rocket Tip
 96-Well Archive	 384 Probe Head CO-RE Tips 50 µl	 Rocket Tip
 384-Well	 384 Probe Head CO-RE Tips 50 µl	
 384-Well Archive	 384 Probe Head CO-RE Tips 50 µl	
 1536-Well	 384 Probe Head CO-RE Tips 50 µl	

The use of special Nested Tip Racks (NTRs) equipped with 96 tips per rack (PN 235993, PN 235447, PN 235695 sterile, PN 235825 sterile; 96 of the 384 positions filled with tips) converts the CO-RE 384 Probe Head into a 96 pipetting channel head with a volume range of 50µl.

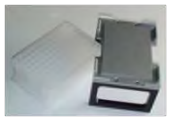
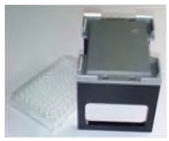
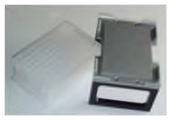
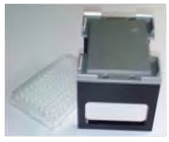
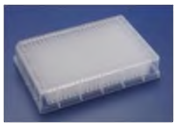
The use of HAMILTON's Rocket Tips (PN 235974) converts the CO-RE 384 Probe Head into a 96 pipetting channel head with a volume range of up to 300µl. Pickup of one or more columns, rows, a single quadrant or even a single Rocket Tip is feasible from the raised tip rack with the CO-RE 384 Probe Head is possible from the standard.

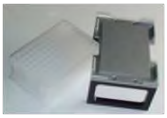
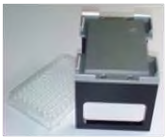
The following table gives an overview of carriers and modules which can be used with the CO-RE 384 Probe Head. Other combinations may be possible, but require special attention.

NOTE

If a type of carrier or module is not shown, it cannot be used for that purpose.

See Section 8.1 Appendix A: Ordering Information for a complete listing.

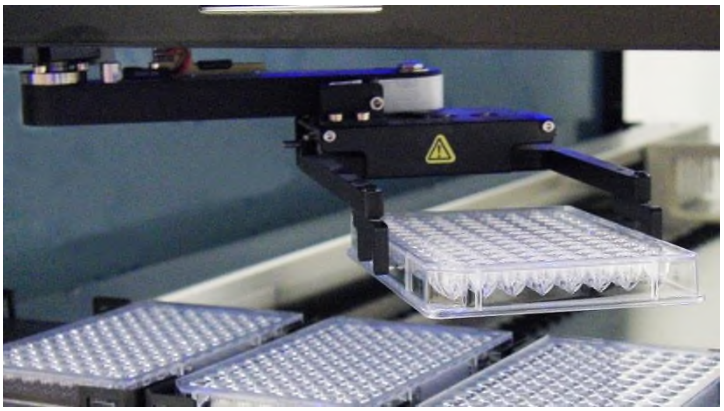
CO-RE 384 Probe Head	Carriers and Modules			
 96-Well	 Plate Carrier for 96/384 Well Plates or  Deep Well Plates	 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	 Multiflex MTP Module or  Multiflex DWP Module
 96-Well Archive	 Plate Carrier for Deep Well Plates	 MFX MTP Nested		 Multiflex DWP Module
 384-Well		 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	
 384-Well Archive		 MFX MTP Nested		

CO-RE 384 Probe Head	Carriers and Modules			
 1536-Well		 MFX DWP Nested or  MFX MTP Nested	 Plate Carrier for 1536-Well Plates	

2.3.4 iSWAP (Landscape)

The iSWAP (internal Swivel Arm Plate Handler) is a robotic arm that transports microplates, lids of microplates, archive plates, filter plates to and from positions on the instrument deck. Plates can be placed in landscape or portrait orientation or rotated. It is possible to pick up labware from and place it to labware any position inside or even outside of the deck. Typical handling tasks like loading/unloading of plates outside the instrument becomes very simple with the iSWAP. The iSWAP can also stack plates or tips with specially provided carriers.

Among the special features of the iSWAP is a sensor which signals when a labware object is gripped, which monitors the presence of an object. Like the ML STAR pipetting channels, the iSWAP has a "traverse height" of 145 mm above the deck (245 mm above the origin).



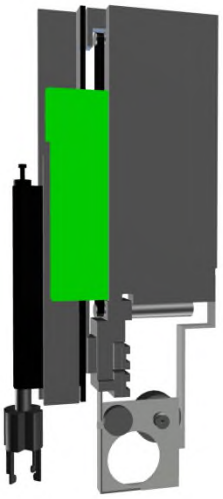
iSWAP Robotic Arm at work Transporting Microplates

The iSWAP is mounted on the pipetting arm. It does not affect the movement of the pipetting tools.

2.3.5 Tube Gripper

The Tube Gripper transports tubes to and from positions on the instrument deck. The tube-gripper tool is mounted to a separate channel. It can handle tubes with diameters from 8 mm up to 20 mm and up to a length of 120 mm.

The tube-gripper is moved independently on the y-axis, as well as on the z-axis. The use of high-precision motors and electronics allows the tube-gripper to reach any position on the deck without the need of teaching.



Tube Gripper Channel

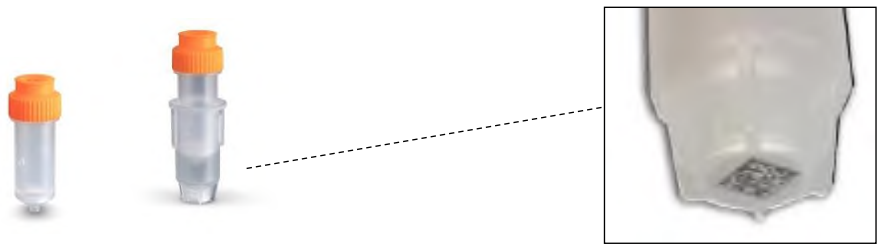
Like ML STAR pipetting channels, the tube-gripper has a "traverse height" of 145 mm above the deck (245 mm above the origin) between the tube and the deck of the instrument. This means that when a tube is moved from one location on the deck to another, it automatically does so at that particular height. This is a safety precaution, so the transported tube will not collide with any items on the deck.

2.3.6 AutoLys Tube Channel

The AutoLys channel handles AutoLys tubes. It is mounted to a separate channel.

AutoLys Channel

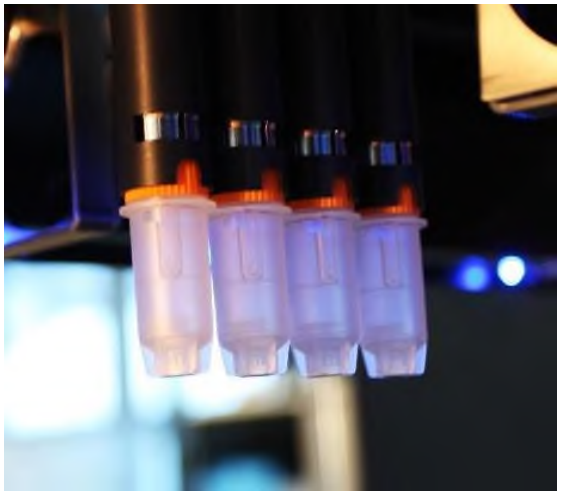
The AutoLys tube is an automation-friendly consumable designed for sample lysis and extraction. It is a 3-part assembly consisting of an outer tube with a 2D barcode, an inner tube with flow-thru filter and a threaded cap. The AutoLys tube facilitates processing and filtering of up to 500µl of lysate. The lysate is usually captured by centrifugation.



AutoLys Tube (right: spin position)

2D Barcode on the bottom of the AutoLys Tube

Four AutoLys Tool Channels



Four AutoLys Tool Channels holding AutoLys Tubes

The AutoLys Tool Channel is moved independently on the y-axis, as well as on the z-axis. The use of high-precision motors and electronics allows the AutoLys tool channel to reach any position on the deck without the need of teaching. It automates the entire process of AutoLys tube handling during a lysis procedure:

- De-capping the AutoLys tube, holding the cap and re-capping the tube
- Transfer the AutoLys tube to and from racks
- Lifting the inner tube and locking it in the "spin position"
- Removing and holding the inner tube post centrifugation
- Replacing the inner tube in the original outer tube

2.3.7 Tube-Twister Channel and Decapper Module

2.3.7.1 Tube-Twister Channel

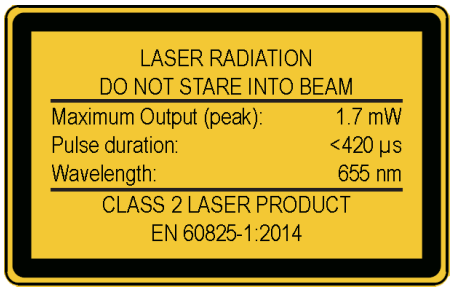
The Tube-Twister Channel can grip and rotate or mix tubes, read their barcodes and transport the tubes to and from positions on the instrument deck. The Tube-Twister is mounted to a separate channel. It can handle tubes with diameters from 15 mm up to 38 mm and caps from 18 mm up to 44 mm. The maximum tube length including cap is 120 mm.

The Tube-Twister Channel is moved independently on the y-axis, as well as on the z-axis. The use of high-precision motors and electronics allows the Tube-Twister Channel to reach any position on the deck without the need of teaching. The attached Tube-Twister Head is specific for the Tube-Twister Channel and includes two drives. One Twister Drive is able to grip tubes with its four fingers. The other Twister Drive rotates the Gripper Tool. This motion can be used to rotate the tube for barcode identification and for alignment, placing the tube in a carrier with the barcode in the correct position. A 1D barcode reader is attached to the Tube-Twister Channel. Barcode reading is executed after gripping the tube. The motion can also be used for mixing liquids inside the tube.

ATTENTION

LASER BEAM

The Barcode Scanner on the optional Tube Twister Channels of the ML STAR has a Class II Laser Diode. Do not stare into the beam.



Optional Tube Twister Channels

Four Tube Twister Channels holding tubes

Like ML STAR Pipetting Channels, the Tube-Twister has a "traverse height" of 145 mm above the deck (245 mm above the origin) between the tube and the deck of the instrument. This means that when a tube is moved from one location on the deck to another, it automatically does so at that particular height. This is a safety precaution, so the transported tube will not collide with any items on the deck.

In combination with the De/Re-Capper Module, the Tube-Twister can also be used for decapping and recapping of tubes.

ATTENTION

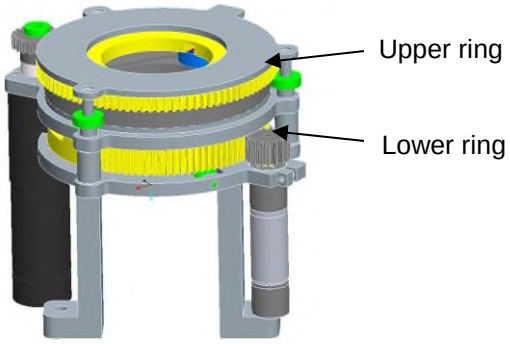
Do not overfill the tubes: fill approximately 15 mm below the top of the tube to avoid spillage.

When mixing liquid, the mixing speed has to be adjusted to the type of tube and the volume within the tube to prevent cross-contamination due to spillage.

Do not mix liquid (spinning), while tubes are open. Ensure that the tubes are completely closed before processing.

2.3.7.2 Decapper Module

In combination with the Tube-Twister, the Decapper Module allows opening and closing tubes with screw-top caps. A decapper module consists of up to four decapper positions, where each position consists of two rings. The upper ring clamps and turns the cap, the lower ring clamps the tube. For reliability and increased process control, the de/re-capper module is equipped with independent upper ring drive and lower ring drive.



De/Re-Capper Module

ATTENTION

Glass tubes cannot be opened or closed with the Decapper Module.

Tubes with equal or smaller diameter caps cannot be opened or closed with the Decapper Module.

For de-capping, a capped tube is presented to the Decapper Module using the Tube Twister Channel. The lower ring of the Decapper is closed in order to hold the tube. The upper ring is closed to hold the cap and disengage the cap from the tube. The Tube Twister Channel unscrews the cap and discards it into the waste or holds it for further re-capping. The lower ring of the module releases the tube. At this stage, the pipetting channels have access to the liquid in the tube. Alternatively, the tube can be transported to a different location by the Tube Twister Channel.

For re-capping, an uncapped tube is presented to the Decapper using the Tube Twister Channel. The lower ring of the Decapper is closed in order to hold the tube. The Tube Twister Channel gets a cap from a cap supply and screws the cap to the tube with minimal force. The cap is then finally locked using the upper ring of the Decapper. The capped tube is released and can be transported by the Tube Twister Channel.

2.3.8 Imaging Channel

The purpose of the imaging channel is to capture the digital image from any object placed on the instrument deck. A high-resolution CCD-Camera is fixed on a separate channel and allows acquiring images from a target which is as big as the SBS format. The images are sent to the image analyses software for further investigation.

A typical application is the image analysis of bacteria colonies. In the proprietary easyPick software, the user defines parameters and determines the weighting for the typical criteria to identify colonies: coordinates, size, shape (circularity), color, proximity to the next colony, distance to the margin, etc.

Reliable automation requires proper quality of the analyzed images. Appropriate lighting of the object of interest is fundamental. In case of image analyses of bacteria colonies a back-light table is used in the system. For other applications different lighting options can be applied.

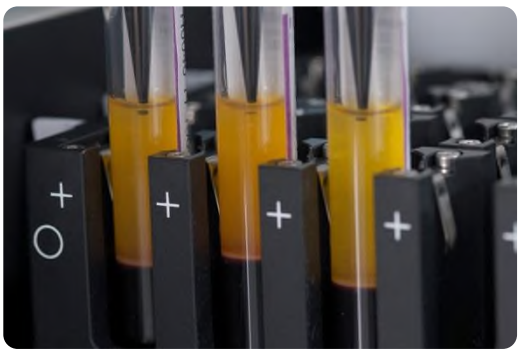
2.3.9 Imaging Applications

- Camera on a separate Channel
- Light Table Carrier
- Image Analysis Software

2.3.9.1 easyBlood

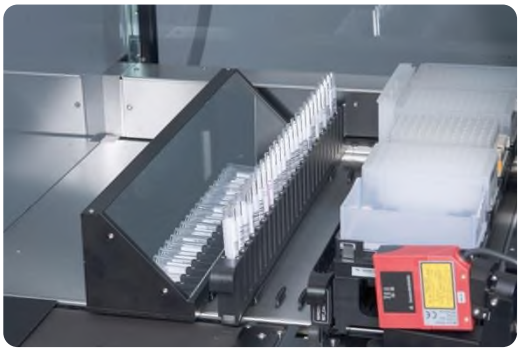
Camera Channel:

- Required for the fraction detection
- Fixed focal length
- Takes picture of 4 tubes at once



Mirror Carrier:

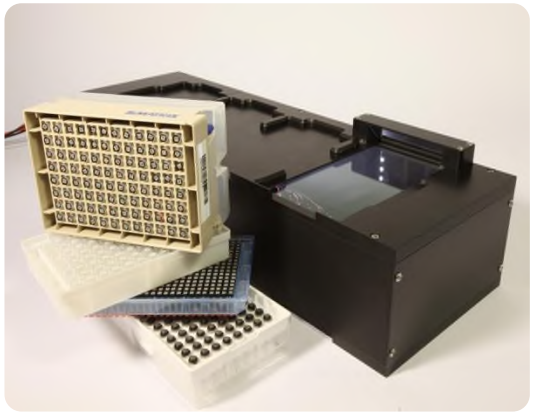
- Presentation of Tubes to the Camera



2.3.9.2 easyCode Carrier

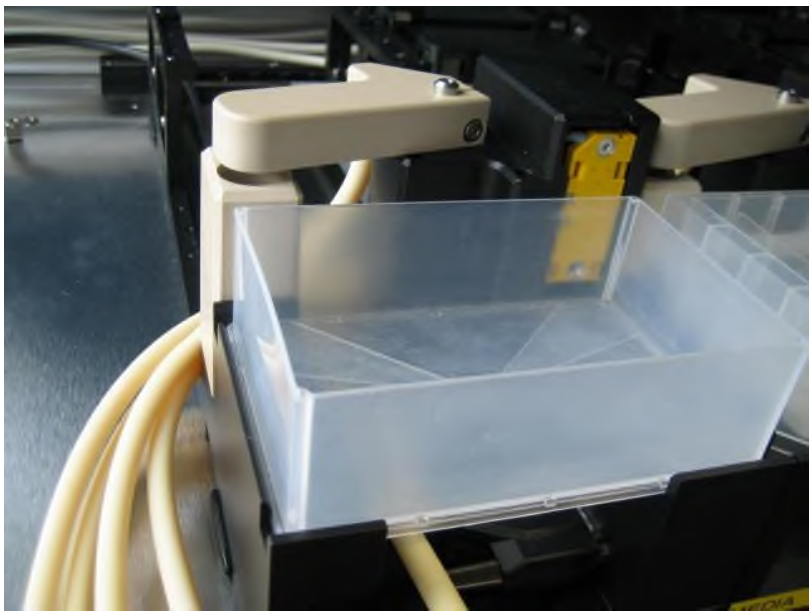
easyCode Carrier:

- Code Identification achieved with integrated HAMILTON 2D Code scanner carrier
- Small Footprint: 6T (7T) Carrier
- Reading time: 1 sec
- Decoding time: 96 Tubes -> 3 sec
- For all types of racks with 12, 24, 48, 96, 384 tubes
- Reads all types of codes:
- FluidX, Matrix, Micronic, NUNC, Abgene, REMP/Nexus, HAMILTON, Greiner, Axygen, Corning, all Datamatrix ECC 200
- "No tube"- recognition, differentiation between missing tubes and unreadable codes



2.3.10 Multiflex Media Line

The Multiflex Media Line is used to fill a SBS Reagent Container on the Deck of a STAR Instrument, from which a MPH or single pipetting channels can aspirate medium. In comparison to the Multiflex Liquid Dispenser, which has a similar function, the Media Line is more sterile. Also, the temperature of the medium is controlled by the Pre-Heating Module. All tubing including Dispense Arm and Drain can be autoclaved and can be replaced easily by the customer. Typically, one medium is used to fill the Reagent Container, whereas a disinfectant and water is used for cleaning and rinsing.



The Media Line is mounted on a special Multiflex Base Plate with an overfill basin. The Liquid Level of the Reagent Trough is controlled by a Capacitive Liquid Level Sensor. One Peristaltic Pump Unit fills and maintains the level in the SBS Reagent Container, the other Peristaltic Pump Unit transports the liquids from the drain into a Waste Container (provided by the customer). The Dispense Arm with a spout has two defined positions: one over the Reagent Container for filling, the other over the drain for cleaning and rinsing the tubing. Up to 3 Media Lines can be mounted onto one Base Plate.

Optionally, a controlled heater plate can be mounted underneath the Media Trough.

For each Media Line, there is a safety overflow stop mounted in the overflow basin. Should the Liquid Level Sensor (due to a fault) fail to detect that the Reagent Container is full, the fill pump will be stopped automatically as soon as the safety overflow stop detects any fluid in the overflow basin.

2.3.11 CO-RE Gripper

The CO-RE gripper is a plate handling tool picked up by two pipetting channels during a run. This tool is only available if the system has at least two individual pipetting channels.

The CO-RE gripper transports microplates, lids for microplates, archive plates, filter plates, etc. to and from positions on the instrument deck. Plates can be gripped in landscape or portrait format within the working area. Rotation of plates is not an option. The traverse height of the pipetting channels with the gripping jaws is the same as with tips: 145 mm above the deck.

Given that there is no sensor implemented; you must ensure that the CO-RE gripper does not grip the plates too tightly which can cause step loss (blockage) of the pipetting channels when moving in the Y-direction. When the grip is too loose, it can cause a rattle and possibly drop the plates.

CO-RE Grippers mounted on waste block: 1000 µl Pipetting Channels (inner positions) and 5 ml Pipetting Channels (outer positions)

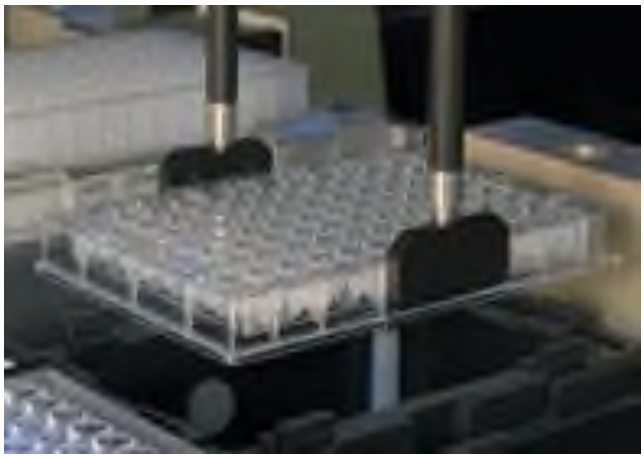


Plate being moved with CO-RE Gripper using 1000 µl pipetting channels

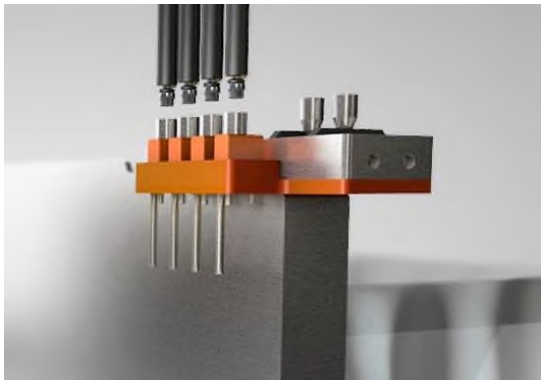
The CO-RE grippers are available for waste block- and for plate carrier mount. Plate carrier mount requires one freely selectable SBS position on deck, whereas waste mount does not consume space on deck.

NOTE

When using plate stacks or filter plate systems with an elution plate on the CVS, the stacked plates and elution plates are only accessible by the CO-RE gripper, NOT with the iSWAP.

2.3.12 FlipTube Tool

HAMILTON FlipTubes are 1.5 ml reagent tubes with a lid attached which can close the tube tightly for convenient and reliable preparation, centrifugation and storage of small volume samples. The FlipTube tool is used for opening and closing HAMILTON FlipTubes. It is picked up by a 1000 µl pipetting channel during a run. Up to four FlipTube tools can be used in parallel when picked up by four 1000 µl pipetting channels, if available. The traverse height of a channel with the FlipTube tool is the same as with tips: 145 mm above the deck.



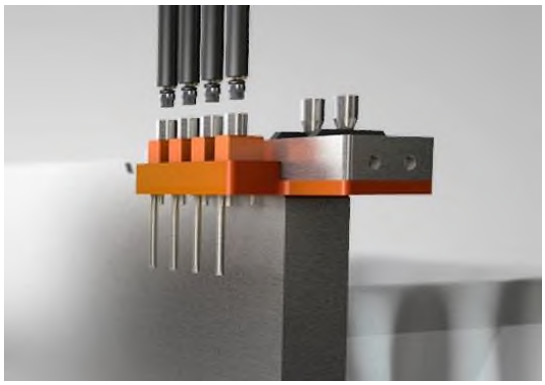
A Set of 4 FlipTube Tools (consisting of four fingers) within the holder

HAMILTON FlipTube A (PN 235454)

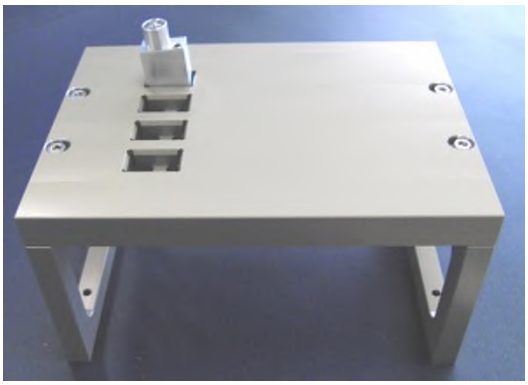
NOTE

The FlipTube Tool can be used for HAMILTON FlipTubes only, not for other locked tubes.

Park positions (holders) for FlipTube Tools are available for a Multiflex Carrier or for the waste block. The park position on the waste block has the advantage that it does not consume a plate position.



FlipTube Tool Park Position on the Waste Block.
The long rod faces to the left hand side on the deck.



FlipTube Tool Park Positions for a Multiflex Carrier in their correct orientation on the deck,
FlipTube Tool positioned on the left hand side, long rods facing left.

NOTE

FlipTube Tools have to be aligned on the deck such that the long rod is on the left hand side of the park position.

FlipTubes must be fixed tightly in a special carrier or adapter for opening and closing by a FlipTube tool. This is achieved by a specific sample carrier, an adapter for a Multiflex Module or an adapter for the HAMILTON Heater Shaker.

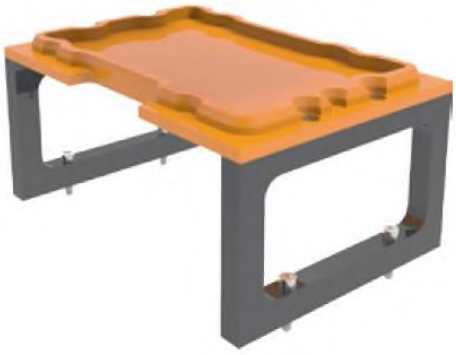
Sample Carrier for 32 FlipTubes SMP_CAR_32FT (PN 809030)

NOTE

The Multiflex adapter for FlipTubes and the HHS adapter for FlipTubes have to be aligned on the deck such that the lid of the tube faces to the left hand side.

NOTE

The Multiflex adapter for FlipTubes and the HHS adapter for FlipTubes have to be aligned on the deck such that the lid of the tube faces to the left hand side.



Multiflex Adapter for FlipTubes (PN 814275). Mounted such that the lids of the tubes face left.

HHS 3 (PN 199085) equipped with a built-in adapter for a FlipTube Rack.

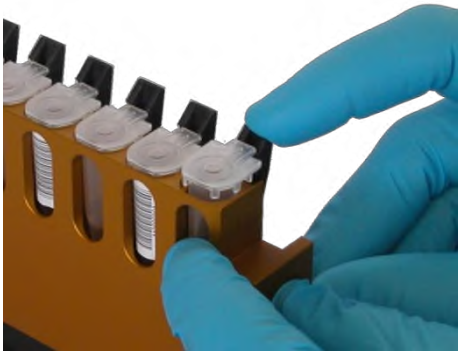
ATTENTION

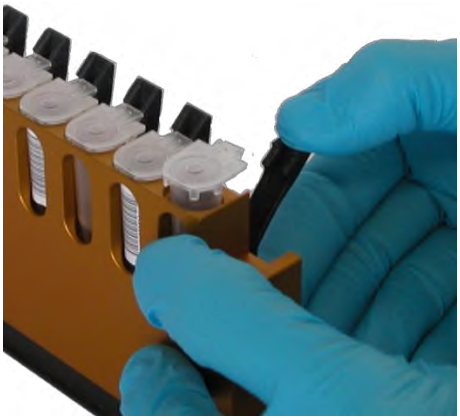
For best performance:

- *Use a demo method for optimal process safety*
- *Tip has to dispense inside the tube*

ATTENTION

Carefully use the spring elements of the FlipTube Carrier.





NOTE

FlipTubes are normally mounted on the waste block or in the SBS position; however, when the waste block is occupied by the CO-RE gripper, either the SBS position or the FlipTube tool will be used instead. For more information about FlipTubes, please refer to the FlipTube Manual. There is a spring on the carrier or adapter which can be adjusted when necessary.

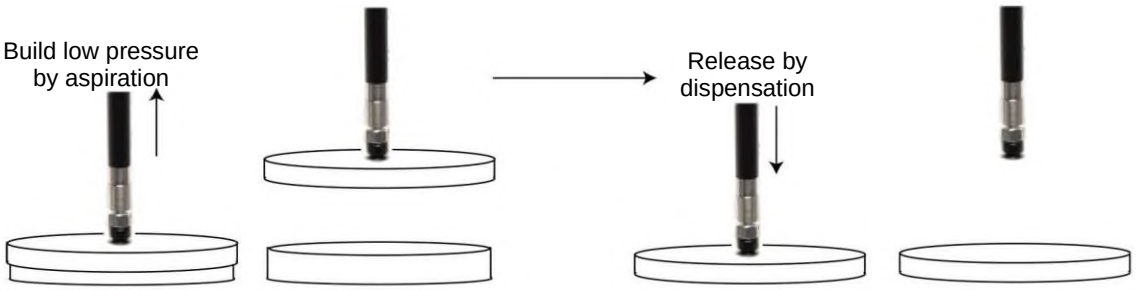
2.3.13 CO-RE Lid Tool

The CO-RE lid Tool (CLT) is a suction cup device for picking up and transporting lightweight lids of petri dishes, plates, troughs and reservoirs. It can be picked up by a 1000 µl pipetting channel. The plunger drive generates a negative pressure (vacuum) on the surface of flat clean labware objects so that they can be lifted. When not in use, the CO-RE Lid Tool is parked on the waste block in one of the teaching/maintenance needle positions.

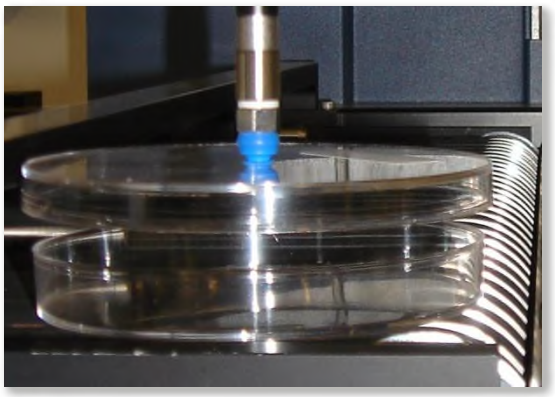
The CO-RE Lid Tool can also be parked on a SMP-CAR-32 (PN 173410).

- Labware: Petri-dish lids, microplate lids, trough and reservoir covers can be transported
- Park position: on the waste block in one of the teaching/maintenance needle positions
- Transport device: 1000ul-pipetting channel
- Move-Control: Move-control by a *.hsl library

Labware pickup and eject positions / tool park position / number of 1000ul-pipetting channels has to be defined.



Mode of operation of the CO-RE Lid Tool



CO-RE Lid Tool picking up the Lid of a Petri Dish

CO-RE Lid Tool on its Park Position on the Waste Station

For operation of the CO-RE lid tool, a special library is required. The labware pick-up and eject positions, the tool park position and the channel to be used have to be defined.

NOTE

Please consult the local HAMILTON representative for the specific library to use the CO-RE lid tool.

2.4 Accessories

NOTE

This list of accessories does not claim to be complete. Please contact your HAMILTON representative for more information.

Accessories are defined as additional automation components. They provide a high degree of adaptability and permit customizing for multiple applications. These components can be ordered later by the customer and installed by HAMILTON authorized personnel.

NOTE

An accessory may require a library, labware definitions, etc. for use with the VENUS software. Please consult a local HAMILTON representative.

NOTE

The availability of accessories is subject to change. Please consult a local HAMILTON representative or visit www.hamiltonrobotics.com.

2.4.1 Carriers

Labware such as plates or tubes are placed on special carriers that are loaded onto the ML STAR instrument deck. HAMILTON provides a wide range of standard carriers for microplates, tubes, tips, etc. All standard carriers can be loaded to the deck manually or by the Autoload option. The naming of carriers follows a systematic nomenclature "**X_CAR_Y_Ann**" where

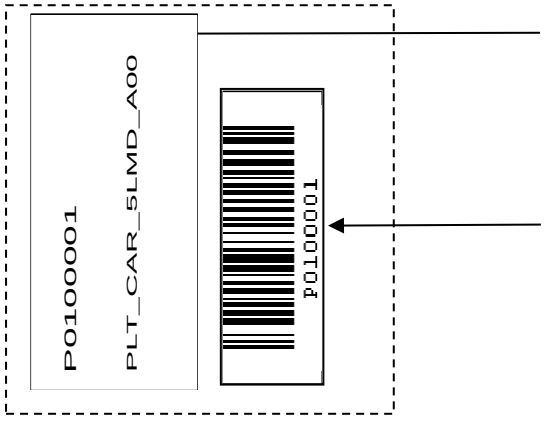
X represents the type of labware placed on the carriers, e.g.
TIP (= tips)
PLT (=plates)
SMP (=samples)
RGT (=reagent)

CAR carrier

Y describes the labware details, e.g.
L: landscape orientation
P: portrait orientation
Number: number of items placed on the carrier (plates or tips)
MD: medium density (96- or 384-well micro-plates)
HD: high density micro-plates (1536)
AC: 96-well archive plates

Ann identifies the part number revision (e.g. A00)
A: variant
nn: revision

Example: PLT_CAR_L5MD_A00 is a plate carrier for 5 medium density (96- or 384-well) micro-plates in landscape orientation.



Identification label on the front side of the carrier:

- carrier identification number
- carrier name

Carrier barcode label at the back of the carrier with carrier identification number:

- character 1 to 3: carrier type
- the rest is serial numbering

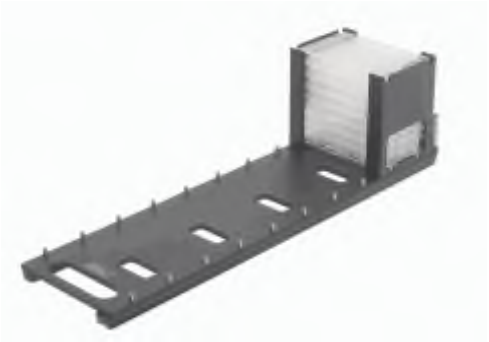
Carrier barcode labels

A carrier must always be identified in the VENUS software (e.g. in deck layouts and methods) by the unique description with which it is tagged.

2.4.2 Multiflex Carrier

The Multiflex Carrier consists of a multiple-use carrier base offering space for up to five modules. Several modules can be chosen, such as: tip rack module, microplate module, plate stacker module, module for heating or cooling labware, reagent trough module, tube or cup module, etc. The list of available modules can be found in Section 8.1 Appendix A: Ordering Information.

Multiflex Tip Module on the Multiflex Carrier



Multiflex Stacker Module (landscape) on the Multiflex Carrier

NOTE

For configuring Multiflex Carriers, a Multiflex Carrier Assistant Software Tool is required. Please consult a local HAMILTON representative.

NOTE

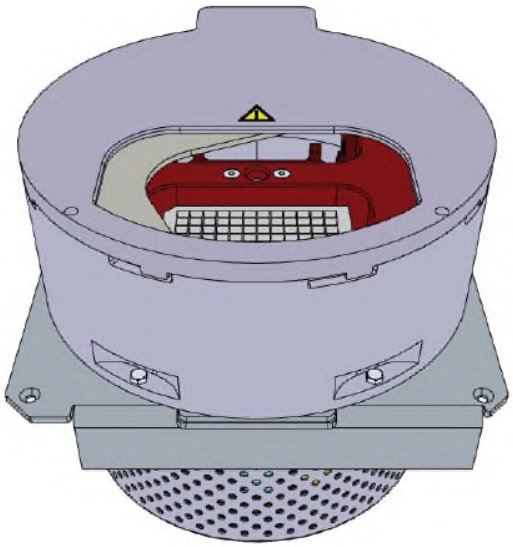
The availability of accessories is subject to change. Please consult a local HAMILTON representative for an up-to-date list, or consult www.hamiltonrobotics.com.

The modular design of the Multiflex Carrier allows space optimizing and customizing the instrument. This carrier will occupy 6 tracks of the instrument deck.

2.4.3 HAMILTON SBS Centrifuge

The HAMILTON SBS Centrifuge is a dedicated device for automated processing of SBS plate formats such as DWP, standard microplates, AutoLys racks, etc. The HAMILTON SBS Centrifuge is integrated into the ML STAR Instrument, and not used as a stand-alone device.

For detailed information about the HAMILTON SBS Centrifuge, refer to the HAMILTON SBS Centrifuge Operator's Manual.



2.4.4 HAMILTON Heater Shaker

The HAMILTON Heater Shaker (HHS) is designed to heat and shake microplates in SBS format. Loading and unloading as well the independent heating or shaking function of the HHS is fully controlled by the VENUS software.

For detailed information about the HAMILTON Heater Shaker, refer to the HAMILTON Heater Shaker Operator's Manual.

2.4.5 Multiflex Cooling-Heating Module

Generally; the Multiflex Cooling-Heating Module will be installed by a HAMILTON Trained Field Service Engineer.

Multiflex Cooling-Heating Module

The intended use of the Multiflex Cooling-Heating Module is to cool down or heat up a labware and keep it at that defined temperature. It is not a thermocycler and not a standalone module.

2.4.6 Multiflex Heating and Multiflex Cooling Module

The Multiflex Heating Module heats microplates whereas the Multiflex Cooling Module cools microplates. The temperature range for the heating module is from ambient +5 °C to 65 °C. The temperature range for the cooling module is from 15 °C to 4 °C. A temperature selection dial is located on the top of each module for setting the desired temperature. The status LED "**OFF**" indicates the Multiflex Heating or Cooling Module is coming to temperature. When the LED is "**ON**", the temperature of the Labware adapter is within ±1 °C of the desired temperature. It is possible to place up to 4 Multiflex carriers on one ML STAR instrument.

Multiflex Heating Module

Multiflex Cooling Module

ATTENTION

Do not touch the Multiflex heating module during run time and shortly after finishing a run. Wait until the Multiflex heating module has cooled down to room temperature, which can take up to 1 hour.

2.4.7 Multiflex Carrier Base

Instead of microplate shakers, however, the shaker carrier base can be equipped with ordinary microplate modules. The list of available modules can be found in Section 8.1 Appendix A: Ordering Information.

This carrier base will occupy 7 tracks of the ML STAR deck.

Carrier base with Multiple Modules

2.4.8 On Deck Thermo Cycler (ODTC)

The ODTC is for "on deck" integration, a superior quality product from INHECO adapted for the HAMILTON STAR Line. Any molecular biological workflow that needs repeated heating or cooling steps can be realized.

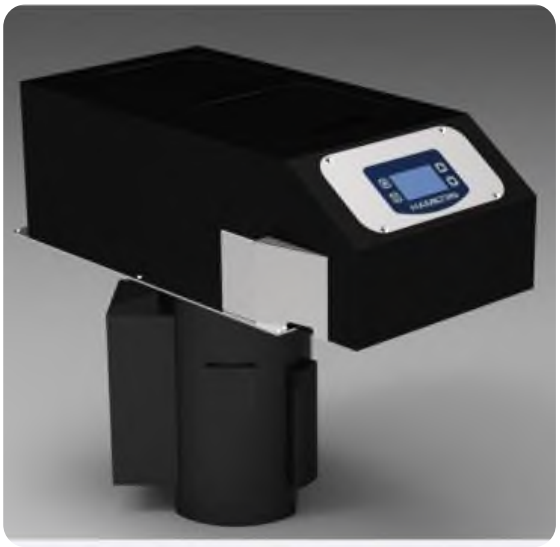
For detailed information about the ODTC Module, refer to the ODTC Module Operator's Manual.

2.4.9 Freezer Carrier

On-deck freezer carrier with two chambers for labware (microplate, mircotubes, plate frames) in SBS format. Temperature range -20 ºC – +50 ºC. The cooling/heating is performed by a Stirling engine and does not require any tubing, cryostat or cooling agents.

The freezer carrier is in total 11 T wide (10T for the module, 1T for air circulation) and requires space below the deck to accommodate the Stirling engine. A level 2 APE integration is required.

- Labware: Standard and Customized
- Temperature: -30 ºC up to ambient
- Control: manual or by hsl Library
- Load/unload: manual / CO-RE gripper



ATTENTION

Due to the design of the carrier, it can only be placed in certain positions on the deck and requires a special table. Also multiple tools are required for operation (power box, CO-RE grippers). Contact APE for further information!

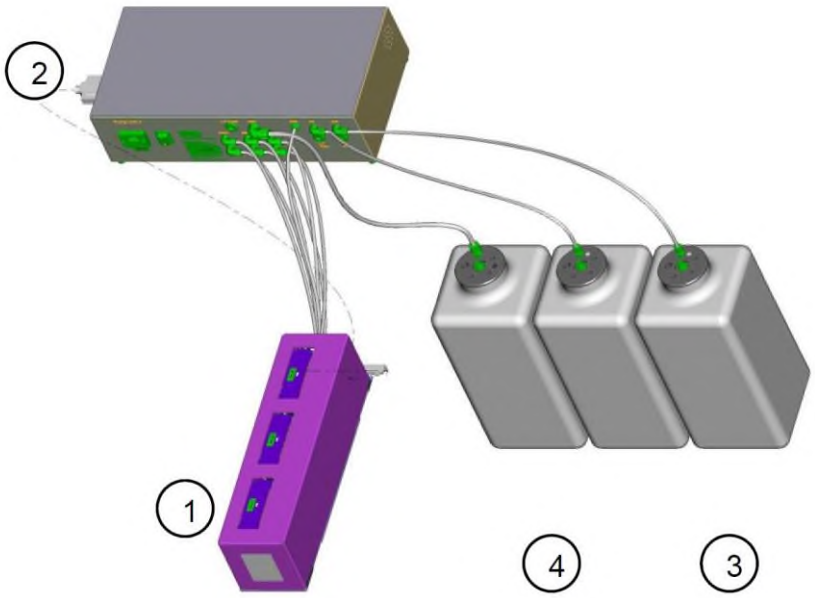
2.4.10 CR Needle Wash Station and Pump Unit

2.4.10.1 CR Needle Wash Station

The CR Needle Wash Station is a device for the washing of up to 24 steel needles in parallel to the pipetting process. The wash station has the width of a standard microplate carrier (6 tracks), and up to two needle wash stations can be mounted on the ML STAR instrument deck. The carryover of the wash station depends on the wash settings. Typical values are 10^{-5} to 10^{-6}. A set of default parameters is given for the wash process within the relevant dialog boxes of the VENUS Software. Wash parameters can be adapted.

Each wash station (item 1) is supplied with a pump station (item 2) with two reservoir containers (item 3) for wash solutions and one waste container (item 4). The wash and the waste containers have a capacity of 12 liters. LLD is provided in the wash containers to indicate when they are near empty and in the waste container to indicate when it is near full. The pump station is placed under the bench of the instrument.

A HAMILTON Trained Field Service Engineer must perform the installation of the Needle Wash Station.

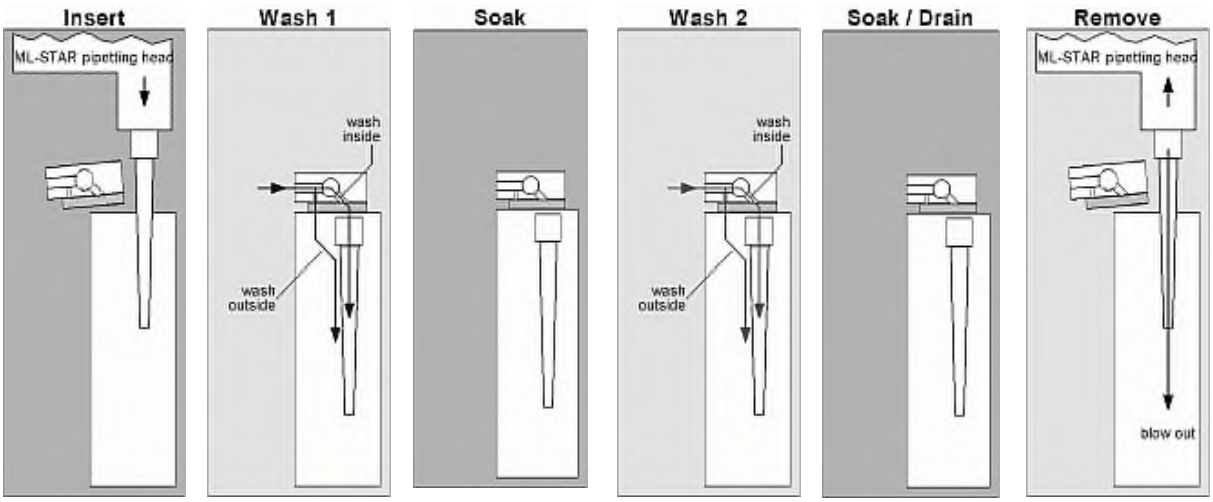


The wash station consists of three individual 8-fold wash modules. The modules are able to wash the 1000 µl needles, 300 µl needles or the 10 µl needles. Within one wash station module all 3 needle types can be washed.

The wash cycle works in parallel to the pipetting steps. In case of an 8-pipetting channel instrument, the following steps can take place:

1. The needles from the first module are picked up; they are used for pipetting, and then placed back into the same module. Needle washing starts.
2. While the needles of the first module are washed, the needles from the second module are picked up. After pipetting these needles are placed back into the second module. The wash cycle of this module is started parallel to the next steps.
3. While the instrument starts pipetting with the needles of the third module, the first module has finalized needle washing. After pipetting the needles are placed back into the third module. Then the wash cycle of this module can start.
4. Clean needles are picked up from the first module again. The process is then repeated.

The principle of the wash station is illustrated in the following figure which shows a typical procedure.



Schematic drawing of needle wash process: needles are placed into the wash module, washed from inside and along the outside using 1) wash and 2) rinse solution. The pipetting channels blow air through the needles to expel any residual liquid.

NOTE

Disposable tips may not be used in the needle wash station.

HAMILTON recommends replacing the needles every 6 months.

For a 16 pipetting channel instrument, two independent wash stations are necessary to enable high-throughput pipetting with one needle type.

NOTE

A table of chemical compatibilities with the wash station can be found in Section 8.2 Appendix B: Chemical Compatibility. The information provided is based on laboratory tests with raw materials and should be interpreted as a guideline only.

Consider local regulations for handling and storage of wash liquids regarding toxicity, contamination, fire protection, etc.

ATTENTION

The needle wash station for the ML STAR is not explosion-proof. When working with flammable or explosive fluids or vapors, necessary precautions must be taken.

2.4.10.2 CR Needle Wash Station Pump Unit

The CR Needle Wash Station Pump Unit includes two wash pumps and one waste pump. The wash pumps fill the wash chambers in the washer unit. The waste pump empties the wash chamber and/or the overflow chamber.

The Pump Unit for the Needle Wash Station has its own power supply. The main plug is on the rear of the pump station (see picture below).

Ensure that the needle wash station is correctly grounded when connected to the electrical outlet.

Ensure that the voltage selector on the pump station of the needle wash station is correctly set before operating the needle wash station. The needle wash station does not automatically switch for different voltage levels (115 VAC or 230 VAC).

The fuses for the pump station are located in the main socket (see picture below). The pump station has two fuses for the power supply which can be accessed by opening the cover underneath the main socket.

The technical specifications regarding electrical power and fuses to be used for the wash station are listed in Section 7.13 CR Needle Wash Station Specifications.

2.4.11 Dual Chamber 96/384 Wash Station

The Dual Chamber 96/384 Wash Station is an optional device for washing disposable tips of the CO-RE 96 and 384 Probe Heads. Low volume CO-RE tips, 50 µl volume CO-RE tips, standard volume CO-RE tips, 300 µl Rocket Tips, and high volume CO-RE tips without filter can be washed.

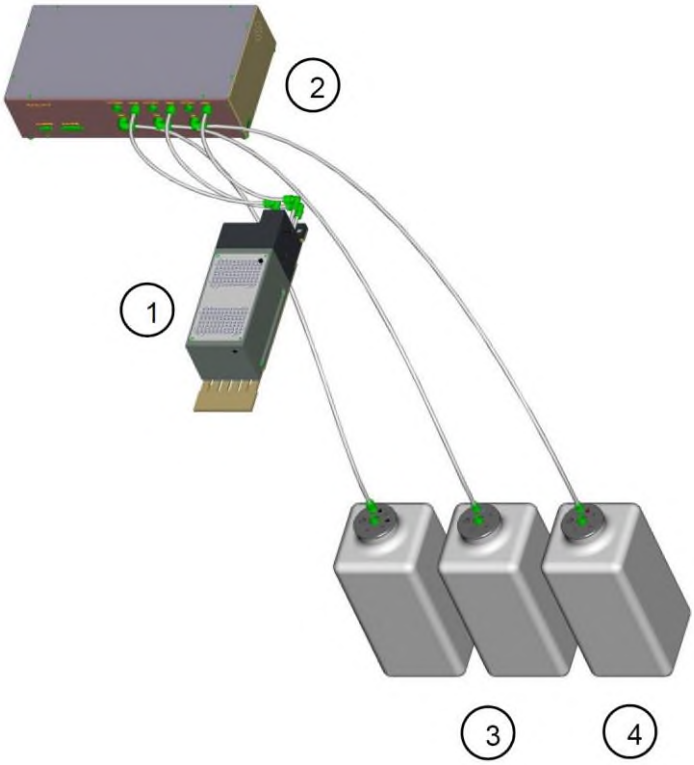
ATTENTION

Washing of filter tips is not recommended and not intended because there is a risk of contamination and a risk of damage to the pipetting device.

Washing takes place by aspirate/dispense cycles with the CO-RE 96 Probe Head or the CO-RE 384 Probe Head respectively.

The wash station has the width of 6 tracks and is mounted on the ML STAR instrument deck. Two wash chambers make the process of washing tips efficient. Each chamber can be filled individually with wash liquid out of two source containers.

Each wash station (item 1) is supplied with a pump station (item 2) with two reservoir containers (item 3) for wash solution and one waste container (item 4). Both the wash and the waste container have a capacity of 12 liters. LLD is provided in the wash containers to indicate when they are near empty and in the waste container to indicate when it is near full. The pump station is placed under the bench of the instrument.



The pump unit includes two wash pumps and one waste pump. The wash pumps fill the wash chambers in the wash station. The waste pump empties the wash chamber and/or the overflow chamber.

NOTE

When re-filling the wash liquid container the waste liquid container must be emptied as well.

When re-using washed tips, pipetting precision may decrease.

ATTENTION

Only use water or DMSO maximum 30%, as wash liquid for the wash station.

ATTENTION

The wash station for the ML STAR instruments is not explosion-proof. When working with flammable or explosive fluids or vapors, necessary precautions must be taken.

NOTE

Older versions of wash stations, i.e. the 96 chamber wash station and the 384 dual chamber wash station are tip pattern specific!

If 50 µl tips of the 384 Probe Head need to be washed in these older versions, the 384 dual chamber wash station must be used.

If Rocket Tips need to be washed in these older versions, the 96 chamber wash station must be used.

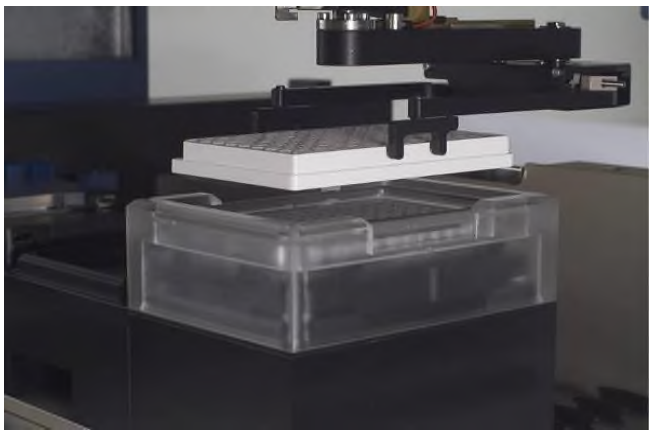
There is the possibility to field-upgrade an old version Wash Station to the Wash Station 96 / 384 Dual. Please consult a local HAMILTON representative for this possibility.

2.4.12 Crystal Vacuum System

2.4.12.1 Overview

The Crystal Vacuum System (CVS) replaces the older Basic Vacuum System (BVS) and allows automation of vacuum based kits for SPE, LC-MS, genomics and proteomics. CVS consists of a 7-track-wide carrier base equipped with the vacuum manifold, the park position for the manifold top and two microplate positions. By default the manifold top and insert for standard filter plates are included. For processing deep well filter plates, the deep well kit is required. The DWP Kit can be found in Section 8.1 Appendix A: Ordering Information.

The manifold top can be handled either by the iSWAP (see Section 2.3.5 iSWAP (Landscape)) or the CO-RE gripper (see Section 2.3.12 CO-RE Gripper). If working with the iSWAP, four tracks next to the CVS carrier have to be empty, usually on the right side.



The iSWAP in action to Load a Filter Plate to the CVS

The CVS accommodates a wide variety of 96-well and 384-well filter plates. With the height adjustable inserts inside the vacuum chamber, almost any kind of elution plates can be used. Four sets of nuts and bolts are included for adjusting the height as required.

The CVS carrier is mounted on the ML STAR instrument deck. It is possible to place up to four CVS carriers on one instrument.

NOTE

The Crystal Vacuum System is dependent on the elution plate's depth. If it deeply recessed, there is a need for it to be handled with the CO-RE grippers.

"Standard" Formats:

Deep Formats:

2.4.12.2 Vacuubrand Pump

The vacuum inside the chamber of the CVS is generated with the Vacuubrand pump. The maximum flow rate of the pump is ~5 m^3/h, and the possible final vacuum ~80 mbar. There is a built-in sound absorber that considerably reduces the pump noise level.

The vacuum controller CVC 3000 regulates the pump. Communication with the computer is via an RS232 cable, and its software is seamlessly integrated into the VENUS Software.

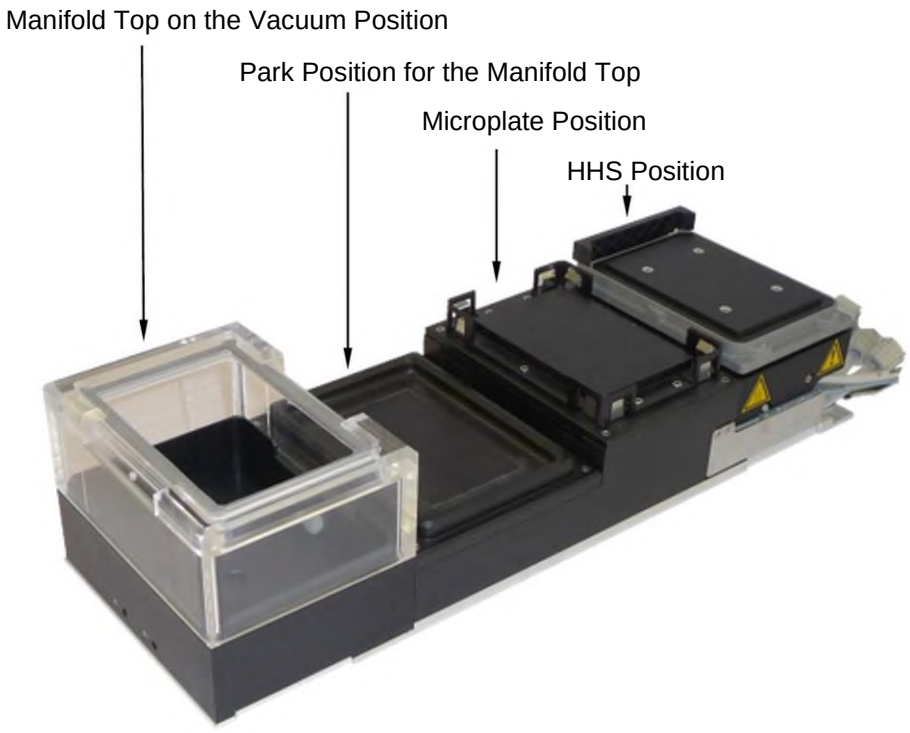
Vacuubrand Pump

Controller CVC 3000

The air-bleed valve and the pressure sensor are both mounted inside the CVS carrier.

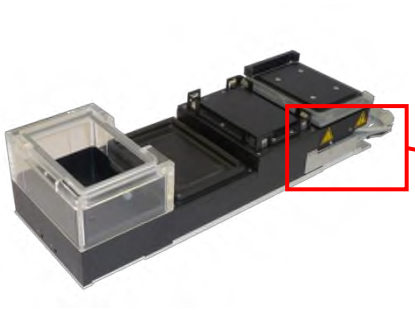
2.4.12.3 Shaker on the CVS

Optionally, the rear microplate position can be replaced by a HAMILTON Heater Shaker or by third party heater shakers. The list of available modules can be found in Section 8.1 Appendix A: Ordering Information.

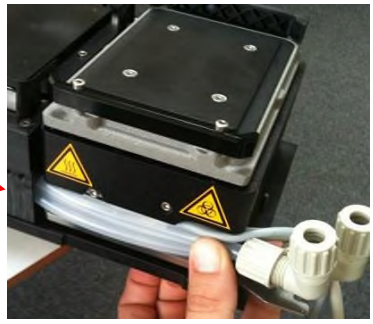


HAMILTON Heater Shaker on the CVS

The HAMILTON Heater Shaker will be installed by a HAMILTON Trained Field Service Engineer. The CVS Carrier comes with a metal plate for fixation for the heater shaker cables and tubing, it must be retained in a safe place, including screws for mounting, for a later retrofit of a heater shaker.



Position of the metal plate on the CVS Carrier for fixation of heater shaker cables and tubing



HHS, tubing and cables, shown before fixation by the metal plate

2.4.12.4 Waste Bottle for BVS / CVS

The CVS has to be connected either to a suitable in-house liquid disposal system or to a HAMILTON waste bottle which collects liquid that is extracted during the vacuum step. The waste bottle is connected to the vacuum chamber and to the pump unit. Waste bottles come assembled with a bucket which prevents spillage in case of leakage.

BVS / CVS Waste Bottle 4L in a Bucket

NOTE

When using alternative Waste systems, be aware that bottles and tubing's may be exposed to a differential pressure of approximately -1 bar.

The BVS / CVS waste bottles are not part of a CVS, they have to be ordered separately. Please refer to Section 8.1 Appendix A: Ordering Information. One 4 liter waste bottle is available. The waste bottle shall be placed below the instrument.

2.4.13 SCAP (Star Clean Air Protection)

The ML STAR instruments can be retrofitted with SCAP, a HEPA (*H*igh *E*fficiency *P*articulate *A*ir) Filter Hood for air filtration. The SCAP creates a laminar flow of filtered air from the top of the instrument to the deck surface. Room air is drawn in from the top and forced through a one stage filter. The air is discharged from the filters over the pipetting area inside the instrument and exits through the opening in front and vents in the acrylic glass cover on the back of the workstation to protect the assay from the environment outside the workstation.

Form fitting hoods are available for all three sizes of the STAR.

NOTE

The HAMILTON SCAP is designed to draw ambient air in through the filter assembly and to provide an enclosed, 'clean air zone' within the ML STAR instrument workstation pipetting environment. For more information about the SCAP, please refer to the SCAP Filter Hood Manual.

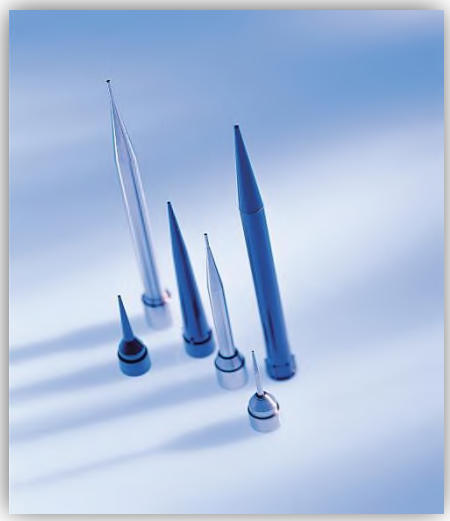
A UV light option is also available to further reduce germ count inside the instrument (e.g. before and after a working day). When retro-fitting the UV option, the instrument covers shall be exchanged from those made of standard acrylic glass to a UV resistant acrylic glass.

ATTENTION

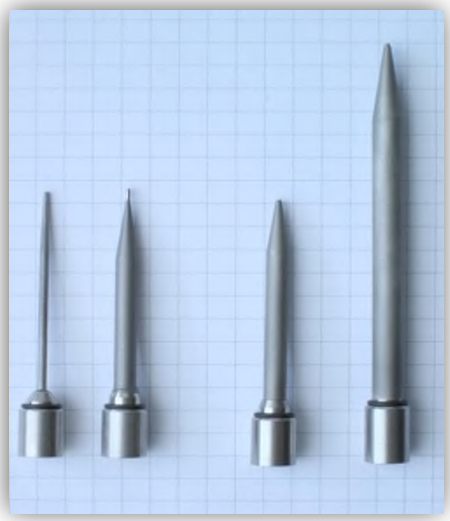
Do not expose yourself to the UV light(s), do not stare into the UV light source.

2.5 Tips and Needles

HAMILTON offers a large variety of disposable tips and steel needles for use with the pipetting channels and MPH heads.



Disposable Tips
Left to right: Low Volume (10 µl), 50 µl, Standard Volume (300 µl), and High Volume (1000 µl)



Steel Needles
10 µl, 300 µl, 1000 µl

HAMILTON Disposable Tips and Steel Needles for the ML STAR

HAMILTON´s CO-RE tips are the only tips designed to work with the ML STAR instruments developed to meet the demands of the most flexible and reliable automated pipetting tasks. The pipetting channels lock into the tip rather than the standard method of forcing a tip on a mandrel (see Section 2.1.6 Tip Pick-up with the CO-RE Technology for the CO-RE technology). This design assures superior tip alignment, seals the tip to the pipette channel, eliminates tip distortion, and mitigates aerosol contamination.

The use of non-HAMILTON tips is not supported on the ML STAR instruments. Non-HAMILTON tips do not have the annular space to receive the O-ring. This may result in non-intended deformation of the O-ring and the tip material itself. It may lead to misalignment of the tip, improper sealing, reduced O-ring life, improper drop-off, and potential aborted runs from randomly dropped tips. Therefore, HAMILTON does not support the use of non-HAMILTON tips on the ML STAR instruments.

ATTENTION

Only HAMILTON disposable tips and needles shall be used for coupling to the pipetting channels and pipetting heads of the ML STAR instrument. Other tips may cause contaminated or damaged pipetting channels.

NOTE

Pipetting specifications are only guaranteed when using HAMILTON Tips.

HAMILTON´s proprietary technologies for tip attachment (CO-RE) and air displacement pipetting together with the use of HAMILTON tips facilitate maximum pipetting accuracy and reliability.

HAMILTON disposable CO-RE tips are produced under clean room conditions (class 8), based on ISO 14644 standards. "Biological purity tested" tips are free of DNA, DNase/RNase, PCR inhibitors and endotoxin (non-pyrogenic). In addition to these criteria, "Biological purity^{PLUS}" tips are sterile, according to ISO 11135, and free of ATP.

Filter tips are available for preventing aerosol contamination.

Clear tips are available as well as conductive (black) tips specifically designed for cLLD.

NOTE

Make sure to match the tip type(s) used with the method and channel(s) / probe head(s). Please refer to Section 2.1.7 Tip Recognition.

NOTE

Be aware that different tip classes (with and without filter, black or clear, non-sterile versus irradiated, etc.) can have different performance when pipetting liquids. Pipetting parameters and liquid class settings may have to be adjusted when changing from one tip type to another in order to achieve accuracy and precision which is within published specifications.

NOTE

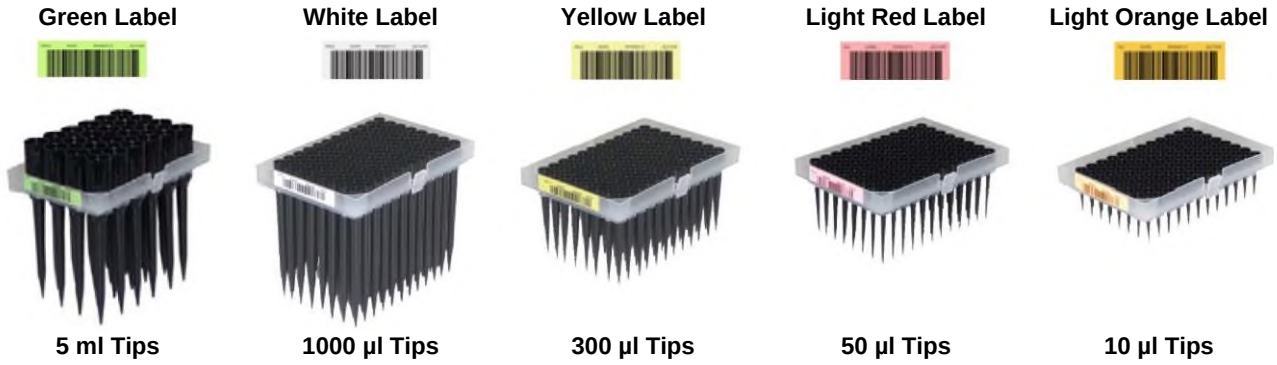
All new or special tip types require additional settings (such as configuration file entries, liquid classes, etc.) in the VENUS Software. Please consult a local HAMILTON representative for implementation of non-standard tips.

For ordering information see Section 8.1 Appendix A: Ordering Information, or contact a local HAMILTON representative.

2.5.1 Tip Packaging

All CO-RE disposable tip types are available in racks with a sealed paper lid (blister pack). The tip racks are barcode labeled for automatic identification during the loading process. All instruments equipped with a barcode reader are able to check for the proper tip type being loaded.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.



Examples of Barcode Labeled Tip Racks

NOTE

The tip type is printed in plain text on the barcode label of the tip rack for visual identification (e.g. "5 ml" for the 5 ml CO-RE tips).

Generally, blister packs contain several racks of 96 tips each. 5 ml tips come in blister packs containing racks of 24 tips each.

CO-RE tips are available in boxes of 3840 high-volume tips (1000 µl), 5760 standard-volume tips (300 µl and 50 µl) and 5760 low-volume tips (10 µl).

The racks are compatible with the respective tip carriers.

Tip Carrier TIP_CAR_480 containing five Tip Racks

10 µl, 50 µl and 300 µl CO-RE disposable tips without filters are stackable. These tip types are available in Nested Tip Racks (NTR). This high packing density and high number of tips make longer runs possible without reloading.

Nested Tip Racks (NTR's)

Five stacks of four NTR's each

The Nested Tip Rack (NTR) is designed to stack disposable tip racks which increase the amount of disposable tips by a factor of 4 per SBS position compared to one layer tip locations. NTR's come with 5 stacks of 4 layers each and have to be placed onto NTR carriers of the ML STAR instrument.

A transport system (e.g. iSWAP, CO-RE Gripper) is required to remove and dispose empty NTR frames away from the NTR Stacks to allow access with the Pipetting Tools to the next filled NTR.

Forcing Pipetting Tools to go down lower or to the lowest layer of the NTR stack may have limitations due to the required space between the Pipetting Tool and neighboring Carriers, as well as its corresponding labware (also to other stacks, neighbor NTR's, etc.). In these cases, an intermediate Tip pickup position becomes necessary. This intermediate tip pickup position is designed and available as an NTR Multiflex Module (MULTIFLEX NTR 96 MODULE, and MULTIFLEX NTR 384 MODULE).

The transport system, as mentioned above, must then be used to move NTR's to these intermediate tip pickup positions, which enables constant tip pickup conditions in terms of the same X, Y and Z coordinates of a single NTR. Once the tips out of the NTR frame of the intermediate tip pickup position are used up, the frame needs to be removed and replaced by a new filled NTR.

When single, row or column tip pickup is required with Multi Probe Heads, an additional tip pickup position equipped with a tip adapter is necessary. The tip adapter may be positioned onto a tip rack carrier as well as on a Multiflex tip module (MULTIFLEX TIP MODULE).

The recommended workflow would be:

1. Transportation of one NTR from a loaded NTR Carrier to the intermediate Tip pickup position.
2. Tip pickup with MPH out of the intermediate tip pickup position.
3. Tip eject into the tip adapter.
4. As desired; single, row or column (shifted) tip pickup out of the tip adapter.
5. Ready for pipetting.

2.5.2 Tips for Pipetting Low Volumes from 0.5 µl – 10 µl

The low volume CO-RE tip, 0.5 µl – 10 µl, is available as a conductive (black) tip for use with cLLD and as a clear tip. It can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Clear and Conductive Low Volume CO-RE Tip,
0.5 µl – 10 µl

Light Orange Label

Tip Rack and Color Code Label
for Low Volume CO-RE Tips, 0.5 µl – 10 µl

Low volume CO-RE tips, 0.5 µl – 10 µl come in racks of 96 tips. One blister pack contains 5 racks. They are also available in the NTR Tip Rack Pack reload system with five stacks of four 96 tip racks in a blister pack.

2.5.3 Tips for Pipetting Sample Volumes from 0.5 µl -50 µl

The low volume CO-RE tip, 0.5 µl – 50 µl, is available as a conductive (black) tip for use with cLLD, with or without filter and also as a clear tip. It can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Clear and conductive Low Volume CO-RE Tip,
0.5 µl – 50 µl

Light Red Label

Tip Rack and Color Code Label
for Low Volume CO-RE Tips, 0.5 µl – 50 µl

Low volume CO-RE tips, 0.5 µl – 50 µl come in racks of 96 tips. One blister pack contains 5 racks. They are also available in the NTR Tip Rack Pack reload system with five stacks of four 96 tip racks in a blister pack.

ATTENTION

Caution when using the CO-RE tips,0.5 µl – 50 µl and CO-RE tips,10 µl – 300 µl on the same system. Do not mix up one tip type with the other during the process of loading the deck. The wrong tip causes either bad pipetting results or damaged pipetting channels.

Use the Autoload function for loading tip carriers or the tip type recognition feature of the VENUS Software. Please consult the local HAMILTON representative for the specific library which is needed.

2.5.4 Tips for Pipetting Sample Volumes from 0.5 µl – 50 µl with 384 MPH

The 384 head CO-RE tip, 0.5 µl – 50 µl, is available as a conductive (black) tip for use with cLLD and also as a clear tip. It can be used with the CO-RE 384 Probe Head.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Clear and Conductive 384 Probe Head CO-RE Tip, 0.5 µl – 50 µl

NTR with 384 (left) and 96 (right) 384 Probe Head CO-RE Tips, 0.5 µl – 50 µl

384 head CO-RE tips, 0.5 µl – 50 µl come in NTRs of 384 or 96 tips per rack. The NTR's equipped with 96 tips per rack enable using the CO RE 384-Probe Head as a 96 channel pipetting head. The NTR pack reload system contains five stacks of four tip racks in a blister pack.

NTR pack containing five stacks of four 384 head CO-RE tip racks

2.5.5 Tips for Pipetting Sample Volumes from 10 µl – 300 µl

The CO-RE tip, 10 µl – 300 µl, is available as a conductive (black) tip for use with cLLD, with or without filter and also as a clear tip. It can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

It is compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Clear and Conductive CO-RE Tip, 10 µl – 300 µl

Yellow Label

Tip Rack and Color Code Label for CO-RE Tips, 10 µl – 300 µl

CO-RE tips, 10 µl – 300 µl come in racks of 96 tips. One blister pack contains 5 racks. They are also available in the NTR Tip Rack Pack reload system with five stacks of four 96 tip racks in a blister pack.

ATTENTION

Caution when using the CO-RE tips,1-50µl and CO-RE tips,10-300µl on the same system. Do not mix up one tip type with the other during the process of loading the deck. The wrong tip causes either bad pipetting results or damaged pipetting channels.

Use the Autoload function for loading tip carriers or the tip type recognition feature of the VENUS Software. Please consult the local HAMILTON representative for the specific library which is needed.

2.5.6 Tips for Pipetting Sample Volumes from 10 µl – 1000 µl

The CO-RE tip, 10 µl – 1000 µl, is available as a conductive (black) tip for use with cLLD, with or without filter. It can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

It is compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Conductive CO-RE Tip, 10 µl – 1000 µl

White Label

Tip Rack and Color Code Label for CO-RE Tips, 10 µl – 1000 µl

CO-RE tips, 10 µl – 1000 µl come in racks of 96 tips. One blister pack contains 5 racks.

ATTENTION

Caution when using the CO-RE tips, 10 µl – 1000 µl and Slim tips, 10 µl – 300 µl on the same system. Do not mix up one tip type with the other during the process of loading the deck. The wrong tip causes either bad pipetting results or damaged pipetting channels.

Use the Autoload function for loading tip carriers or take extra care and identify the tip type by reading the label on the barcode when loading the tip carriers manually.

2.5.7 Tips for Pipetting Sample Volumes from 50 µl – 5000 µl

The CO-RE tip, 50 µl – 5000 µl, is available as a conductive (black) tip with or without filter. It can be used with 5 ml pipetting channels.

It is compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Conductive CO-RE tip, 50 µl – 5000 µl

Green Label

Tip rack and color code label for CO-RE tips, 50 µl – 5000 µl

CO-RE tips, 50 µl – 5000 µl come in racks of 24 tips. One blister pack contains 5 racks. In addition, individually wrapped, preloaded 1T disposable racks of 4 tips each are available in which the tips are physically separated by individual sheaths. They ensure contamination free tip loading.

2.5.8 Special Tips: Rocket Tip

The unique Rocket tip, 2 µl – 300 µl, fits to four pipetting channels of a CO-RE 384 Probe Head in parallel and turns it into a 96 pipetting head on the fly. It is available as a conductive (black) tip for use with cLLD. Rocket tips can be used with Deep Well Plates and microplates with well diameters ≥ 3.7 mm.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Rocket tip, 2 µl – 300 µl

Tip rack and color code label for Rocket tips, 2 µl – 300 µl

Rocket Tips, 2 µl – 300 µl are packaged in racks of 96 tips per rack. One blister pack contains 5 racks.

2.5.9 Special Tips: Piercing Tip

The robotic piercing tip, 2 µl – 250µl is optimized for use with various sealed labware which can be pierced by applying a force of ≤ 10 N. The special tip design is compatible with most commercially available seals.

The tip is available as a conductive (black) tip for use with cLLD, with or without filter. It is compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Piercing tip, 2 µl – 250 µl

Yellow Label

Tip rack and color code label for piercing tips, 2 µl – 250 µl

Piercing tips, 2 µl – 250 µl come with 96 tips per rack. One blister pack contains 5 racks.

For piercing of tubes, additional equipment is required to hold the tubes in their position during the piercing process and to prevent them from being lifted when retracting the piercing tip. The fixation frame for this purpose is dependent on the labware used. It is mounted onto a MTP park position (PN 188041) on a Multiflex carrier (PN 188039). The standard fixation frame (PN 188313) is designed for 1.4 ml matrix tubes. For all other tube types, a customized fixation frame is required.

For piercing of plates, the Multiflex MTP fixation frame (PN 188095) helps to lock the sealed plates in position on a Multiflex carrier during piercing.

Fixation frame for 1.4 ml matrix tubes on a MTP park position (PN 188313)

Multiflex MTP fixation frame on a MTP park position (PN 188095)

Multiflex Carrier (PN 188039)

NOTE

In general, piercing is a challenge. It requires a fixation frame for the labware to be pierced and it is highly dependent on the shape of the needle or tip and the material and thickness of the seal. It is crucial that any piercing method is tested first before routine application.

2.5.10 Special Tips: Extra-long and Slim Tip

Extra-long and slim tips minimize the risk of liquid overflow in cluster tubes, Deep Well Plates and two-phased liquid applications. The Slim tip, 10 µl – 300 µl reduces the displaced liquid volume by 40%, compared to a CO-RE tip, 10 µl – 1000 µl. The tip is available as a conductive (black) tip for use with cLLD, with or without filter. It can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

The Slim tip is compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Slim Tip, 10 µl – 300 µl

Tip Rack and Color Code Label for Slim Tip, 10 µl – 300 µl

Slim tips, 10 µl – 300 µl come with 96 tips per rack. One blister pack contains 5 racks.

ATTENTION

Caution when using the CO-RE tips,10 µl – 1000 µl and Slim tips, 10 µl – 300 µl on the same system. Do not mix up one tip type with the other during the process of loading the deck. The wrong tip causes either bad pipetting results or damaged pipetting channels.

Use the Autoload function for loading tip carriers or take extra care and identify the tip type by reading the label on the barcode when loading the tip carriers manually.

2.5.11 Special Tips: Wide Bore Tips

For applications which require minimizing shear forces applied to the samples during pipetting, for highly viscous fluids or for particulate samples, HAMILTON has designed CO-RE tips with various wide distal end orifices. The CO-RE tip wide bore 10 µl – 300 µl with a length of 57.23 mm which orifices size 0.71 mm is a filter tip. The CO-RE tip wide bore 10 µl – 300 µl with a length of 51.60 mm which orifices size 1.55 mm and the CO-RE tip wide bore 10 µl – 1000 µl with a length of 80.14 mm which orifices size 3.2 mm are available with or without filter. Wide bore tips are conductive (black) for use with cLLD and they can be used with 1000 µl pipetting channels and CO-RE 96 Probe Head / CO-RE 96 Probe Head TADM.

Wide bore tips are compatible with Monitored Air Displacement (MAD) and Total Aspiration and Dispense Monitoring (TADM).

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

Yellow Label

Wide bore tip, 10 µl – 300 µl, orifice size 0.71 mm

Yellow Label

Wide bore tip, 10 µl – 300 µl, orifice size 1.55 mm

White Label

Wide bore tip, 10 µl – 1000 µl, orifice size 3.2 mm

Wide bore tips come with 96 tips per rack. One blister pack contains 5 racks.

2.5.12 Needles

Steel needles can be used for pipetting instead of disposable tips in combination with a needle wash station when the application does not require an exchange of a tip. Needles are rinsed between pipetting tasks and are resistant to many chemicals (refer to Section 8.2 Appendix B: Chemical Compatibility).

10 µl, 300 µl and 1000 µl needles are available for use with 1000 µl pipetting channels.

See Section 8.1 Appendix A: Ordering Information for a complete listing, or contact a local HAMILTON representative.

NOTE

HAMILTON recommends replacing steel needles every six months.

2.6 Computer Requirements

The ML STAR instrument is controlled by a dedicated VENUS Software which controls all functions for daily work routine, method programming, running methods, and other services.

HAMILTON offers standard computers for the VENUS Software. For specifications, please refer to Section 7.11 HAMILTON Standard PC Specifications. For ordering information, please refer to Section 8.1 Appendix A: Ordering Information.

NOTE

Computer requirements as well as operating systems described in this manual are subject to change without notice, respectively may become obsolete.

Depending on the ML STAR instrument configuration and processor board implemented, the following interfaces are available for linking the control computer (see Section 2.8 Electronics and Interfaces):

- a Serial Interface (RS-232C, with the Dual Processor Board)
- a Unified Serial Bus Interface (USB, with the Dual Processor Board or the LAN Dual Processor Board)
- Ethernet (with the LAN Dual Processor Board)

The communication interface used on the PC has to be set by the configuration editor. For further information about the recommended PC model, refer to Section 7.1 Basic ML STAR Specifications.

To avoid data loss, using of an Uninterruptible Power Supply (UPS) for the PC is recommended.

2.7 VENUS Software

The VENUS Software provides everything to control an ML STAR instrument.

It is a Windows-based, menu-driven interface allowing the user to define deck layouts and methods, and then to run the instrument.

The VENUS Software allows programming and running different methods for aspirating and dispensing liquids, also to control accessories such as a wash station, etc.

It is possible to upgrade from previous software versions to VENUS four. A software upgrade requires compatible firmware.

ATTENTION

ML STAR functionality has been verified using Windows 7 and Windows 10. Running the instruments under any other operating system may lead to severe problems and/or malfunction. Each programmed method shall be validated by the customer.

NOTE

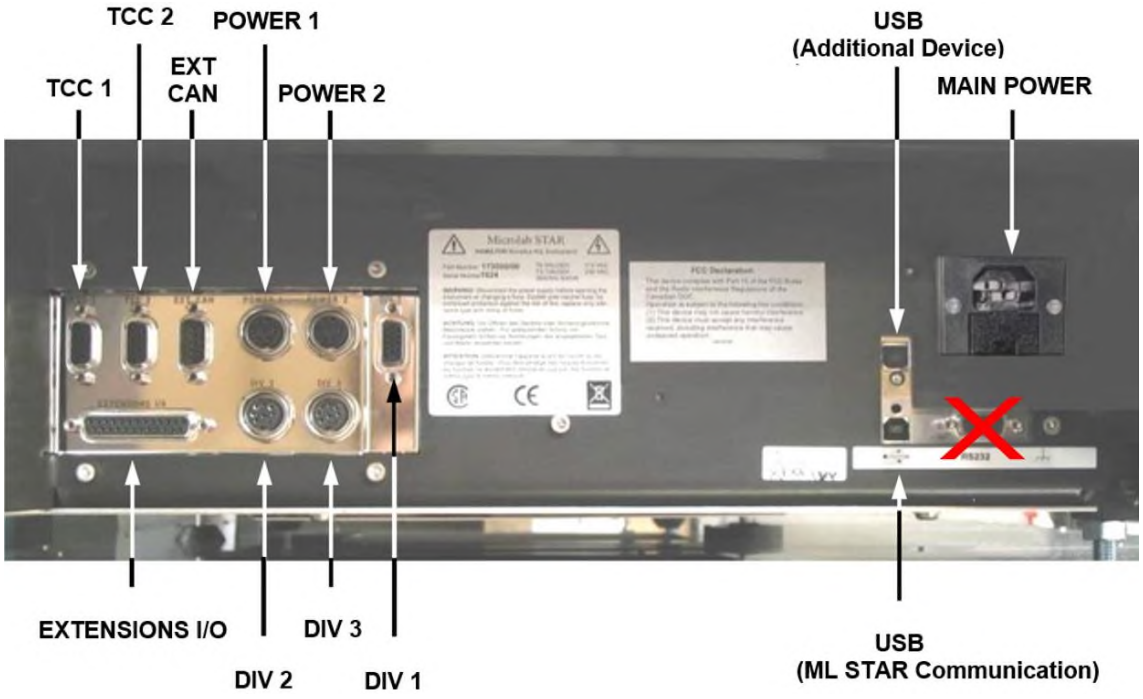
Windows 7 is no longer supported by Microsoft.

For more details refer to the VENUS Software Programmer's Manual. If not using a HAMILTON computer, the customer needs to ensure that a suitable control PC is available for installation of the VENUS Software. The ML STAR instrument will be unpacked and installed and the initial set-up will be performed by a HAMILTON Trained Field Service Engineer according to the IQ Form 610911.

2.8 Electronics and Interfaces

All the electrical connections are placed on the left side of the instrument, as shown below.

2.8.1 Interfaces of the Dual Processor Board



Interfaces of the Dual Processor Board

The main power connection is located near the front of the instrument (on the right side in the picture above). The communication connections to the PC are positioned beneath this main power connection. The ML STAR communicates via the USB. A second USB connection is reserved for an additional device (e.g. an optional CCD Camera).

ATTENTION

The RS232 is no longer supported for communication. The connector is still available, however, without communication.

ATTENTION

Do not attempt to install a 3rd party device via the electrical connections of the ML STAR instrument on your own! Always consult a local HAMILTON representative for installation.

Various connections for external devices are available. Up to two Temperature Controlled Carriers (TCC's) or HAMILTON Heater Shakers can communicate and be powered via the connectors labeled "TCC 1", "TCC 2".

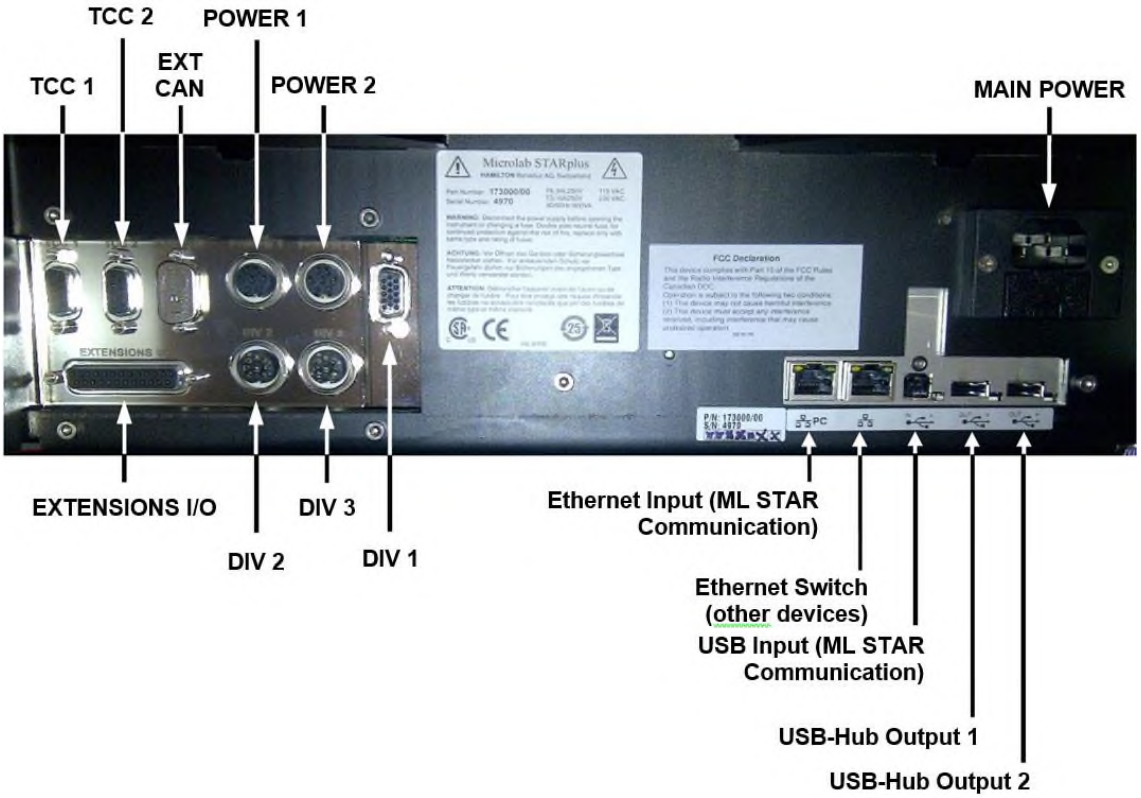
The CR Needle Wash Station is connected with the "EXT CAN" connector.

The "POWER 1" and "POWER 2" connectors provide different power supply voltages and also a CAN bus for communication.

The "DIV 1", "DIV 2", "DIV 3" connectors, and the "EXTENSIONS I/O" connector deliver several digital input/output signals as well as pulse-width-modulated (PWM) outputs, CAN Bus and TTL levels.

2.8.2 Interfaces of the LAN Dual Processor Board

Depending on the configuration, some ML STAR instruments are configured with a LAN Dual Processor Board (e.g. in combination with a CO-RE 96 Probe Head TADM). The electrical connections of these instruments are as shown below:



Interfaces for the LAN Dual Processor Board

NOTE

With the LAN Dual Processor Board installed, an RS232 connection is no longer provided.

For further details on all connections, please consult a local HAMILTON representative.

2.9 ML STAR Power / Voltage

600 VA

Make sure that the instrument is connected to a 100 or 115 or 230 VAC (50 Hz or 60 Hz) socket. The ML STAR automatically recognizes any voltage within that range, without user intervention.

1000VA

Make sure that the instrument is connected to a 115 or 230 VAC (50 Hz or 60 Hz) socket. The ML STAR automatically recognizes any voltage within that range, without user intervention.

General

We recommend using an Uninterruptible Power Supply (UPS) for the ML STAR instrument.

Ensure that the instrument is correctly grounded when connected to the electrical outlet.

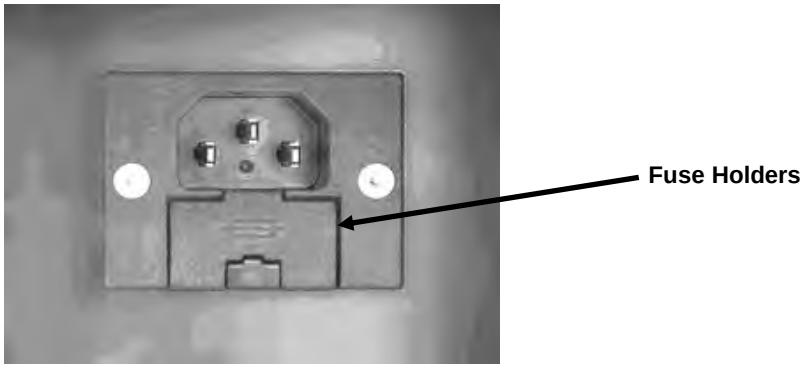
The main plug is on the left-hand front side of the instrument.

The fuses for the instrument are placed in the main power socket (see picture below). During installation / IQ, the appropriate fuse is selected by a HAMILTON Trained Field Service Engineer.

Plug the main cables for the computer and the instrument into the same electrical outlet. Connect them only to a grounded outlet.

ATTENTION

Do not exchange the supplied/installed power cord. In case of damage or loss of the power cord, consult a local HAMILTON representative for a replacement. If replacing the power cord by inadequately rated power cords, the protection provided by the equipment may be impaired.



Fuses

ATTENTION

When replacing a fuse, make sure to use the appropriate fuse (see Section 7.1 Basic ML STAR Specifications) and place it into the main power switch before switching on the instrument.

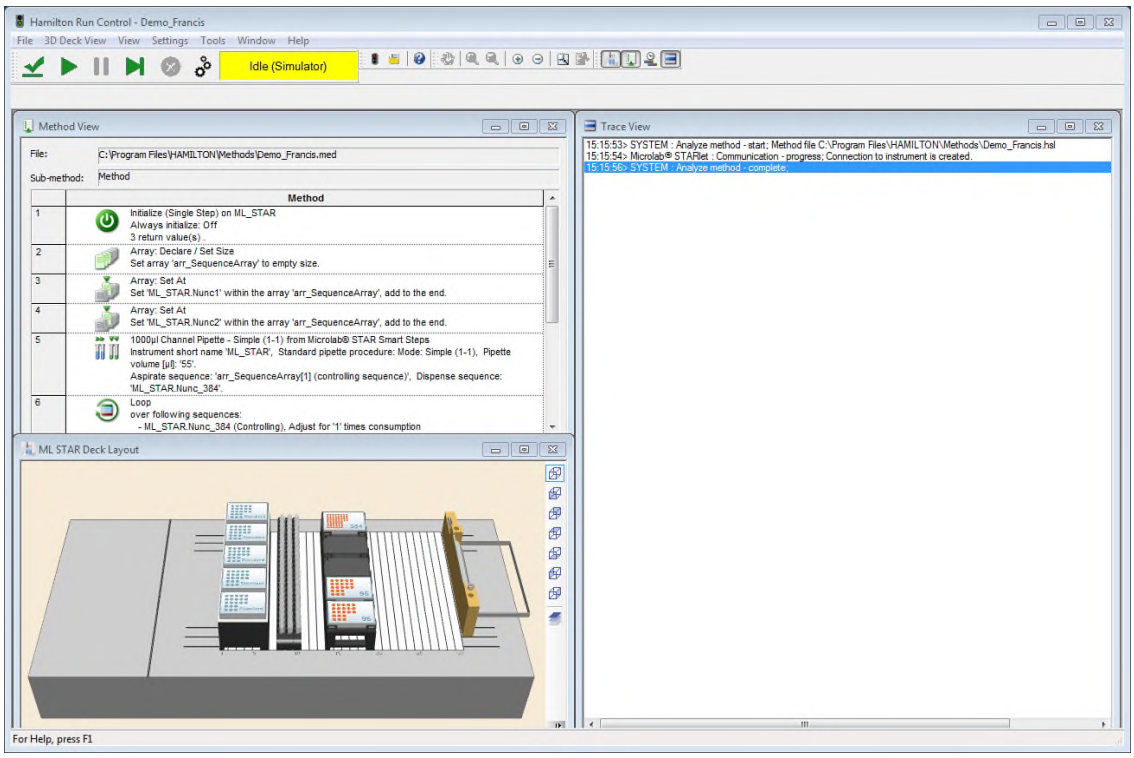
3. Routine Use

3.1 Loading the ML STAR Instrument

To run a method:

1. Double-click the "**Microlab STAR Run**" Icon on the desktop:

2. From the File menu of Run Control, select "Open", to open a method (*.med). The method is now loaded:



3. The free area visible after starting "Run Control" can display multiple windows:
   - Deck View
   - Method View
   - Trace View
   - Schedule View
   - HSL Debugger (if installed)
   - Watch View (if HSL Debugger is installed)

4. The windows may be enabled/disabled by means of the "Run Views" menu (click the desired menu entry)

   or

   by clicking on the appropriate icon in the "Run Views" toolbar  (if enabled).

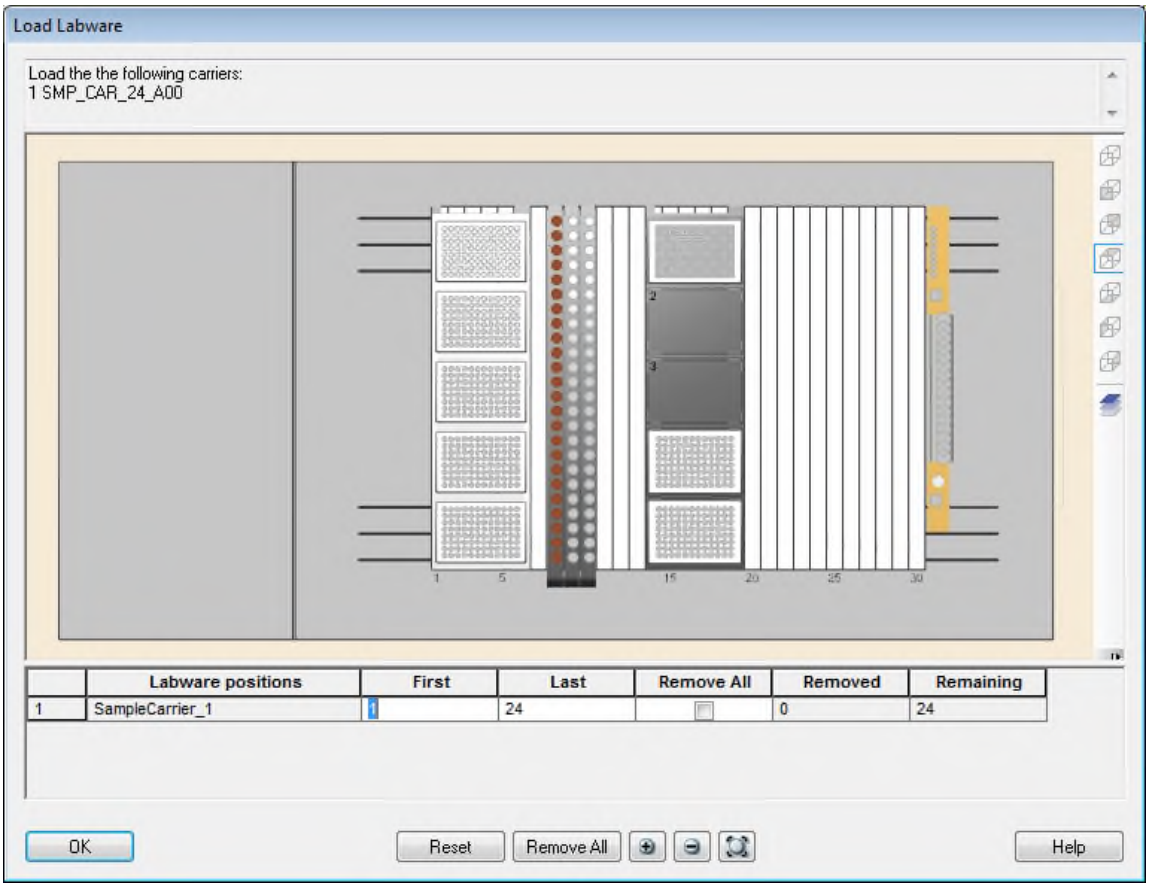
5. Arrange the windows according to preference using the "Window" menu. In the window shown above, the deck layout and the method log (trace) of a run as it is generated are shown.

6. Press the [Start] button (the one which looks like a [Play] button)

   to run the method. The steps in the method are traced to the log frame.

7. If a loading step is part of the method, a dialog appears that requests a reduction of the number of positions (e.g. on the sample carrier), as well as a start position for the tips to be picked up. Both pieces of information are optional.



Loading Dialog

8. Enter the number of samples, tips or wells for this run, delete items graphically from the highlighted positions on the deck (first select the appropriate item in the table), or accept the default.

9. Generally, whenever the system finds a 'Load Carrier' command in the method, the operator is requested to place the carrier holding the appropriate labware onto the Loading Tray. The correct position is highlighted by blinking LED's.

ATTENTION

It is important to ensure that adequate amounts of the correct liquids are placed in the correctly labeled containers.

When pouring reagents into the reagent containers, ensure that there is no foam on the surface of the liquid.

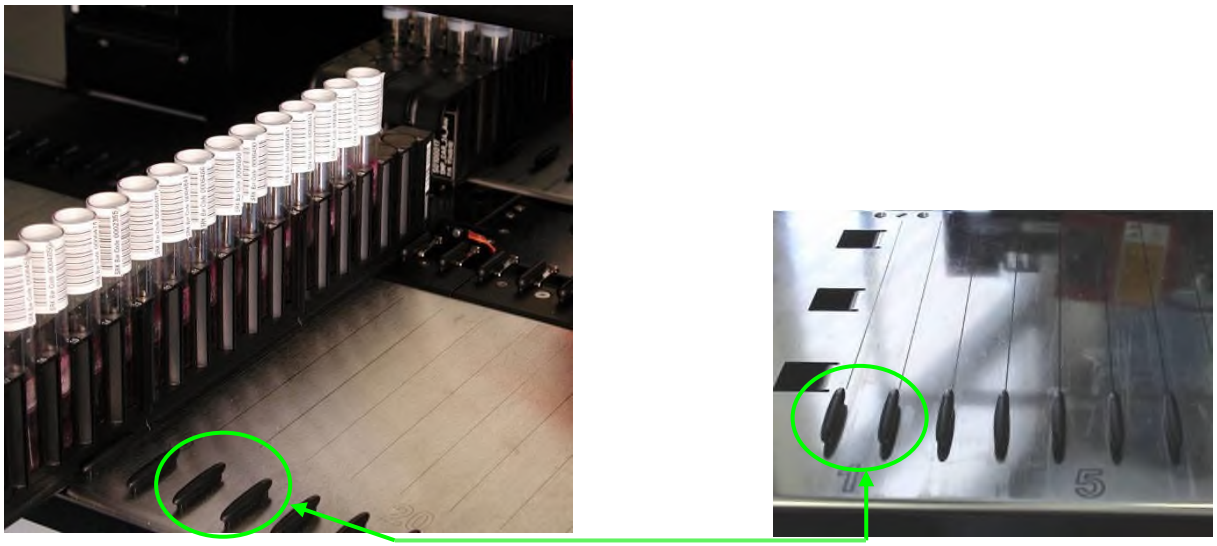
Do not overfill reagent containers: fill approximately10 mm below the top of the container.

Always use the proper labware (tips, microplates, tubes, etc.) corresponding to the definitions of the method's layout.

Position microplates correctly such that well #A1 is placed according to the deck layout.

Handle any 1-track carrier (such as a sample carrier) with particular care, as this type of carrier can fall over and cause injury or contamination. Position it onto the Loading Tray (see figure below), or place several carriers together to minimize this risk.

10. Insert the carriers into the tracks between the front and rear slide blocks of the Loading Tray until they touch the stop hooks on the far side of the tray.



Slide Blocks for Carriers

Stop Hooks for Carriers

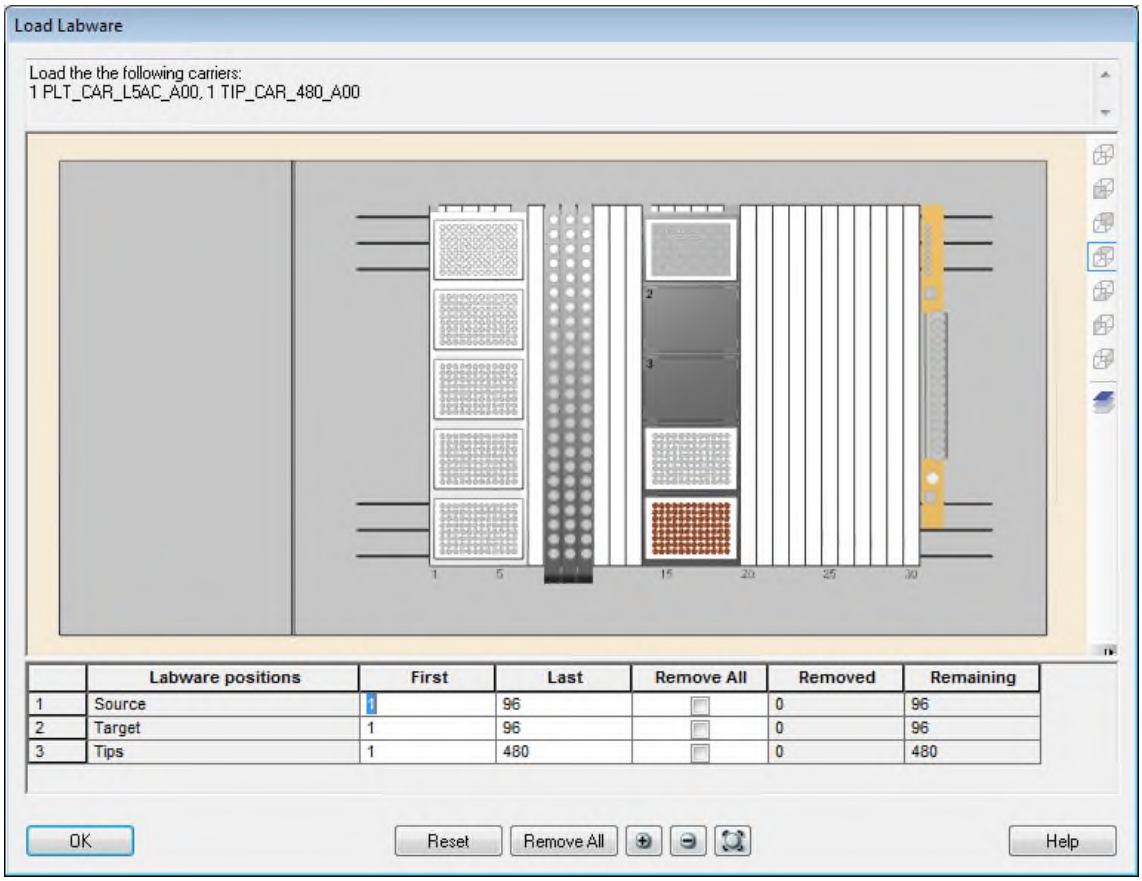
ATTENTION

Make sure the carriers are inserted completely as far as they will go on the loading tray.

11. Click **[OK]** in the dialog. The carriers are loaded onto the deck automatically by the 'Load Carrier' command in the method. During loading, the barcodes of the carriers and labware are read and stored in a file.

    or

    Alternatively, load the carriers onto the defined positions of the Loading Tray before starting the method. Loading and barcode reading will then be performed without operator input. Still the following dialog is displayed:



12. Enter the number of wells (e.g. on the source plate for this run), delete wells graphically from the sequence or accept the default (copy the entire plate).

For a Manual Load System only:

13. Load the deck with the carriers mentioned in the upper part of the dialog box (the two plate carriers and two tip carriers). Don't forget to place labware onto the correct positions.

For an Autoload System only:

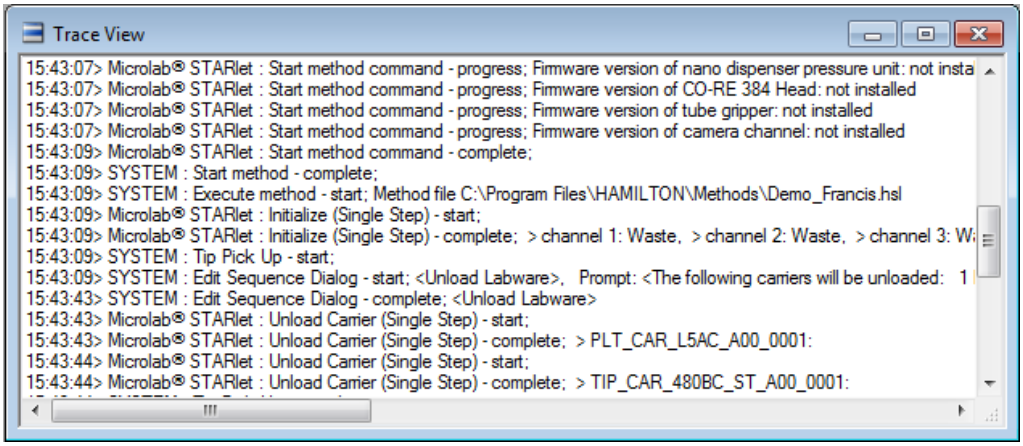
14. Whenever the system finds a "Load Carrier" command in the method, the user is requested to feed the carrier holding the appropriate labware onto the Loading Tray. The correct position is highlighted by LED's on the instrument. Alternatively, all carriers can be placed directly in their correct positions on the Loading Tray.
15. Click **[OK]** in the dialog box to start loading.

For a Manual Load System only:

16. Click **[OK]** in the dialog. Unload all the carriers manually. The unloading will be checked by the system.

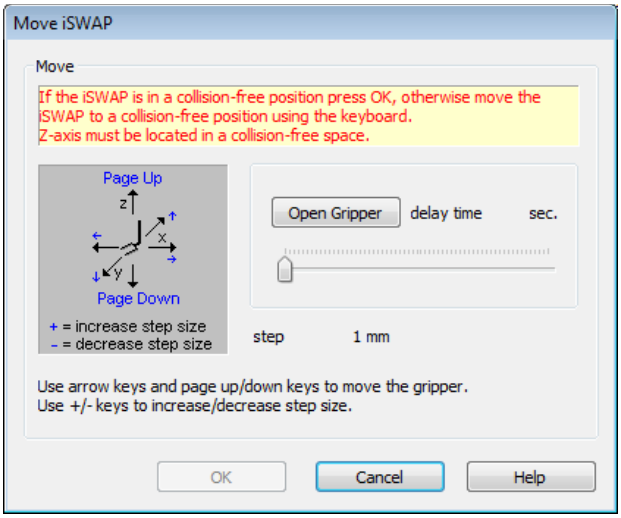
For an Autoload System only:

17. Click **[OK]** in the dialog. The carriers are unloaded to the Loading Tray.
18. The method is now finished. Method completion information is visible in the "Trace View" window.



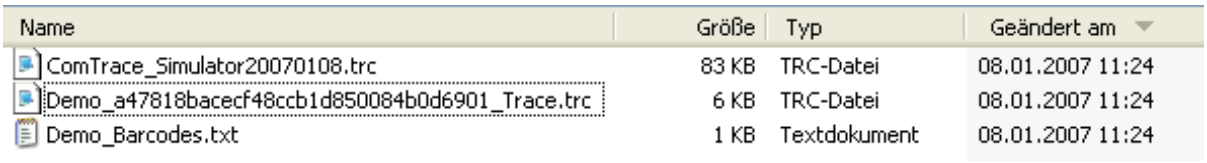
For the iSWAP only:

19. Usually, at the first run of a day, the iSWAP needs to be initialized. There is also a possibility that there is still a plate in the gripper of the iSWAP. In case a plate is caught by the gripper, it can be safely removed.



3.1.1 Method Trace File

Every run creates a separate method trace file. The method trace file is stored under "{method namex32x}.trc" within the "...\LogFiles" directory. Each method trace contains the method name, and a unique number consisting of 32 hexadecimal digits, here represented by "x32x", within the file name. The method traces are not overwritten or appended.



3.1.2 Communication Trace File

The communication trace file is created every day or when the instrument is switched on. A communication trace file is also generated under the "communication name" (Simulator, HxUSBComm) and current date, where the communication of each run of the method will be appended. If barcode information is generated, its data will be stored in a file labeled with the method's name followed by **"*_Barcodes.txt".**

NOTE

All unused method traces and com traces shall be deleted by the operator from the hard disk as the information will fill the hard disk. (backup on a separate device).

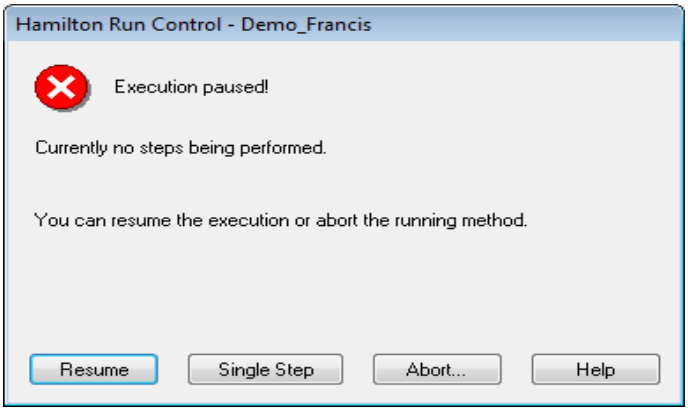
3.1.3 Single Step Mode

The method can be tested using Single Steps. It is always possible to start a run in Single Step mode (like TipPickUp, Aspirate, Dispense, etc.).

1. Click the **[Single Step]** button, to start the method in single step mode.

   or

   Pause a method at any time by clicking on the **[Pause]** button on the Run Control Window. The step currently running will be terminated, and the following message box appears:



2. Click anywhere in the pause dialog to stop the beeping.

   During the pause, it is possible to open the front cover of the instrument.

3. Before continuing the method, make sure the cover is closed again.

4. Resume the paused method by clicking **[Single Step]**

   or

   Resume method execution by clicking **[Resume]**, to run the method without further breaks

   or

   Abort method execution by clicking **[Abort...]**. It will be prompted to confirm the abort.

NOTE

An abort may cause loss of data.

Aborted methods cannot be restarted again, unless explicitly programmed.

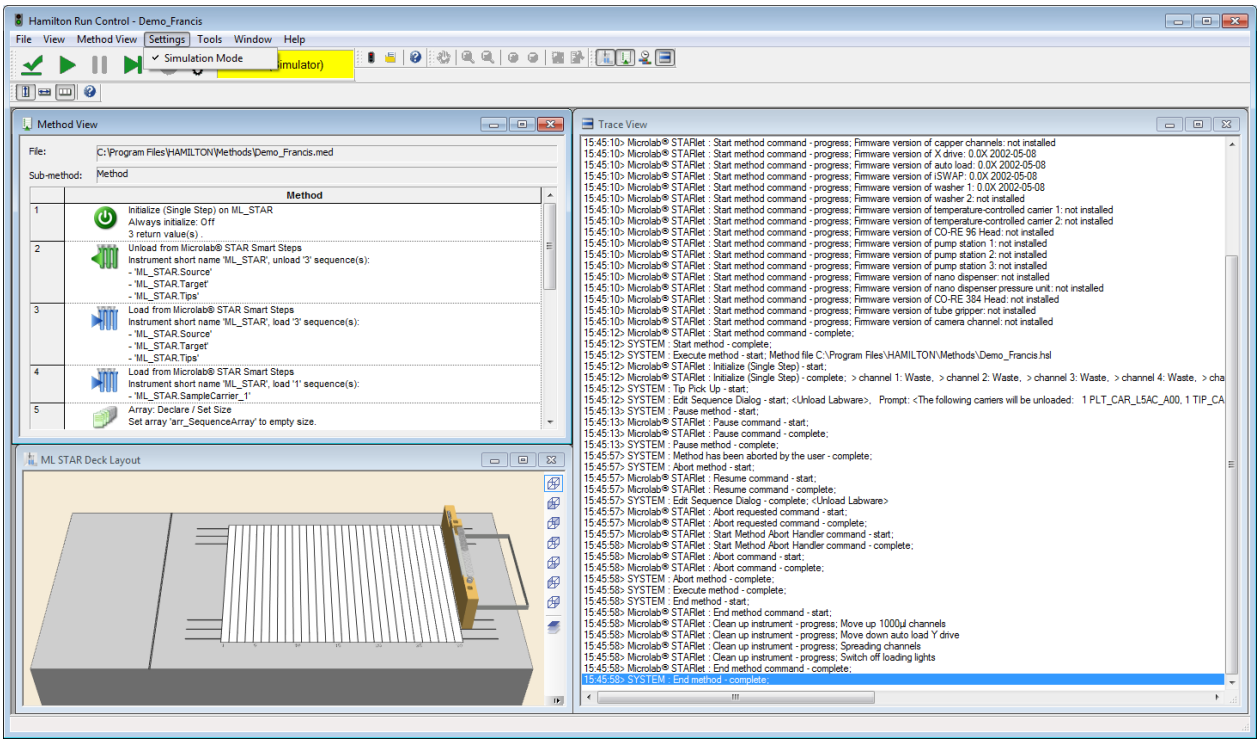
A fast abort can always be done by opening the front cover of the ML STAR instrument during a run.

*A method can be aborted even if **[Pause]** is active.*

*To prevent loss of data or bad pipetting results, **[Pause]** will finish the proceeding step and then stops.*

3.1.4 Run Simulations

It is also possible to run a simulation instead of the actual run. It is recommended to always simulate a newly created method first, before running it on the instrument. The run simulation is switched on in the menu "Settings":

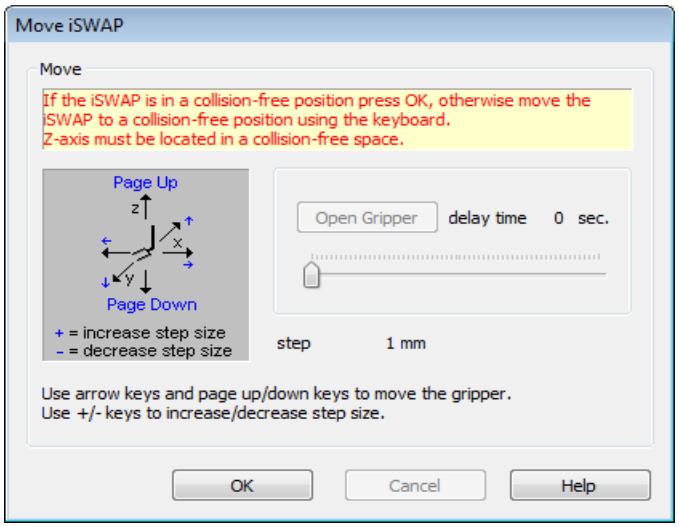


1. Enable the simulation by clicking to **[Settings / Simulation Mode]**.
2. The run simulators check the consistency of each (single) step of the system, together with minimum aspiration and dispense volume management.
3. The speed for simulations is adjustable. If the default speed (at maximum is best guess under real runs) is too fast the speed can be reduced by the simulator delay via the configuration editor. Refer to the VENUS Software Programmer's Manual.

3.2 Retrieving Objects

3.2.1 Retrieving Objects from the iSWAP

Usually, at the first run of a day, the iSWAP needs to be initialized. There is a possibility that there is still an object (e.g. plate) in the gripper of the iSWAP. In case an object is caught by the gripper, it can be removed safely using the "Move iSWAP" dialog.



Removing a gripped object:

1. Use the "+"/"-" keys to define the distance (e.g. 10 mm) the gripper shall be opened to release the object.
2. Define a "delay time" moving the pointer of the slide bar.
3. Calculate the time (e.g. 5 seconds) you will require to walk from the PC to the instrument and place your hand underneath the object.
4. Open up the front cover of the ML STAR instrument and check if you can retrieve the object.
5. If you cannot access the object move the X-Arm using the arrow keys and page up / page down keys.

ATTENTION

Make sure the selected step size and moving direction will not crash the iSWAP. Always wear gloves, eye protection and lab coat for manipulations inside the instrument.

ATTENTION

When using the iSWAP to transport plates, a plate carrier must be used as listed in this manual (see Section 8.1 Appendix A: Ordering Information). If other carriers are used, make sure that the plate position is verified.

6. If there is enough time and space to hold the object and the gripper parameters are set, click the **[Open Gripper]** Button.
7. If the iSWAP cannot move up to the traveling height without a collision (e.g. shelving position where the iSWAP would collide with the shelf above it), it must be guided to a safe area using the keyboard (arrow keys, page up / page down keys, + / - keys).

ATTENTION

Make sure the selected step size and moving direction will not crash the iSWAP.

8. If the iSWAP is ready for initialization, click the **[OK]** Button.

3.2.2 Retrieving objects from the Tube Twister Channel

Usually, at the first run of a day, the Tube Twister Channels need to be initialized. There is a possibility of objects (e.g. tube) still gripped by one or more Tube Twister Channels.

The situation may appear when a run was aborted either automatically by the system (in case of irregularities detected by the instrument) or when a manual abort was triggered by the operator.

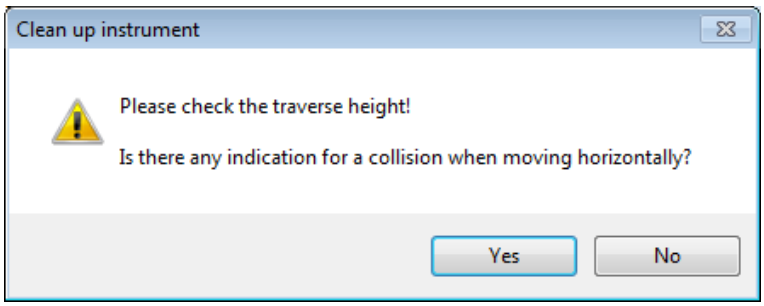
ATTENTION

Retrieving objects from the Tube Twister Channel requires completing steps within specified time frames. Read and understand the complete instructions given in this section before retrieving objects.

ATTENTION

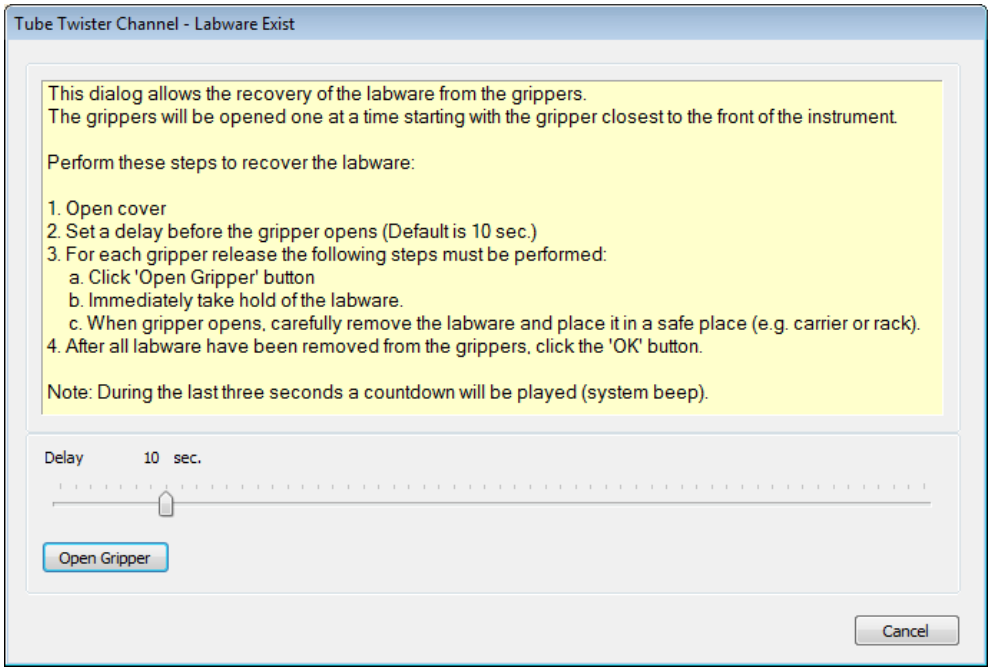
Always wear gloves, eye protection and lab coat for manipulations inside the instrument.

1. Depending on the status of the instrument, the following dialog may first appear.



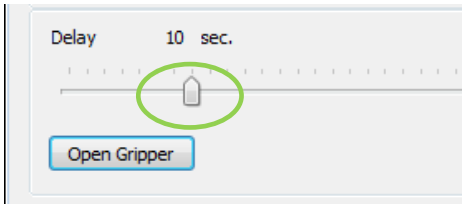
Check for the situation of the gripped object.

2. If for any reason, the Tube Twister Channel cannot move horizontally, press **[Yes]** and recover from the situation manually after shutdown of the instrument.
3. If there is no indication for a collision, press **[No]**.
4. In case one or more objects are caught by the grippers, it can now be removed safely using the **Tube Twister Channel – Labware Exist** Dialog.



Removing a gripped object:

5. Remove any carrier from the Loading Tray.
6. Open the front cover of the ML STAR Instrument and check if you can retrieve the gripped objects.
7. If you cannot access the object, do not open the gripper. Click **OK** and handle the situation manually after shutdown of the instrument.
8. If you can access the object, calculate the time (e.g. 10 seconds) you will require to walk from the PC to the instrument and to place your hand underneath the gripped object. Use the slider to set the required "**Delay Time**".



9. If there is adequate time set and space to hold the gripped objects click the **[Open Gripper]** Button. Immediately take hold of the object.

10. For the next object, click the **[Open Gripper]** Button. Immediately take hold of the object.
11. Repeat the step until all objects are released from the grippers.
12. Remove the objects from the deck.
13. Close the front cover.
14. Click the **[OK]** Button to proceed.

3.2.3 Retrieving objects from the Decapper Module

Usually, at the first run of a day, the Decapper Module needs to be initialized. There is a possibility of objects (tubes) still sitting in one or more Decapper Stations.

The situation may appear when a run was aborted either automatically by the system (in case of irregularities detected by the instrument) or when a manual abort was triggered by the operator.

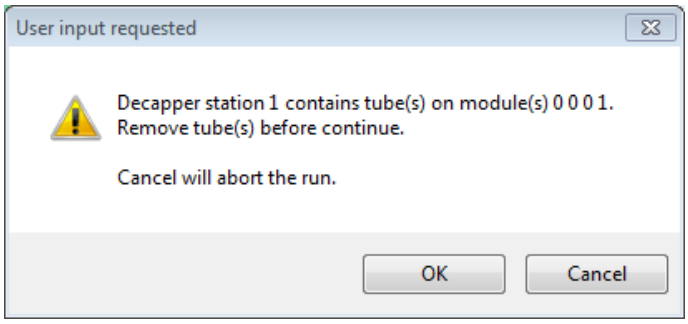
ATTENTION

Retrieving objects from the Decapper Modules requires special attention. Always use the specific pliers supplied with the Decapper Module to avoid pushing down the vial into the Decapper Module and damaging the locking springs. Take special care not to spill liquid into the Decapper Module.

ATTENTION

Always wear gloves, eye protection and lab coat for manipulations inside the instrument.

In case one or more objects are sticking in the Decapper, the following dialog will be displayed.



Removing an object:

1. Remove any carrier from the Loading Tray.
2. Open the front cover of the ML STAR instrument.
3. Insert the pliers into the open tube or into the depression of its cap and grip the object.

4. Carefully pull out the tube of the Decapper Station. Avoid spilling liquid. Take care not to deform the locking spring within the Decapper Station.
5. Close the front cover.
6. If all Decapper Stations are checked and clear, click the **[OK]** button to proceed.

3.3 Error Handling

3.3.1 Run-Time Error Handling

Before using the run time error handling, several types of problems causing errors have to be solved first. Among these are:

- Syntax errors when programming in HSL (e.g. forgotten ";")
- Logical errors (e.g. "tip eject" before "pick-up", or "asp 20 µl", "disp 100 µl")
- Semantic errors (e.g. wrong pipetting pattern)
- Method/deck interaction errors (e.g. dispense 100 µl into the first well of a 1536-well MTP)
- Liquid handling/application errors (e.g. droplets, foam, non-pipetted wells)
- User-related errors (e.g. wrong deck loading)

These problems *cannot* be handled by any run time error handling. They have to be solved in advance, while or after the programming. Refer to the VENUS Software Programmer's Manual.

Problems that *can* be handled in run time are:

- Not enough liquid
- Liquid level not found (if it occurs as an exception)
- No tip picked up
- Clot detected
- Barcode unreadable (if it occurs as an exception)

- Execution error (channel no. 1 has an error (e.g. not enough liquid), then channel numbers 2-8 have an execution error because they have been stopped before completion of the step)

NOTE

In principle, each channel may have one or more different types of errors at a time.

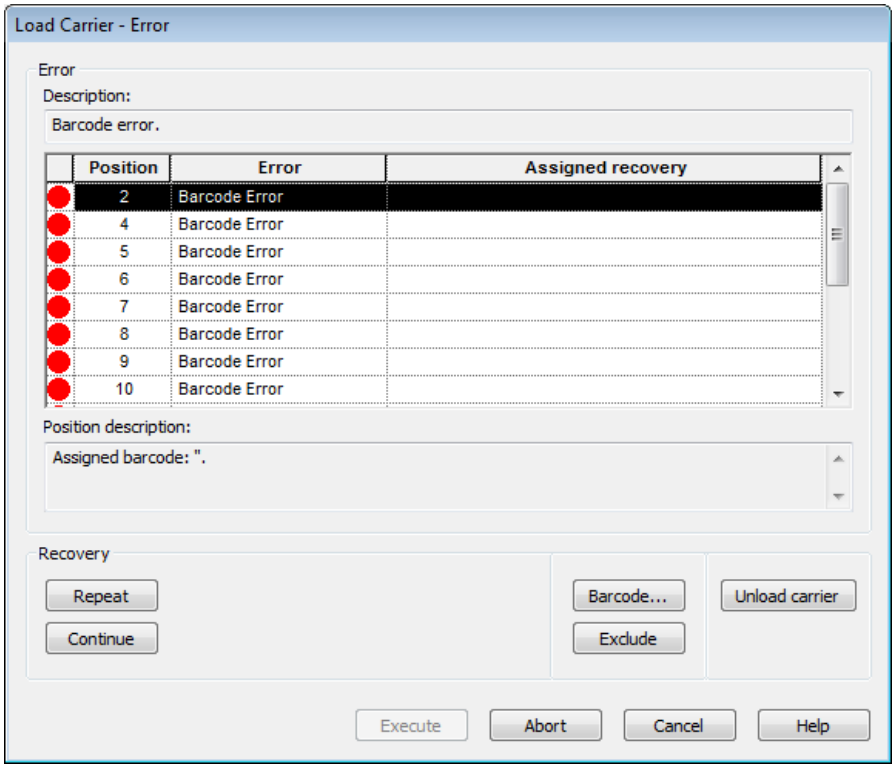
If all channels have the same error at the same time, a collective recovery can be made.

3.3.2 Run-Time Error Handling Examples

We now focus on some important examples. A detailed description is available in the VENUS Help Function. Click on **[Error Settings]** within the single step dialogs of the ML STAR -specific commands and select **[Help]**.

3.3.2.1 Barcode Reading Error

1. In the case of an error, the process can be continued using the error handling procedure. If, for example, a barcode of a carrier cannot be read, a dialog window opens up:



2. The error recovery can be programmed to:

- **Continue** Ignore the error message (here, failure of barcode reading)

  [Continue] makes no sense in the case of a barcode reading error - at least a manual entry has to be made so that barcode data exists for further processing.

- **Repeat** Try to read the barcode once again

  Often a repetition of reading will solve the problem because the reading speed is reduced then. The selected action is displayed in the field **[Assigned recovery]**.

- **Barcode...** Enter the barcode manually

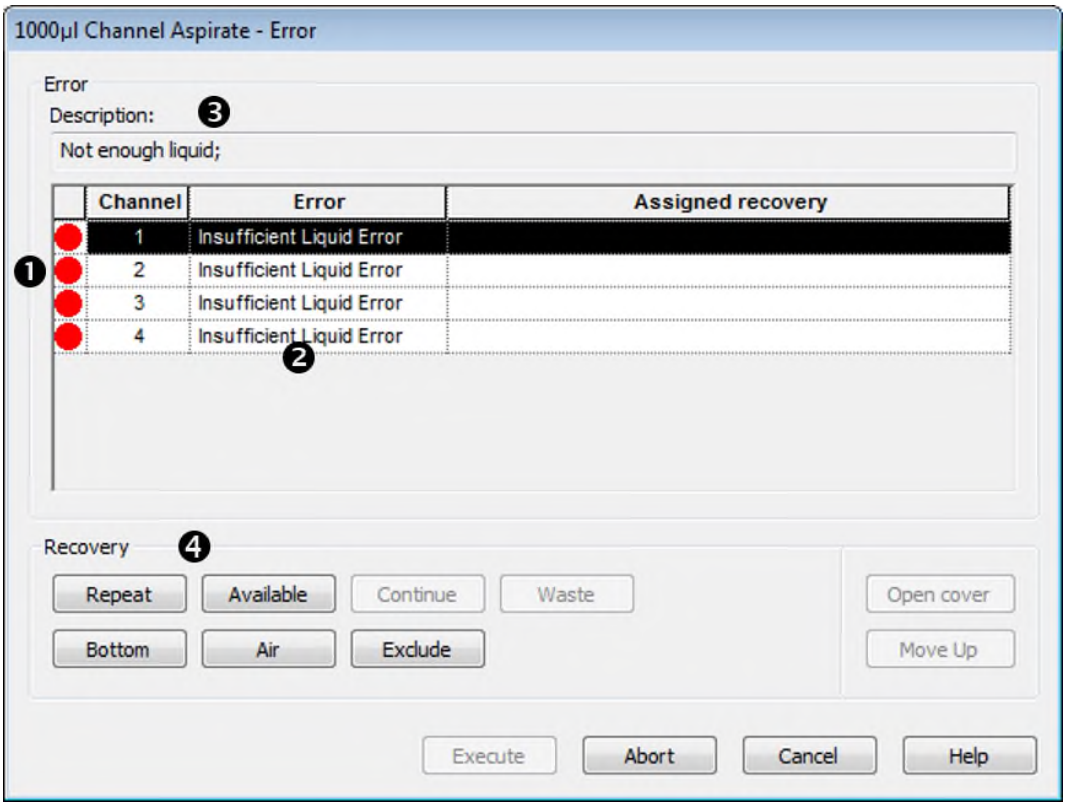
  Clicking **[Barcode]** opens an entry dialog box where a barcode can be entered (no entry is also allowed).

3. Assign a recovery option. The **[Execute]** button becomes active.
4. Click on **[Execute]**. The instrument proceeds with the selected recovery option.

3.3.2.2 Pipetting Error

1. If an error occurs with the pipetting channels, a dialog shows its error state and its recovery options for every single channel (Different channels can have different errors).

   For example, in case of an LLD error, such as no liquid in the container while aspirating, a window similar to the following is displayed:



2. All pipetting channels which produced an error are listed with a red dot on the left ❶. A short error description ❷ is given. A detailed error description is shown at the top text field ❸ labeled "Description:" for the selected pipetting channel. There may be different errors which lead to the same short error description.
3. The **[Recovery]** ❹ frame buttons offer several possible actions to solve the problem:

   - **[Repeat]:** Executes the command which caused the error once again
   - **[Available]:** Aspirates the available volume from the source and fills up the missing volume with air
   - **[Continue]:** Continues as if no error was recognized
   - **[Waste]:** Means that the erroneous tip is ejected to the waste and the channel is excluded
   - **[Bottom]:** Activates the channel to move down to the bottom of the container, and the available volume is aspirated without LLD
   - **[Air]:** Means air is pipetted instead of liquid and the method will continue
   - **[Exclude]:** Allows disabling any further action on the error-affected pipetting channel
   - **[Move Up]:** This is not a real error recovery procedure, but useful (e.g. to manually remove a clot). This causes the following actions (this action can be repeated):
     - moves the barcode reader of the Autoload (if present) to the very right
     - moves the selected pipetting channel up by 10 mm
   - **[Open Cover / Close Cover]:** this button does not start any error recovery actions either, but triggers the following actions:
     - **[Open Cover]:** enables opening the front cover during error recovery
     - **[Close Cover]:** enables closing the front cover before executing error recovery

4. An error recovery needs to be assigned for every error. Selecting a pipetting channel followed by any possible recovery procedure assigns the selected recovery procedure to all error-affected pipetting channels with the same error.
5. If any recovery procedure is assigned to a pipetting channel (even one which is not desired to be assigned), the dot ❶ in the first column (see picture above) changes color to green.
6. Some recovery buttons are disabled, to prevent further faulty steps, (e.g. an error-affected aspiration step cannot be "Continued"), to prevent any later dispense with insufficient volume.
7. When the last faulty pipetting channel is assigned to a recovery procedure, the **[Execute]** button becomes active and the system can proceed.
8. In any case, the method can be aborted without further recovery options.

3.3.2.3 Walk-Away (Predefined) Error Handling

1. The programmer can define a walk-away error handling which uses predefined default settings for different error situations. These settings can be customized for single steps and easy steps only. For Smart Steps, the error recovery is fixed.
2. For every instrument-specific single step of the method, an individual error recovery can be defined. The following can be configured:
   - The appearance of the error recovery dialogs (which buttons are available)
   - A timeout, after which the default recovery is carried out (the dialog automatically closes down)
   - The default procedure (what shall be executed if the timeout runs out)
   - Which error is flagged in the trace file
3. For this purpose, every instrument-specific Single Step and Easy Step has an **[Error Settings]** button. To deal with the subject of error handling consult the VENUS Software Programmer's Manual.

4. Maintenance

The ML STAR operator is instructed to maintain the instrument on a regular basis; this maintenance consists largely of surface cleaning and does not require opening up the instrument (i.e. any unscrewing, removing deck, covers, etc.).

The operator is responsible for changing consumable parts (disposable tips, needles, waste Bag, etc.). The HAMILTON Trained Field Service Engineer generally changes spare parts (PCBs, cables, channels, etc.).

Periodic maintenance routines need to be run in order to ensure safe and reliable operation of the ML STAR instrument and the accessories. The HAMILTON Trained Field Service Engineer will need to service (perform preventive maintenance) the instrument at least twice a year. In addition, the HAMILTON Trained Field Service Engineer may be called to repair a damaged component of the instrument or to resolve a functional problem, which the user cannot resolve himself (such as adjusting and calibrating the pipetting channels).

ATTENTION

When parts and accessories have application engineering (APE) modified parts adapted for a specific application, the original parts must be attached/used when performing adjustments/calibrations with the macro programs. After the adjustments/calibrations have been completed, re-install the modified parts.

Depending on how the VENUS Software is configured (maintenance configuration settings), the maintenance routines might be mandatory. If so defined – until they are completed, the user is reminded by a warning.

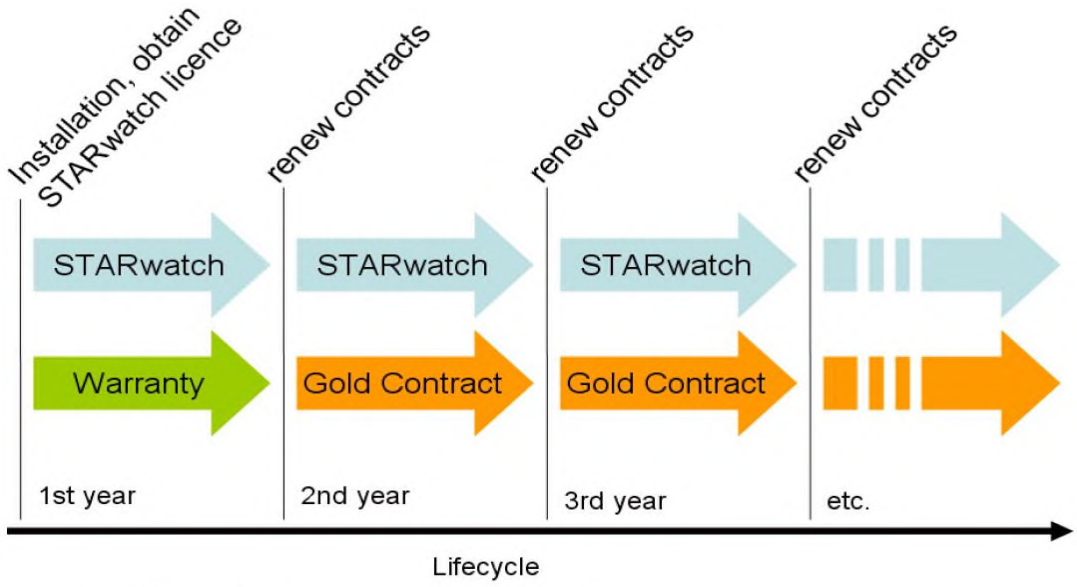
4.1 STARwatch Advanced Service

STARwatch is a pro-active service which increases the uptime of ML STAR instruments.

STARwatch continuously monitors the condition of the dedicated instrument. The captured data is automatically analyzed, and when a critical pattern is recognized, the service organization is immediately informed to provide a proactive intervention.

STARwatch intended use is primarily for high throughput laboratories where a certain consistency of daily routine use is given and where a short down time is critical.

STARwatch is available in addition during the warranty period and later in combination with a service contract. STARwatch has to be obtained on a yearly basis. Its license is valid for one year.



STARwatch Program

Refer to our Technical Note "STARwatch Condition Monitoring System", available on our website www.hamiltonrobotics.com .

4.2 Maintenance Intervals

In order to maintain the ML STAR in good condition, the following maintenance intervals are foreseen:

- **Daily**: Before the ML STAR start-up or shut-down. A Daily Maintenance is valid for 24 hours.
- **Weekly**: At the end of the week before the ML STAR is shut-down. A Weekly Maintenance is valid for 7 days.
- **Six-monthly**: Preventive maintenance service carried out by a HAMILTON Trained Field Service Engineer.

  This is based on a maximum usage of 8 h per day and 5 days per week. Higher numbers of hours per week will make shorter maintenance intervals necessary.

ATTENTION

If the operator decides not to run either daily or weekly maintenance before shut-down of the instrument, these routines must be implemented at the next run start.

If any parts of the instrument, carriers or racks have become contaminated, the weekly maintenance procedure must be performed.

The expiration date of the daily or weekly maintenance is not calculated from the day and time of the execution of the maintenance in question, but from the day and time of the start of the first method after having completed the maintenance. Right after completion of maintenance, the expiration date is not yet defined.

4.3 If Maintenance Fails

If an error is encountered during a maintenance procedure, try to rectify the problem and re-start the maintenance procedure. If this fails again, call your local HAMILTON Trained Field Service Engineer.

4.4 Materials Required

- Disposable latex gloves
- Protective glasses
- Lab coat
- Paper towels
- Lint-free cloths or Q-tips
- Set of 8 teaching needles - please refer to Section 8.1.3 Teaching Needles
- De-ionized water
- Mild Detergent & Disinfectant or MICROCIDE SQ (only available in the USA)

4.4.1 Cleaning Agents

According to the degree of contamination, the following cleaning agents are suitable for cleaning the parts of the ML STAR. It is recommended to clean only the described part.

Suitable Cleaning Agents	Concentration	Risk to instrument	Other comments
Water	Distilled or de-ionized water	-	-
Alcohol	70% ethanol 70% 2-Propanol (isopropanol).	Dissolve plastics, stress crack on transparent plastics	-
Detergents	Mild detergent Special detergents	Check before with vendor of the agents	Follow the vendor instructions for the correct concentration
Tissue	-	-	Use lint-free tissues only

ATTENTION

Strong detergents may dissolve carrier and worktable surface coatings. Use only cleaning agents that are suitable for Laboratory instruments.

The vendor of this disinfectant and cleaning agents can give more information.

Do not clean electronics, this can damage the instrument. Pay very close attention to the exposure times listed by the cleaning agent manufacturer.

Failure to follow manufacturers' recommend-actions can lead to incomplete cleaning and disinfection.

In some cases other decontamination procedures may be desirable, e.g. for reliably destroying infectious materials or DNA/RNA.

Many of these decontamination procedures are very aggressive and can cause damage to the ML STAR. Please follow the guideline given below. If using other decontamination procedures not listed here, be aware that they may increase service and maintenance requirements and may make shorter maintenance intervals necessary.

ATTENTION

Use cleaning, disinfecting and decontaminating fluid in accordance with the manufacturer's instructions. Do not use disinfecting materials, which contain hypochlorite (Javel water, Chlorox), or bleaching fluids.

Prepare disinfectant fluids according to their labeling.

Dilute the concentrated cleaning agent with deionized water according to the instructions on the bottle.

4.4.2 Possible Vendors of Cleaning and Disinfection Agents

Vendor	URL
Borer Chemie AG	http://www.borer.ch
Bode Chemie GmbH	http://www.bode-chemie.de
Schülke & Mayr GmbH	http://www.schuelke.com

4.5 Cleaning Parts of the Instrument

Instrument Part	Cleaning Agent
Stop-disk and Tip Eject Sleeve on the Pipetting Head	Water, alcohol, mild detergent
Waste Station	Water, alcohol, mild detergent
Deck	Alcohol, mild detergent
Housing	Alcohol
Metal parts	Alcohol
Carriers	Alcohol, mild detergent
Racks	Alcohol, mild detergent
Needles	Alcohol
Autoload or other scanner	Alcohol

ATTENTION

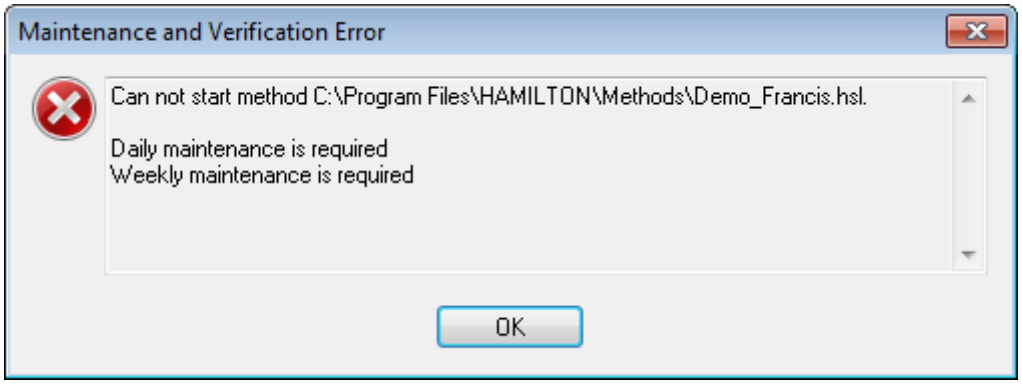
Cleaning procedures others than described are not recommended e.g. washing of carriers in washing machines, autoclaving of instrument components, etc.

4.6 Maintenance Procedures

The operator will be guided by the VENUS Software through the regularly scheduled maintenance procedures.

Depending on the maintenance configuration settings (*optional, warning* or *mandatory*), the instrument will display messages when starting up the instrument:

Displayed warning if maintenance is needed, configuration setting = warning



Displayed error if maintenance is needed, configuration setting = mandatory

ATTENTION

Always wear disposable gloves during maintenance.

Do not clean the instrument in the vicinity of unshielded flames or devices which could create sparks. Do not use hot air blowers to dry the instrument. The liquids used for cleaning may be flammable.

Maintenance work on the iSWAP is not permitted, because adjustment may be lost. Clean the iSWAP only where necessary after spillage and be careful of sharp edges on the gripper jaws.

Any regulations for waste disposal specific for the country of operation must be taken into account and observed.

Routine Completion:

A maintenance routine is completed once the procedure has been fully implemented and the results are within specifications.

Aborting Maintenance Procedures:

Aborting a maintenance procedure will lead to a 'failed' status, and maintenance will need to be started again.

4.6.1 Daily Maintenance

Daily maintenance is configuration dependent. The following tasks are examples belonging to the daily maintenance:

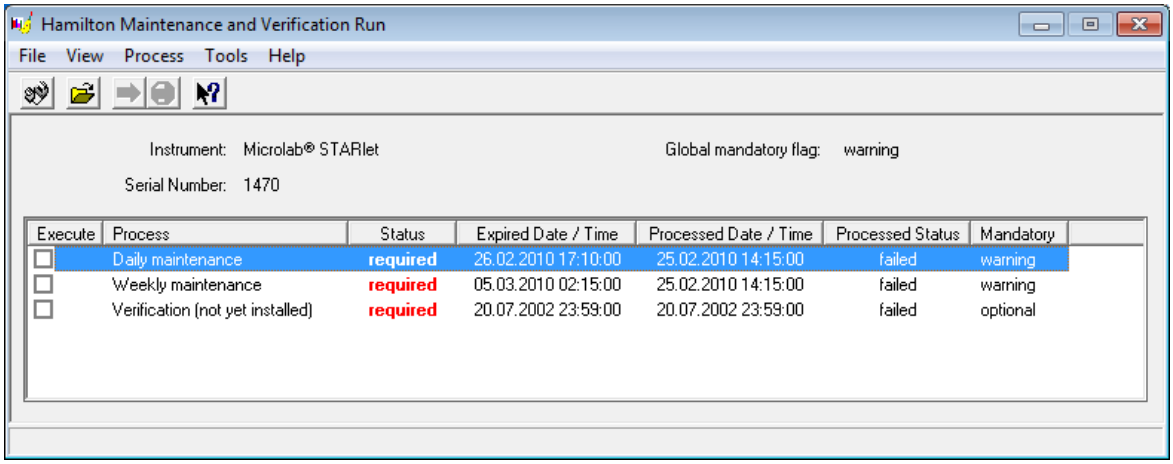
- Check if the deck is clean
- Empty the tip waste
- Check the tightness of the 5 ml/1000 µl pipetting channels
- Verify the cLLD function (5 ml/1000 µl pipetting channels)
- Perform the rinse procedure of the wash station(s)
- Check if the CVS vacuum system is working properly

To execute the Daily Maintenance procedure:

1. Double-click the "**Microlab STAR Maintenance & Verification**" Icon on the desktop:

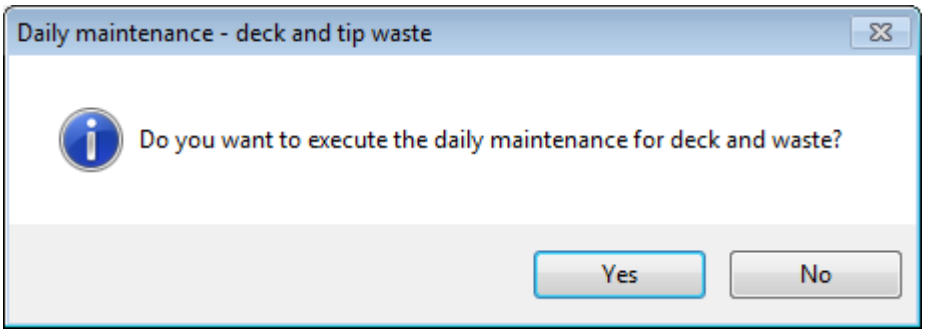
2. In the **Hamilton Maintenance and Verification Run** Main Window, the process status information view lists all maintenance and verification processes for the connected/selected instrument.

   Here the operator can start the necessary commands to perform all maintenance-related routines.



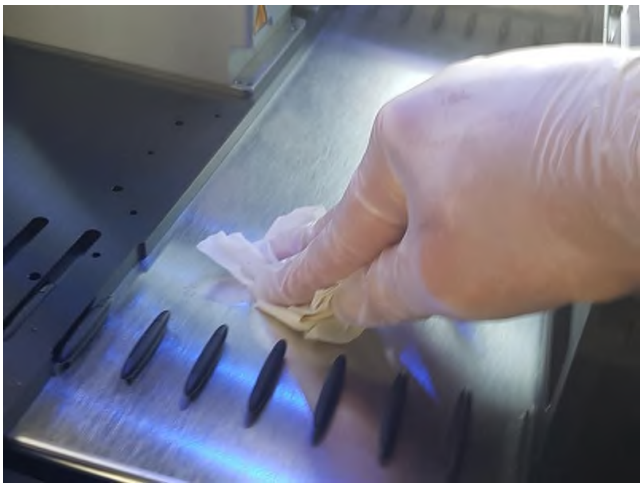
"Hamilton Maintenance and Verification Run" Main Window

3. Select the desired maintenance routine by clicking the specific check box and by pressing the **[Run Process]** button. The VENUS Software will then issue on-screen instructions detailing all procedures required to perform the selected maintenance routine.
4. After instrument initialization, the operator will be asked to execute the daily maintenance:



5. After clicking **[Yes]**, the daily maintenance procedure will be started. Pressing **[No]** will abort the procedure.
6. The front cover (the hinged acrylic glass window that shields the instrument in front) can be opened for user intervention.
7. Once the maintenance procedure has been started, the pipetting arm moves to the left side. The operator now has access to the deck to check if cleaning is needed or not:

   - If the deck is clean, continue with the daily maintenance.
   - If the deck needs to be cleaned, the daily maintenance can be interrupted. Instead of the daily maintenance carry out the weekly maintenance.

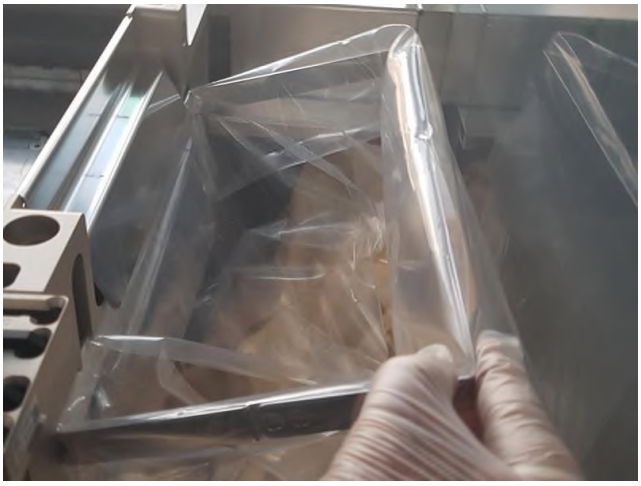


8. Continuing the daily maintenance procedure will lead the user to the next maintenance task. The tip waste needs to be emptied. Dispose of it with the rest of the laboratory's contaminated waste.

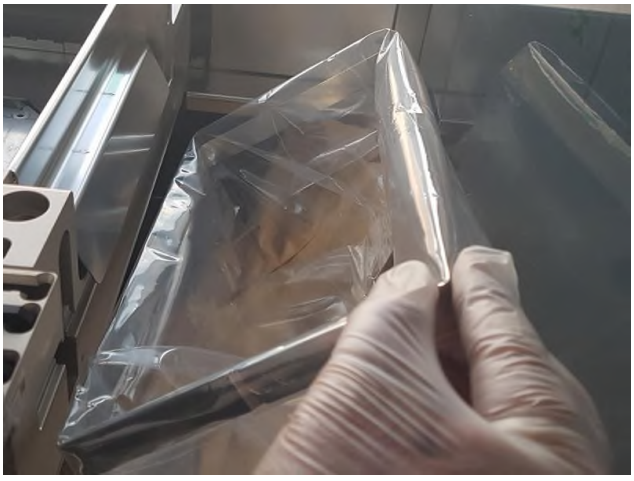
   a. Remove the Tip Waste Bag

      The Tip Waste Bag is removed by:

      - Lifting the right side of the waste frame



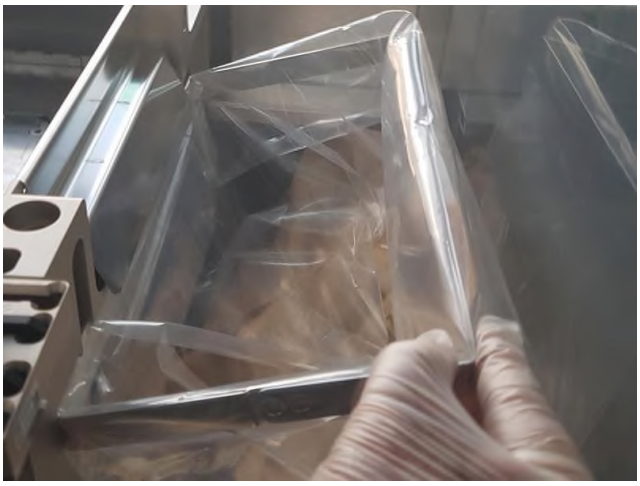
      - Lowering the left side of the waste frame



   b. Remove the Tip Waste Bag from the waste frame

c. Insert a new Tip Waste Bag

d. Place the Tip Waste Bag into the correct position under the ledge of the Waste Block



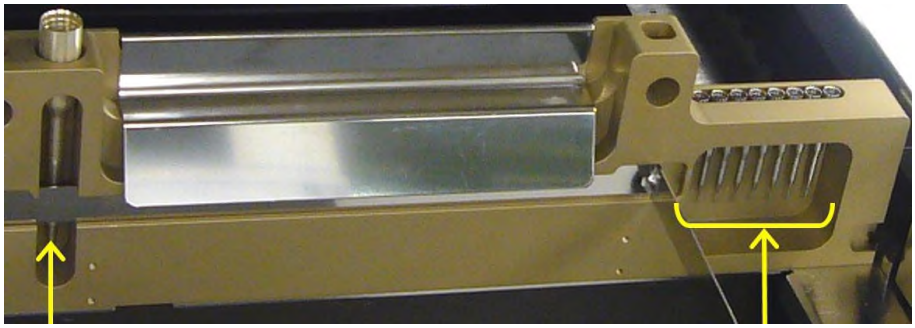
e. Place the frame in a horizontal position

ATTENTION

BIOHAZARD

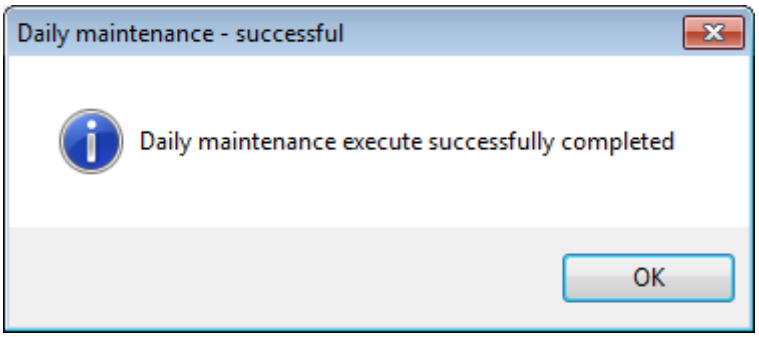
The tip waste and the plastic bag are always to be regarded as contaminated.

9. For the next steps, the maintenance needles are required.



Maintenance Needle 5 ml Pipetting Channel ***Maintenance Needles 1000 µl Pipetting Channels***

10. The procedure continues with the tightness check of the pipetting channels. The pipetting arm will travel to the right hand side to pick up the teaching needles. Two checks are done with the pipetting channels, the over-pressure and the under-pressure check.
11. For the capacitive liquid level detection (cLLD) check, the needles are picked-up again. Each pipetting channel is checked one at a time, for proper cLLD function.
12. If there is a CVS installed, a rinse procedure is started. Refer to Section 4.13 Crystal Vacuum System.
13. If there is a needle wash station installed, a rinse procedure is started. Refer to Section 4.10 CR Needle Wash Station.
14. When the daily maintenance is completed, the "Daily maintenance execute successfully completed" screen is displayed.



15. The daily maintenance process status is saved on the instrument and a report file is created. Refer to Section 4.7 Printing a Report.

NOTE

If any parts of the instrument, carriers or racks have become contaminated, the weekly maintenance procedure must be performed.

4.6.2 Weekly Maintenance

Weekly maintenance is configuration dependent. The following tasks are examples belonging to the weekly maintenance:

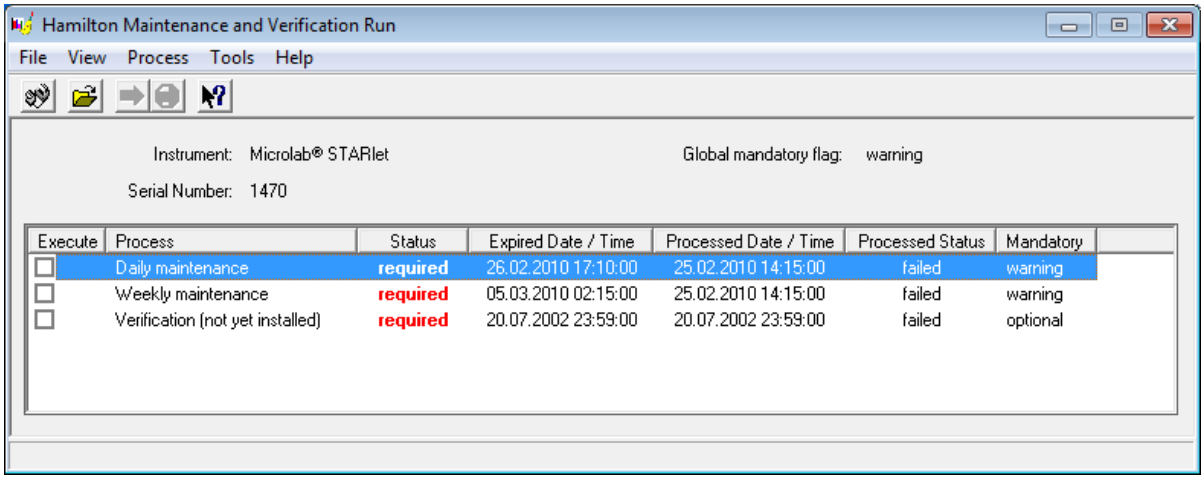
- Clean the deck and carriers
- Check the condition of all carriers
- Empty and clean the tip waste
- Check the tightness of the 5 ml/1000 µl pipetting channels
- Verify the cLLD function (5 ml/1000 µl pipetting channels)
- Perform the maintenance procedures for the wash station(s)
- Clean of the pipetting head: stop disk, O-ring, tip eject sleeve
- Clean of the covers, Autoload protection ribbon
- Check if the CVS vacuum is working properly

To execute the Weekly Maintenance procedure:

1. Double-click the "**Microlab STAR Maintenance & Verification**" Icon on the desktop:

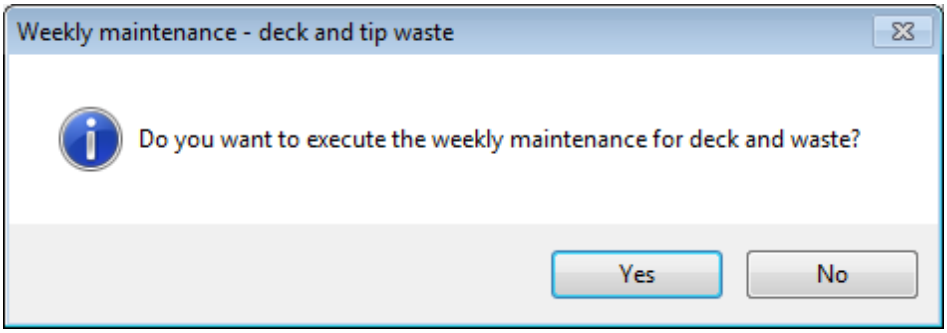
2. In the **Hamilton Maintenance and Verification Run** Main Window, the process status information view lists all maintenance and verification processes for the connected/selected instrument.

   Here the operator can start the necessary commands to perform all maintenance-related routines.

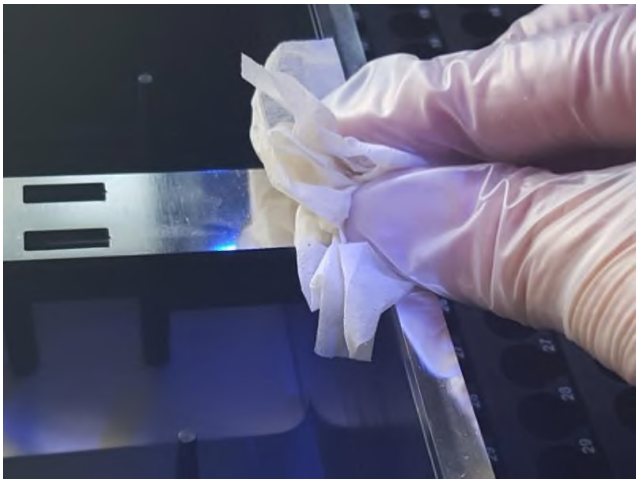


"Hamilton Maintenance and Verification Run" Main Window

3. Select the desired maintenance routine by clicking the specific check box and by pressing the **[Run Process]** Button. The VENUS Software will then issue on-screen instructions detailing all procedures required to perform the selected maintenance routine.
4. After initialization of the instrument, the operator will be asked to execute the weekly maintenance:



5. When the instrument is initialized, the weekly maintenance program advises the user to unload the deck manually. If the Autoload option is available, this step is carried out automatically.
6. Clean all carriers and all modules with an appropriate cleaning agent as described in Section 4.5 Cleaning Parts of the Instrument and leave them to dry. If they are heavily soiled, these carriers may be immersed in the cleaning agent (see Section 4.4.1 Cleaning Agents).
7. Examine each carrier for scratches on the barcode and any signs of damage. If damage is apparent, replace with new carriers.



NOTE

Carriers must be completely clean and dry before re-using.

8. Continuing the weekly maintenance program will advise the Autoload to move to the right hand side of the instrument.

9. Open the front cover and wipe the deck with a cloth saturated with an appropriate cleaning agent as described in Section 4.5 Cleaning Parts of the Instrument. The slide blocks in particular must be checked for cleanliness. Close the front cover.

ATTENTION

Do not spray directly at the Autoload unit, Decapper Module or at electrical boards or connectors.

10. The next step of the maintenance procedure will advise the Autoload (if configured) to move to the left hand side of the instrument. The tip waste needs to be emptied and cleaned. Dispose of tip waste with the rest of the laboratory's contaminated waste. Remove the frame that holds the plastic bag in place, and discard the plastic in the laboratory's contaminated waste. Pull a new plastic bag over the frame and re-attach it.

ATTENTION

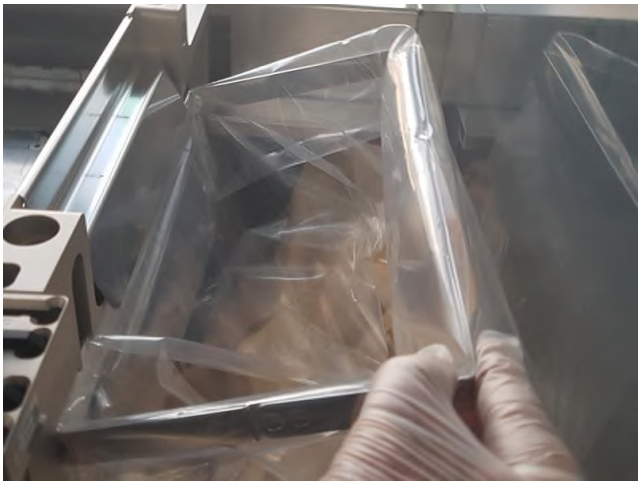
BIOHAZARD

The tip waste, the tip eject plate, and the plastic bag are always to be regarded as contaminated.

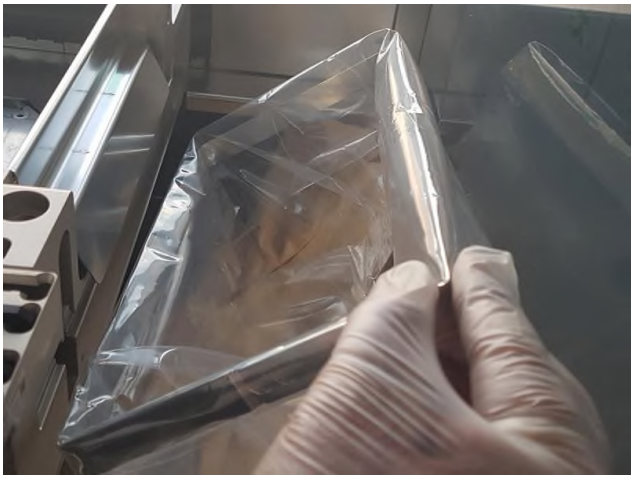
a. Remove the Tip Waste Bag

   The Tip Waste Bag is removed by:

   - Lifting the right side of the Waste Frame.



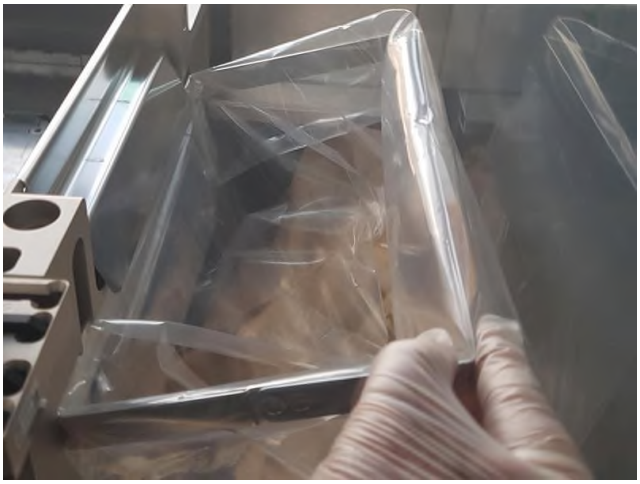
- Lowering the left side of the Waste Frame from under the ledge of the Waste Block.



b. Remove the Tip Waste Bag from the Waste Frame.

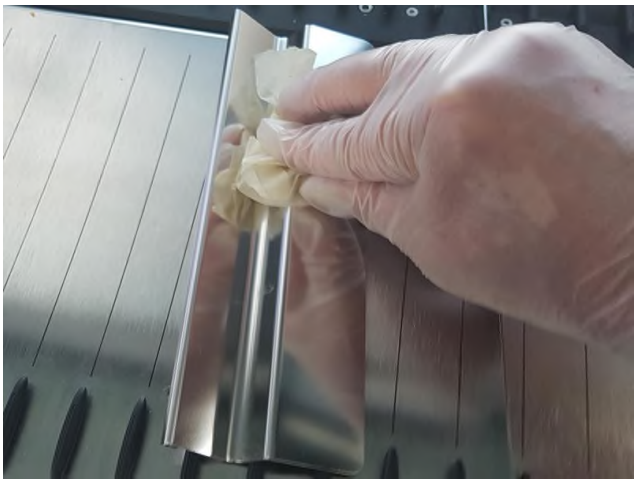
c. Insert a new Tip Waste Bag.

d. Place the Tip Waste Bag and Waste Frame into the correct position under the ledge of the Waste Block.



e. Place the Waste Frame in a horizontal position.

f. Remove the tip eject plate of the waste station and clean it with an appropriate cleaning agent as described in Section 4.5 Cleaning Parts of the Instrument.



g. Put the clean tip eject plate back in place.

11. Clean the maintenance needles.

1000 µl Maintenance Needles

5ml Maintenance Needles

ATTENTION

Do not wet the maintenance needles inside.

12. To prevent unreliable barcode reading, check the laser scanner window of the barcode reader and clean it with a lint-free cloth or Q-tips lightly soaked in ethanol (70 %).

ATTENTION

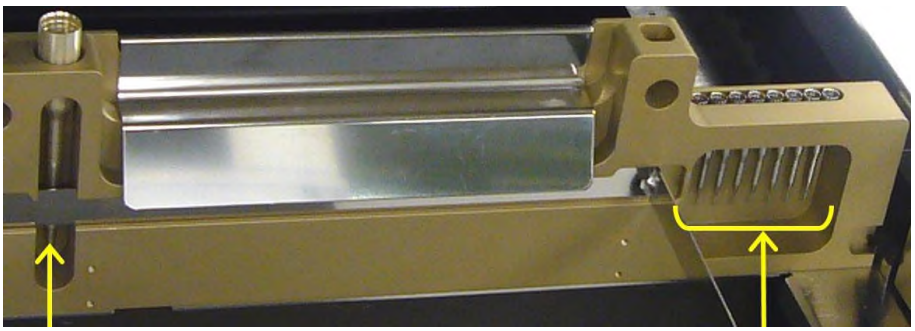
The laser scanner window must be completely dry and free from dust and fibers before the instrument can be reused.

13. Clean the Autoload protecting ribbon with a lint-free cloth and the appropriate cleaning agent and wipe without exerting pressure (see Section 4.5 Cleaning Parts of the Instrument).

ATTENTION

Do not spray directly at the Autoload unit, Decapper Module or at electrical boards or connectors.

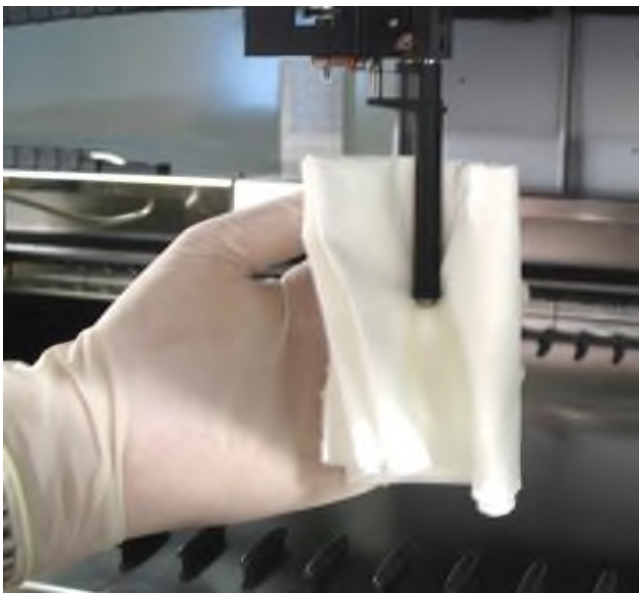
14. For the next steps, the maintenance needles are required.



Maintenance Needle 5 ml Pipetting Channel ***Maintenance Needles 1000 µl Pipetting Channels***

15. The procedure continues with the tightness check of the pipetting channels. The X-Arm will travel to the right hand side to pick up the maintenance needles. Two checks are done with the pipetting channels, the over-pressure and under-pressure check.
16. For the capacitive liquid level detection (cLLD) check, the needles are picked-up again. One pipetting channel after the other is checked for proper cLLD function.
17. If there is a CVS installed, a rinse procedure is started. Refer to Section 4.14 Temperature Controlled Carrier (TCC).
18. If there is a needle wash station installed, a weekly maintenance procedure is started. Refer to Section 4.8 Needle Maintenance.

19. Clean the tip eject sleeve (outer part of the pipetting heads) with a lint-free cloth and the appropriate cleaning agent (see Section 4.5 Cleaning Parts of the Instrument).

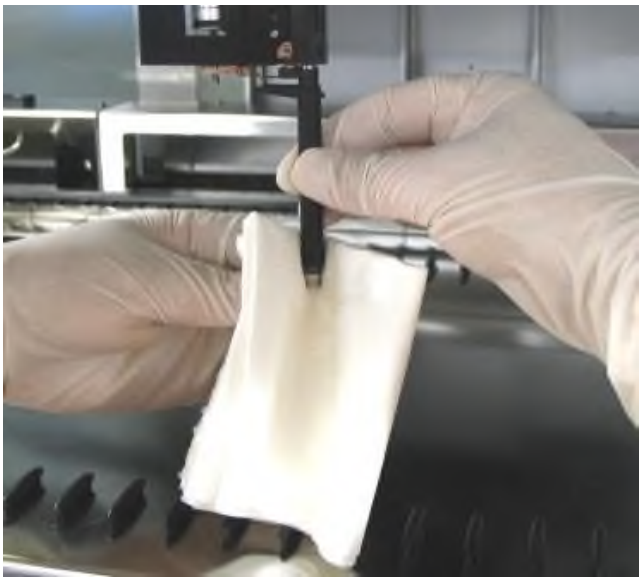


ATTENTION

Take care that no liquid gets inside the pipetting channel.

Whenever it is necessary to move the pipetting channels on the X-Arm, move them gently by pushing close to their Y-slide. Never force them as this may lead to damage. If possible, switch on the instrument as this will result in a smoother motion when the pipetting channels have to be moved on the X-Arm.

20. Clean the stop disk and the O-rings (outer part of the pipetting heads) with a lint-free cloth and the appropriate cleaning agent (see Section 4.5 Cleaning Parts of the Instrument).



ATTENTION

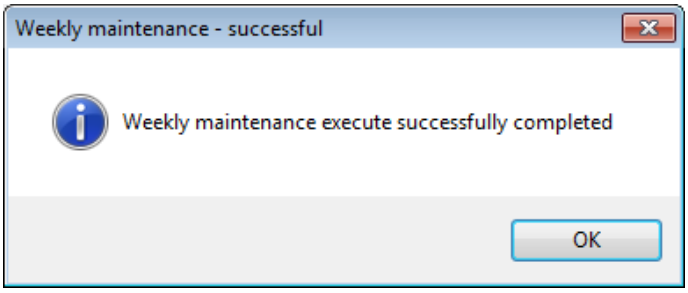
Take care that no liquid gets inside the tip eject sleeve.

Whenever it is necessary to move the pipetting channels on the pipetting-arm, move them gently by pushing close to their y-slide. Never force them as this may lead to damage. If possible, switch on the instrument as this will result in a smoother motion when the pipetting channels have to be moved on the pipetting-arm.

21. Clean the front and side covers with a lint-free cloth and the appropriate cleaning agent and wipe dry (see Section 4.5 Cleaning Parts of the Instrument).

22. Clean the X-guide shaft behind the upper front cover with a dry cloth at least once a month.

23. When the weekly maintenance is completed, the "Weekly maintenance execute successfully completed" screen is displayed:



24. The weekly maintenance process status is saved on the instrument and a report file is created (see Section 4.7 Printing a Report).

4.7 Printing a Report

The maintenance process status can be printed. To print such a report:

1. Go to "**File → Open Report**". All maintenance and verification processes which are found in the default "Report Path" are listed.
2. If necessary, change the report path using the browse button **[...].**
3. Select a report and click the **[Open]** button. The Report Viewer displays the selected report file.
4. Go to "**File → Print**" to print the report file.

4.8 Needle Maintenance

The recommended procedures for maintenance of the needles - if the needles are badly stained – are as follows:

1. Remove the needle(s) with the needle pickup tool (Needle Service Kit, see Section 8.1.3 Teaching Needles).
2. Clean the needles in an ultrasonic bath at 50 °C containing an appropriate cleaning agent for 15 - 20 minutes (see Section 4.4.1 Cleaning Agents). Follow the vendor instructions for the concentration of the cleaning agent.
3. Rinse the needles with warm de-ionized water (50 °C).
4. Put the needles back into the wash module.

NOTE

HAMILTON recommends replacing steel needles every six months.

4.9 CO-RE 96 Probe Head TADM

This section describes the preparation and execution of the maintenance procedures for the CO-RE 96 Probe Head TADM 1000 µl. For more information about the labware, please refer to the TADM Manual or the VENUS Programmer's Manual.

A CO-RE 96 Probe Head TADM 1000 µl must be mounted in a ML STAR to perform the maintenance procedures indicated in this section.

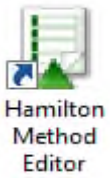
It is required to have either of the carriers (presented in the table below) with a defined plate position for the maintenance tool.

	
PN 188042 MULTIFLEX DWP MODULE	***PN 188041 MULTIFLEX MTP MODULE***
	
PN 182090 PLT_CAR-L5AC	***PN 182365 PLT_CAR_L5MD***
	
PN 199199 Maintenance Tool	

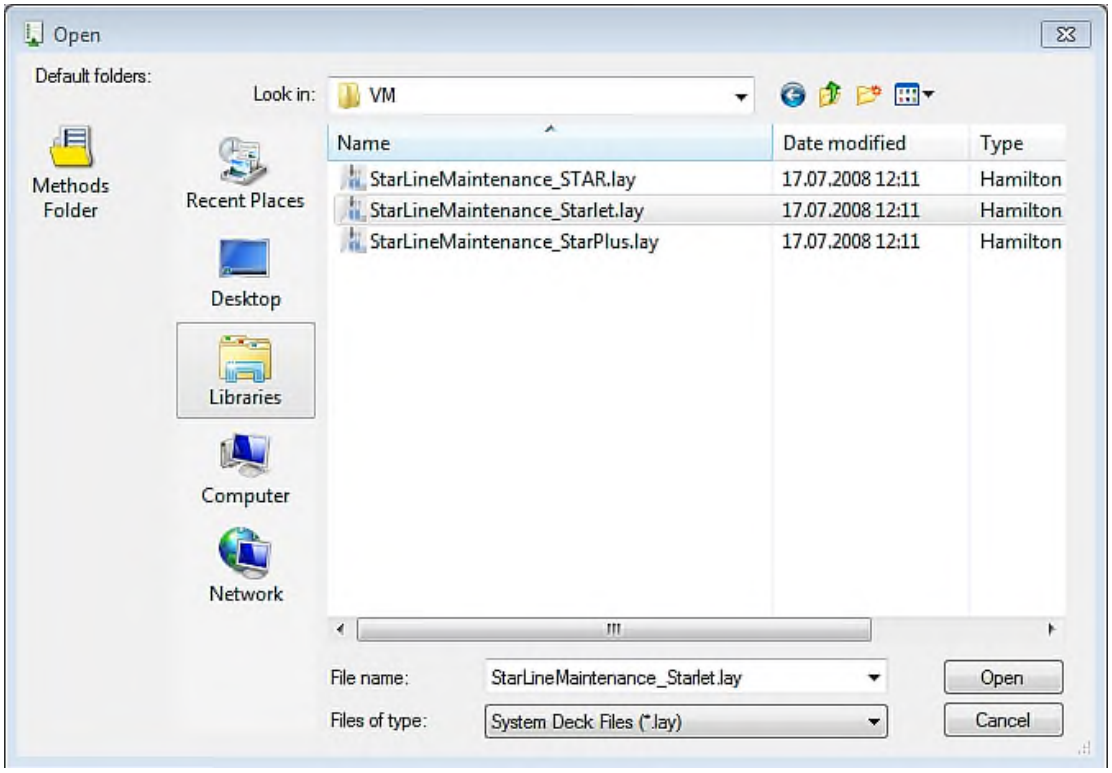
4.9.1 Preparation

To prepare for the maintenance procedure, create a System Deck Layout. Follow these steps:

1. Start the **Hamilton Method Editor** by activating the icon shown:



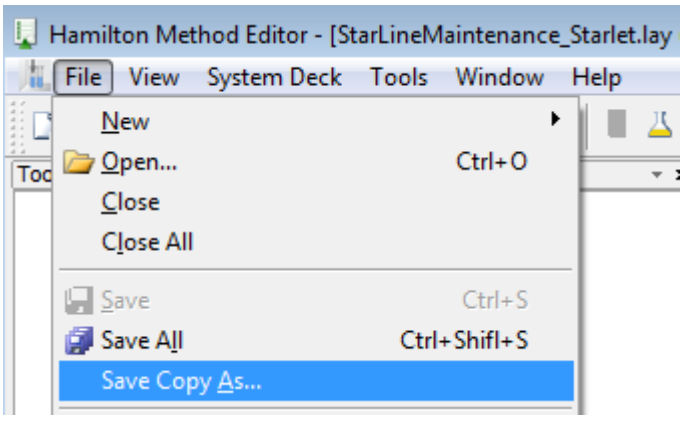
2. Open the existing deck layout found in the directory: \\HAMILTON \Methods \VM
   - For the ML STARlet StarLineMaintenance_Starlet.lay
   - For the ML STAR StarLineMaintenance_Star.lay
   - For the ML STARplus StarLineMaintenance_Starplus.lay

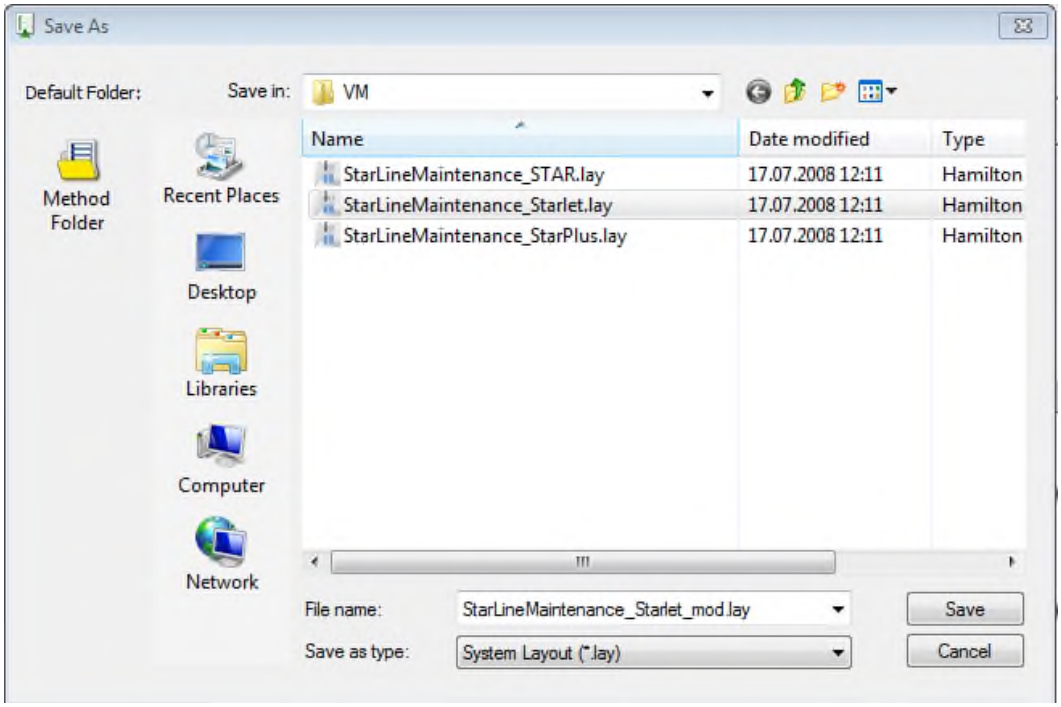


NOTE

Selecting the "Files of type" to System Deck Files (.lay) is necessary. This will make the desired *.lay files visible and selectable.*

3. Create and save a copy of the file selected. In this case, save a copy of the "StarLineMaintenance_Starlet.lay" File and name it as "StarLineMaintenance_Starlet_mod.lay". For uniformity purposes and to avoid confusion in later steps, always use the "_mod" keyword as a prefix to the file copies.



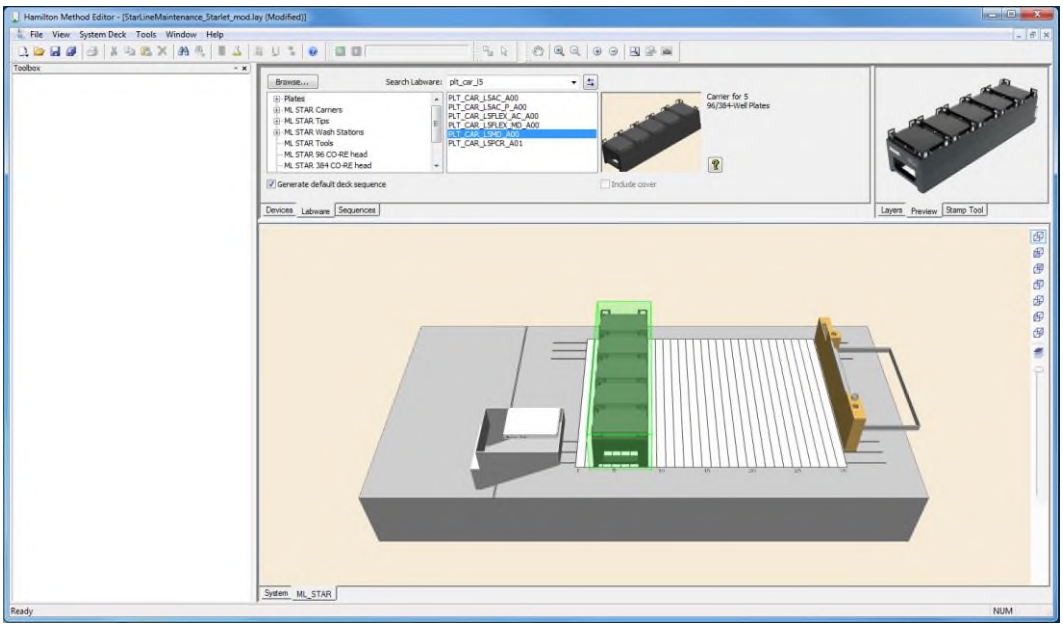


4. Open the file copy "StarLineMaintenance_Starlet_mod.lay", after successfully creating a duplicate.
5. Add the chosen plate carrier from the table, as instructed earlier. Remember to add the plate carrier onto a track position reachable by the CO-RE 96 Probe Head TADM.

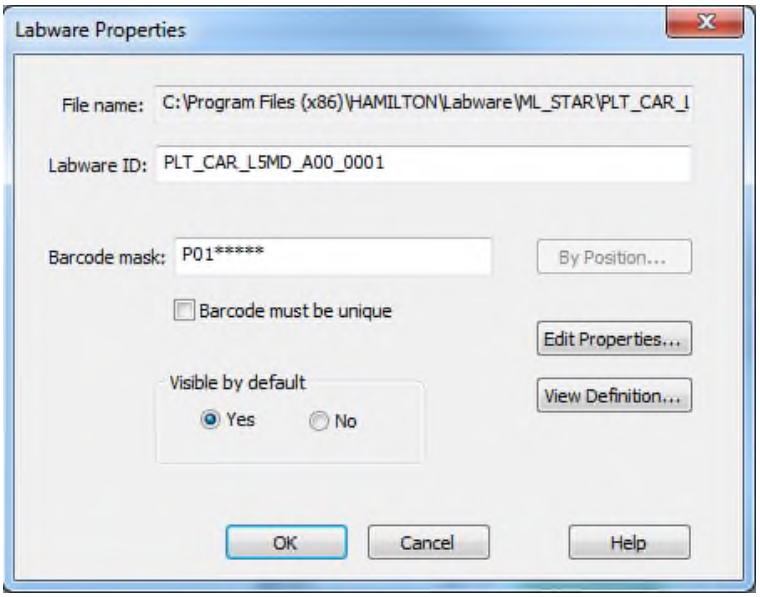
NOTE

The deck position has to be available for the maintenance routine.

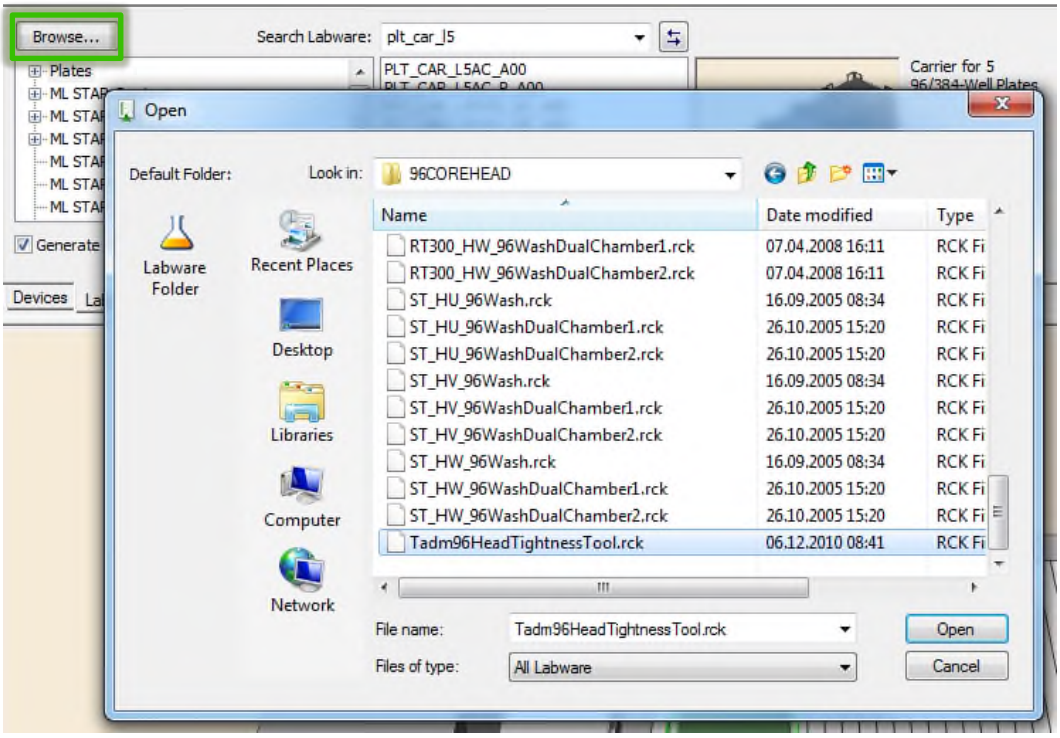
In this example, a PLT_CAR_L5MD (PN 182365) has been chosen.



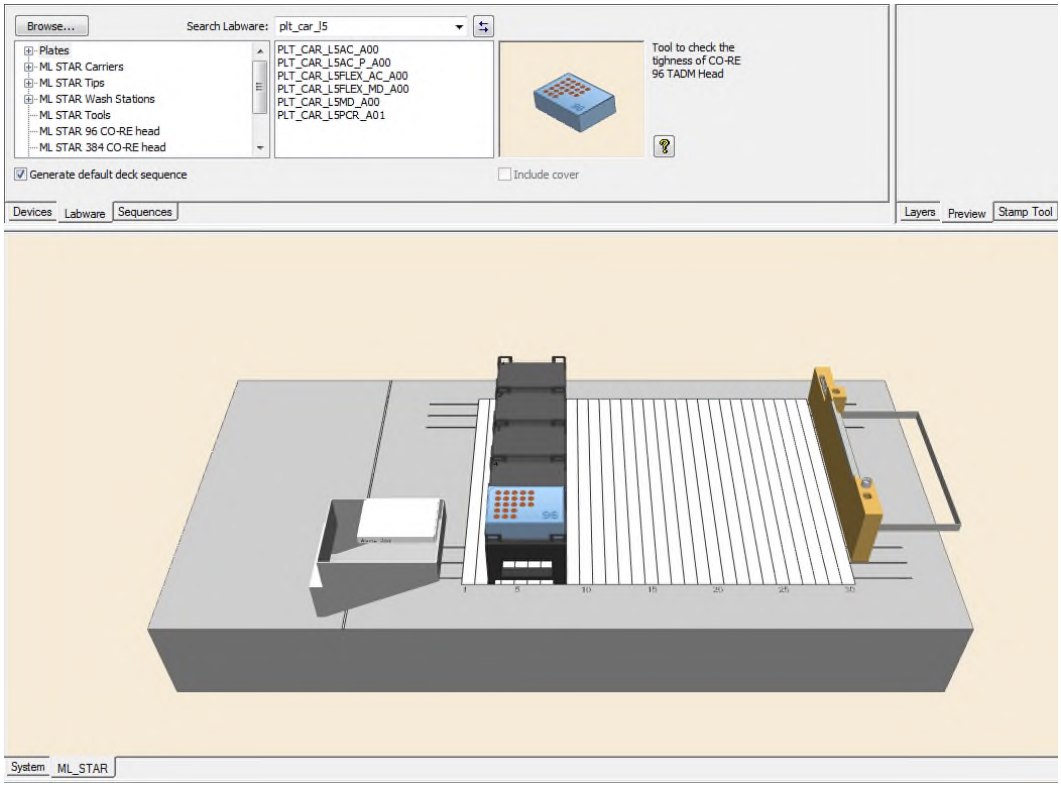
6. Right click on the plate carrier and select Properties. A dialog box with predefined values will display, as shown below. Set the Labware Property "Visible by default" to Yes. This will display the carrier in the deck layout editor during the maintenance process. To close the dialog box, click **[OK]**.



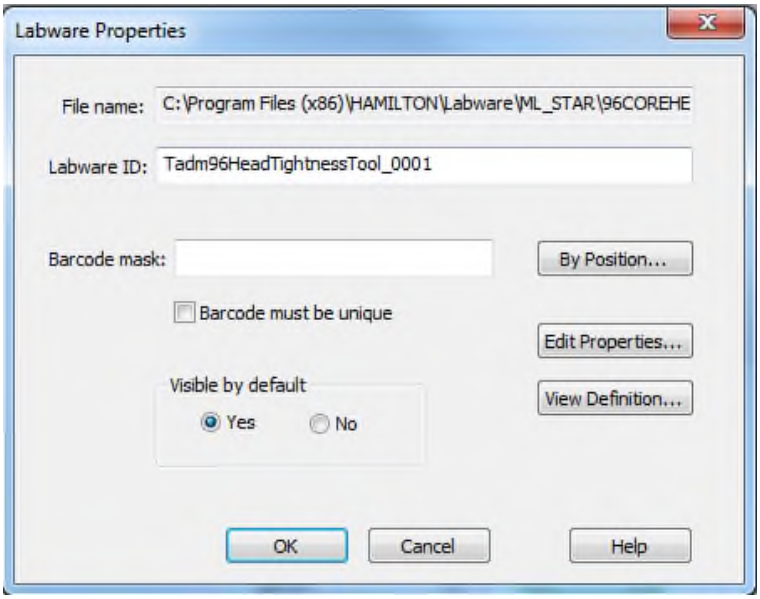
7. To add the Maintenance Tool to the chosen plate carrier, click [Browse] from the Labware Tab of the Method Editor. Locate the file by opening through the folders **Hamilton > LabWare > ML_STAR > 96COREHEAD**.



8. Add the tool onto the plate carrier. Any available position on the plate carrier may be chosen.



9. After having successfully added the tool onto the plate carrier, right click on it and select Properties. A dialog box with predefined values will prompt, as shown below. Set the Labware Property "Visible by default" to Yes. This will display the tool in the deck layout editor during the maintenance process. To close the dialog box, click **[OK]**.



10. Finish by saving the deck layout and closing the Method Editor.

4.9.2 Daily Maintenance

Daily maintenance requires the following tasks to be performed:

- Inspect the workspace/deck for cleanliness
- Empty the tip waste and liquid waste
- Have the instrument perform a tightness check
- Have the instrument perform a cLLD check

ATTENTION

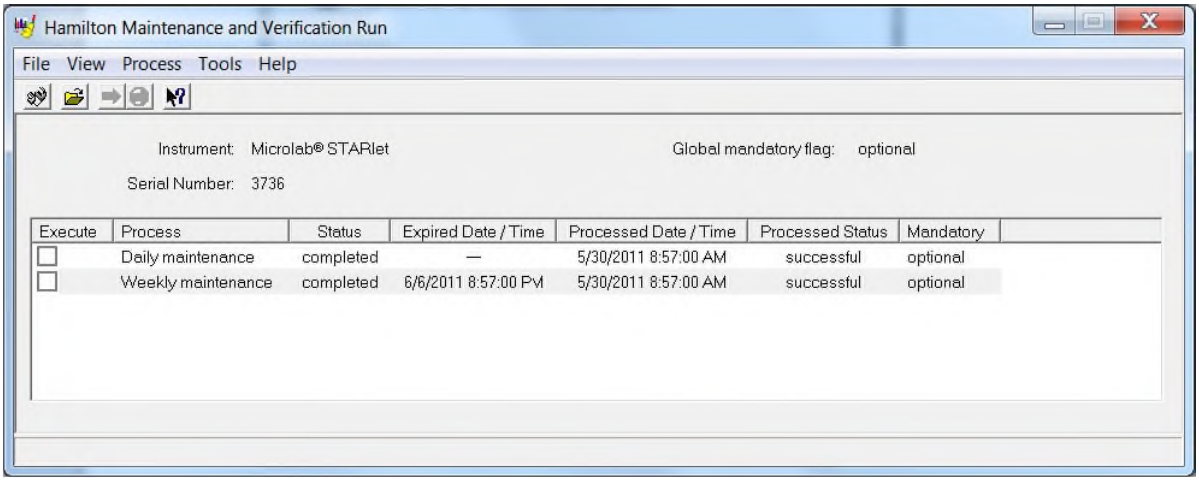
Be aware of moving parts (i.e. X-Arm with its CO-RE 96 Probe Head TADM).

An instruction to place the carrier and the CO-RE 96 MPH Maintenance Tool onto the deck will be prompted. This step requires opening the front cover. After doing so, make sure to close the front cover. This applies to both placing and removing a carrier and a tool.

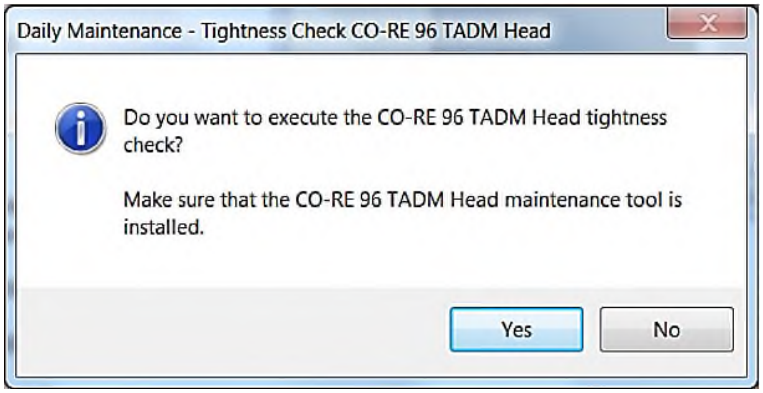
The following steps shall be followed to perform the daily maintenance for the CO-RE 96 Probe Head TADM.

1. Start by activating the Microlab STAR Maintenance & Verification icon shown:

2. Now that the **Hamilton Maintenance and Verification** is running, select "Daily Maintenance" from the choices and start. See image below.



3. Follow the instructions and when prompted with the dialog box shown below, select **[YES]**. This will execute the CO-RE 96 Probe Head TADM tightness check.

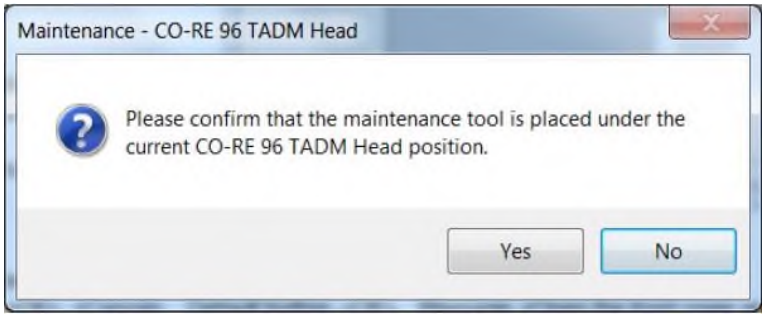


4. Place the carrier with the tool onto the deck. Make sure that the tool when placed on the carrier has a "Maintenance" label engraved on it, as shown in the photo below. The position of the carrier may vary depending on how the system deck has been defined, as described in Section 4.9.1 Preparation.

NOTE

During this step, the front cover may be opened. Once finished, make sure to close the cover. Only then continue to the next procedure.

5. The CO-RE 96 Probe Head TADM will be pre-positioned above the tool. A dialog box to confirm the correct position will be prompted.

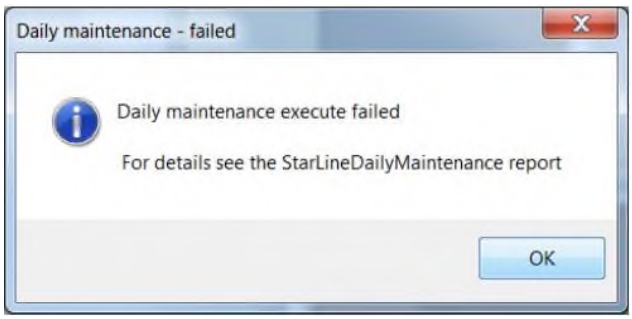


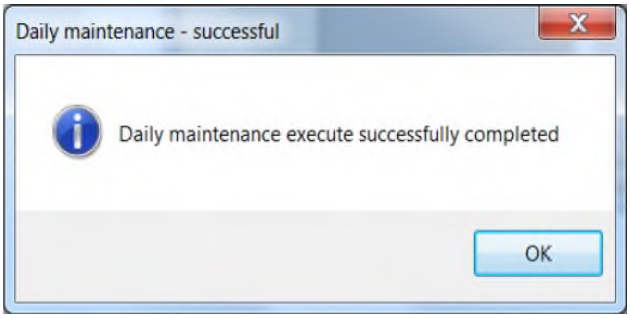
ATTENTION

*Selecting **[YES]** when the position is incorrect may lead to a crash and can damage the CO-RE 96 Probe Head TADM and the Tool.*

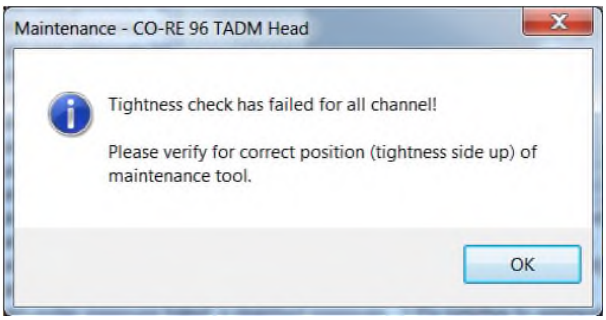
6. The CO-RE 96 Probe Head TADM will pick up the Tool and the tightness check will be executed.

7. Upon the completion of the tightness check, either of the following prompts will be displayed:





OR



8. Now, remove the carrier and the maintenance tool. The CO-RE 96 Probe Head TADM Daily Maintenance is completed.

ATTENTION

Ensure that the carrier and the tool are removed from the deck after the maintenance has been finished.

4.9.3 Weekly Maintenance

Weekly maintenance requires the following tasks to be performed:

- Clean the workspace
- Check the carriers and devices for damage
- Empty and clean the solid and liquid waste
- Clean main instrument modules thoroughly
- Clean the instrument covers
- Have the instrument perform a tightness check
- Have the instrument perform a cLLD check

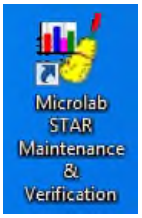
ATTENTION

Be aware of moving parts (i.e. X-Arm with its CO-RE 96 Probe Head TADM).

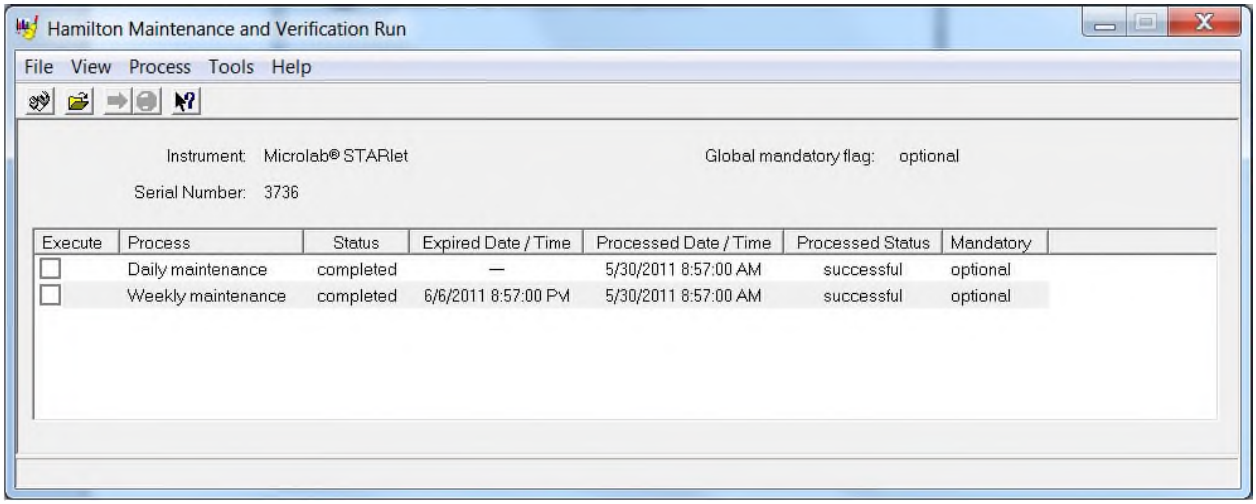
An instruction to place the carrier and the CO-RE 96 MPH Maintenance Tool onto the deck will be prompted. This step requires opening the front cover. After doing so, make sure to close the front cover. This applies to both placing and removing a carrier and a tool.

The following steps shall be followed to perform the weekly maintenance for the CO-RE 96 Probe Head TADM.

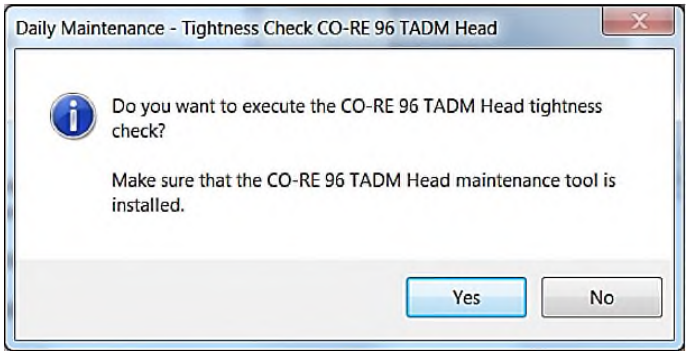
1. Start by activating the Microlab STAR Maintenance & Verification icon shown:



2. Now that the **Hamilton Maintenance and Verification** is running, select "Weekly Maintenance" from the choices and start. See image below.



3. Follow the instructions and when prompted with the dialog box shown below, select **[YES]**. This will execute the CO-RE 96 Probe Head TADM tightness check.

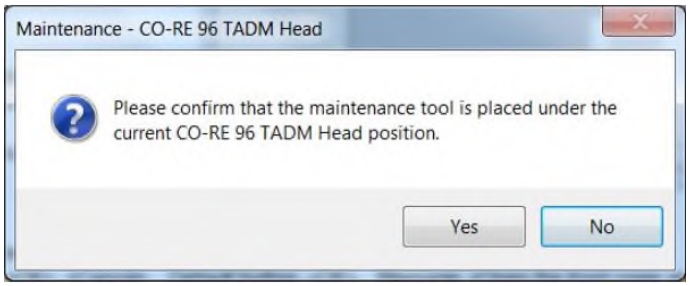


4. Place the carrier with the tool onto the deck. Make sure that the tool when placed on the carrier has a "Maintenance" label engraved on it, as shown in the photo below. The position of the carrier may vary depending on how the system deck has been defined, as described in Section 4.9.1 Preparation.

NOTE

During this step, the front cover may be opened. Once finished, make sure to close the cover. Only then continue to the next procedure.

5. The CO-RE 96 Probe Head TADM will be pre-positioned above the tool. A dialog box to confirm the correct position will be displayed.

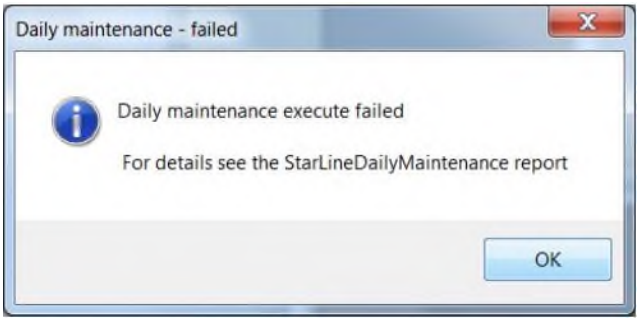


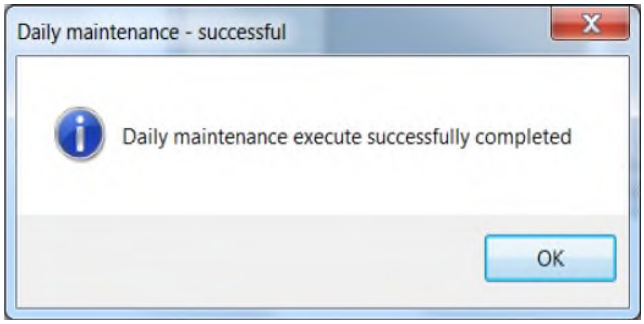
ATTENTION

*Selecting **[YES]** when the position is incorrect may lead to a crash and can damage the CO-RE 96 Probe Head TADM and the Tool.*

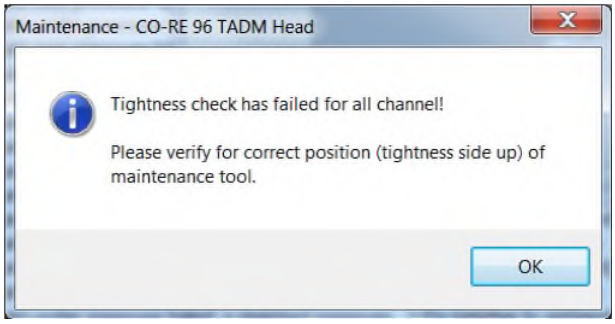
6. The CO-RE 96 Probe Head TADM will pick up the Tool and the tightness check will be executed.

7. Upon completion of the tightness check, either of the following prompts will be displayed:





OR



8. Now, remove the carrier and the maintenance tool. The CO-RE 96 Probe Head TADM Weekly Maintenance is completed.

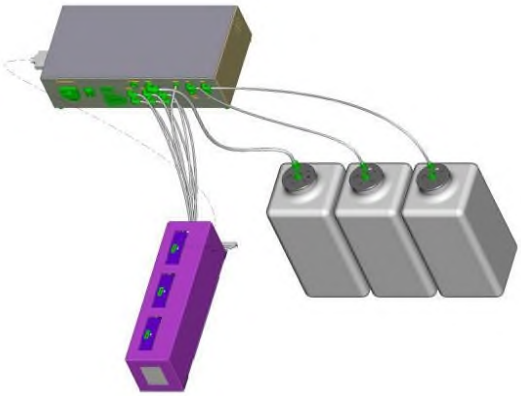
ATTENTION

Ensure that the carrier and the tool are removed from the deck after the maintenance has been finished.

4.10 CR Needle Wash Station

HAMILTON recommends the following daily rinse routine and the weekly maintenance procedure to maintain the functionality of the needle wash station. Excellent washing results can be achieved only with periodic maintenance.

If so configured in the VENUS Software, daily and weekly maintenance are required.

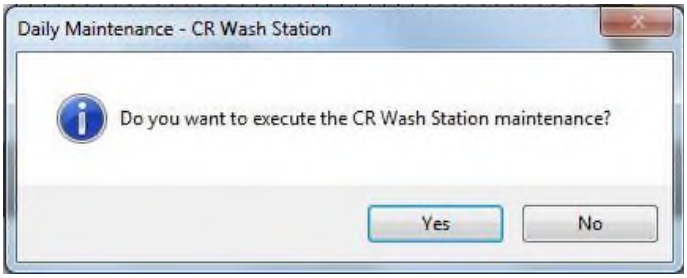


Before starting the maintenance procedures, make sure the needles to be washed are placed in the Needle Wash Station.

4.10.1 Daily Maintenance

The daily maintenance procedure for the CR needle wash station takes 15 minutes. The purpose of this procedure is to rinse the fluid path of the wash station. With this procedure, deposits inside the fluid path can be minimized. If the wash station is seriously soiled, the operator shall start the weekly maintenance instead of the daily maintenance.

Empty any remaining liquid of the wash containers and the waste container. Partially fill the wash solution containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the containers.



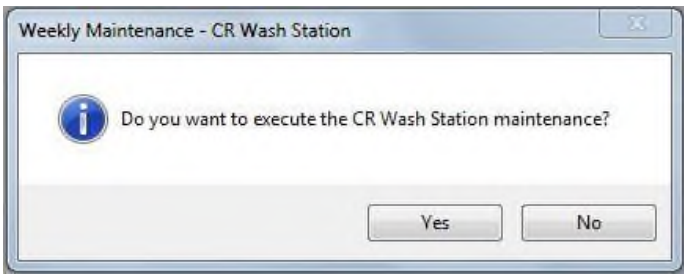
1. Once the maintenance procedure for the CR needle wash station has been started, the software advises the operator to fill Wash Container 1 (red dot on the container lid) with 4 liters of de-ionized water and 40 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
2. 1x wash step (wash solution; Rinse time 60 seconds, soak time 1 second, flow rate 12 ml/second, draining time 12 seconds).

3. The rinse procedure is interrupted by the maintenance program, to advise the operator to empty wash container 1 (red dot on the container lid) and to fill it up with 3 liters of de-ionized water.
4. 2x rinse step (de-ionized water; Rinse time 60 seconds, soak time 1 second, flow rate 12 ml/second, draining time 12 seconds).
5. At the end of the daily maintenance procedure, wash container 1 and the waste container need to be emptied.

4.10.2 Weekly Maintenance

The weekly maintenance procedure for the CR needle wash station takes 40 minutes. The purpose of this procedure is to carry out periodical in-depth maintenance. The fluid path of the wash station and the needles are soaked in a special cleaning solution.

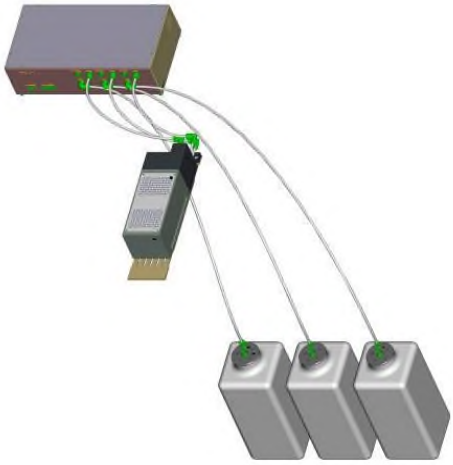
Empty any remaining liquid of the wash containers and the waste container. Partially fill the wash solution containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the containers.



1. Once the maintenance procedure for the needle wash station has been started, the software advises the operator to fill wash container 1 (red dot on the container lid) with 6 liters of de-ionized water and 60 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
2. 1x wash step (wash solution; Rinse time 60 seconds, soak time 300 seconds, flow rate 12 ml/second, draining time 12 seconds).
3. 1x wash step (wash solution; Rinse time 60 seconds, soak time 1 second, flow rate 12 ml/second, draining time 12 seconds).
4. The procedure is interrupted by the maintenance program to advise the operator to empty wash container 1 (red dot on the container lid) and to fill it up with 6 liters of de-ionized water. Partially fill the wash solution container with de-ionized water. With the cap facing upwards, shake the container in the lengthwise direction for a few moments. Empty the container.
5. Fill up wash container 1 with 3 liters of de-ionized water.
6. 2x rinse step (de-ionized water; Rinse time 60 seconds, soak time 1 second, flow rate 12 ml/second, draining time 12 seconds).
7. At the end of the weekly maintenance procedure, wash container 1 and the waste container need to be emptied and let them dry.

4.11 CO-RE 96/384 Dual Wash Station

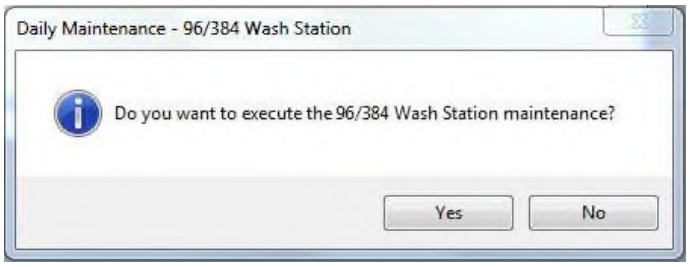
HAMILTON recommends the following daily rinse routine and the weekly maintenance procedure to maintain the functionality of the CO-RE 96/3484 dual wash station. Excellent washing results can be achieved only with periodic maintenance.



4.11.1 Daily Maintenance

The daily maintenance procedure for the wash station 96/384 takes 10 minutes. The purpose of this procedure is to rinse the fluid path of the wash station. With this procedure, deposits inside the fluid path can be minimized. If the wash station is seriously soiled, the operator shall carry out the weekly maintenance instead of the daily maintenance.

Empty any remaining liquid of the wash container and waste container. Partially fill the wash solution containers with de-ionized water. With the cap facing upwards, shake the container in the lengthwise direction for a few moments. Empty the containers.



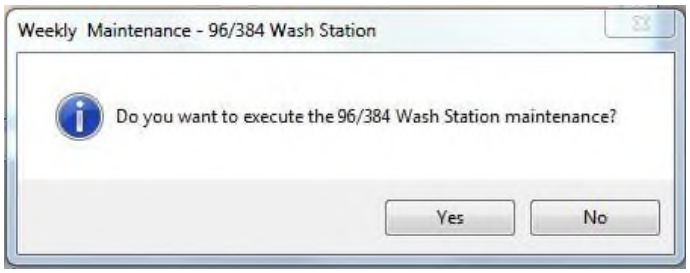
1. Fill the wash containers with 2 liters of de-ionized water and 20 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
2. Empty the waste container.
3. 1x priming (initialize washer, empty).
4. 2x rinse step for both wash chambers (fill washer, empty).
5. Empty the wash containers. Partially fill the wash containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the wash containers. Fill the wash containers with 3 liters of de-ionized water again.

6. 2x rinse step for both wash chambers (fill washer, empty).
7. Empty the wash containers and waste container.
8. Empty any remaining wash solution. Partially fill the containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the containers and let them dry.

4.11.2 Weekly Maintenance

The weekly maintenance procedure for the wash station 96/384 takes 20 minutes. To guarantee excellent wash efficiency, carry out the following steps at least once a week.

Empty any remaining liquid of the wash container and waste container. Partially fill the wash solution containers with de-ionized water. With the cap facing upwards, shake the container in the lengthwise direction for a few moments. Empty the containers.



1. Fill the wash containers with 2 liters of de-ionized water and 20 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
2. Empty the waste container.
3. 1x Priming (Initialize washer, empty).
4. 1x Rinse step for both wash chambers (fill washer).
5. Soaking: Timer of 5 minutes.
6. 1x Rinse step for both wash chambers (empty washer, refill after empty).
7. 1x Empty washer.
8. Empty the wash containers. Partially fill the wash containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the wash containers. Fill the wash containers with 3 liters of de-ionized water again.
9. 2x rinse step for both wash chambers (Fill washer, empty).
10. 1x Empty the washer.
11. Empty any remaining wash solution of the wash and waste containers. Partially fill the containers with de-ionized water. With the cap facing upwards, shake the containers in the lengthwise direction for a few moments. Empty the containers and let them dry.

4.12 HAMILTON Heater Shaker

HAMILTON recommends cleaning the HHS regularly to prolong its lifetime. In case of contamination or spillages clean the HHS immediately. The HHS must be turned off and cooled down for the maintenance procedure.

The recommended procedure for the daily/weekly maintenance of the HAMILTON Heater Shaker:

- Clean all surfaces with a cloth soaked with an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).

ATTENTION

HOT SURFACE

Do not clean the HHS when it is still hot!

Wait until the HHS has cooled down to room temperature before starting the cleaning procedure.

4.13 Crystal Vacuum System

The CVS has to be connected to a HAMILTON waste bottle, or to a suitable in-house liquid disposal system. HAMILTON recommends maintaining the CVS by following the daily and weekly maintenance procedures described below. Only with a periodical maintenance a long lifetime of the CVS can be achieved.

4.13.1 Activating the CVS Maintenance

Activating the CVS maintenance requires modification of the two source files namely "**HslStarLineMaintMetConst.hs_**" and "**HslStarLineMaintMetConst.stp**". They are available in the folder **…\HAMILTON\Library**.

The files can be modified by following the instructions below:

1. Make a copy of both HSL source files namely "HslStarLineMaintMetConst.hs_" and "HslStarLineMaintMetConst.stp".
2. After renaming the files, modify the value of the constant "**isBVSInstalled(hslFalse);**" to "**isBVSInstalled(hslTrue);**".
3. Save the files.
4. Rename the original files to "old_ HslStarLineMaintMetConst.hs_" and "old_ HslStarLineMaintMetConst.stp".
5. Finally rename the modified files generated with the original names.

4.13.2 Daily Maintenance

The daily maintenance procedure for the CVS takes 10 minutes. The purpose of this procedure is to rinse the tubing and the vacuum chamber. With this procedure, deposits inside the fluid path can be minimized. If the CVS is seriously soiled, the operator shall start the weekly maintenance instead of the daily maintenance.

1. Detach the Manifold Top from the vacuum module. It has to be cleaned separately to avoid water spill.
2. Clean the Manifold Top with a lint-free cloth soaked in an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents). Rinse afterwards with de-ionized water and let dry before using it again.
3. Prepare approximately 500 ml of de-ionized water with 5.0 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents). Pour the mixture into the vacuum chamber of the CVS carrier until it is full to the brim. Do not overspill. Use a soaking time of 1 minute.
4. Enter the COM port where the CVS is connected.
5. Vacate the vacuum chamber.
6. Pour in de-ionized water into the vacuum chamber of the CVS carrier until it is full to the brim. Do not overspill.
7. Vacate the vacuum chamber.
8. Re-install the Manifold Top. Close the front cover and press OK to continue. Cancel will abort the daily maintenance.
9. Empty the waste bottle. Fill the waste bottle with de-ionized water to a degree. Ensure that the waste bottle is securely closed. Shake the container in a semi-inclined position for a few moments. Empty the waste bottle.

4.13.3 Weekly Maintenance

The weekly maintenance procedure for the CVS takes 15 minutes time. The purpose of this procedure is to rinse the tubing and the vacuum chamber. With this procedure, deposits inside the fluid path can be minimized.

1. Detach the Manifold Top from the vacuum module. It has to be cleaned separately to avoid water spill.
2. Clean the Manifold Top with a lint-free cloth soaked in an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents). Rinse afterwards with de-ionized water and let dry before using it again.
3. Prepare approximately 500 ml of de-ionized water with 5.0 ml of an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents). Pour the mixture into the vacuum chamber of the CVS carrier until it is full to the brim. Do not overspill. Use a soaking time of 5 minutes.
4. Enter the COM port where the CVS is connected.
5. Vacate the vacuum chamber.
6. Pour in de-ionized water into the vacuum chamber of the CVS carrier until it is full to the brim. Do not overspill.
7. Vacate the vacuum chamber.
8. Re-install the Manifold Top. Close the front cover and press OK to continue. Cancel will abort the daily maintenance.
9. Empty the waste bottle. Fill the waste bottle with de-ionized water to a degree. Ensure that the waste bottle is securely closed. Shake the container in a semi-inclined position for a few moments. Empty the waste bottle.

4.14 Multiflex Cooling-Heating Module

The recommended procedure for the daily/weekly maintenance of the Cooling-Heating Module:

- Clean all surfaces with a cloth soaked with an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).

5. Verification

To perform the verification of the ML STAR instrument, "Field Verification 2" is used. The Field Verification 2 can only be performed by a HAMILTON Trained Field Service Engineer.

Verification specifications (as applicable in the Microlab STAR Field Verification 2 User Manual) are different from the specifications given in the technical specifications of this user manual.

Especially the volume verification specifications (as applicable in the Microlab STAR Field Verification 2 User Manual) are different from the pipetting specifications for disposable tips given in the technical specifications of this user manual. The field verification contains qualified procedures and equipment defined by HAMILTON to demonstrate and to verify the correct function of the instrument according to specifications given by HAMILTON suitable for the field. The field verification is therefore a reference defined by HAMILTON to compare the instrument´s performance according to given procedures valid for a broad operating range.

Based on that, specifications as applicable in the technical specifications will be achieved by maintaining defined environmental conditions in the laboratory, by keeping the operating range as small as possible, by optimizing the methods such as adapting the liquid classes, knowing the sample liquids and the characteristics of used labware, etc. See also Section 1 Information about Safety and Security.

For the 1000 µl pipetting channels, the CO-RE 96 Probe Head and the CO-RE 384 Probe Head, a dye-pipetting procedure followed by gravimetric and photometric analysis is used to verify the trueness and precision. The 5 ml pipetting channels are verified by a gravimetric approach.

Devices such as Heater Shaker, Cooler, barcode reader and cover safety can also be verified with the "Field Verification 2".

Further information can be found in Section 8.1 Appendix A: Ordering Information of this manual; for additional information, consult a local HAMILTON representative.

6. ML STAR Instrument Decontamination

The decontamination method must be adapted to the respective application and the substances associated with it. The user takes full responsibility for the appropriate decontamination of the entire equipment.

Apart from regular decontamination, the user must thoroughly decontaminate the instrument according to local regulations in the following cases:

- Before any maintenance or service work is performed on the instrument
- In case of accidents (e.g. crash, spilt substances, etc.)
- Before the instrument or parts of it are returned to HAMILTON (e.g. for repair)
- Prior to storage of the instrument
- Prior to disposal of the instrument or parts of it
- Generally before the instrument or parts of it leave the user's site

Note

Dilute the concentrated disinfectant agent with deionized water according to the instructions on the bottle.

ATTENTION

Risk of Damage to equipment.

Do not use cleaning or disinfecting solutions which contain hypochlorite, such as bleach.

ATTENTION

Risk of Damage to equipment.

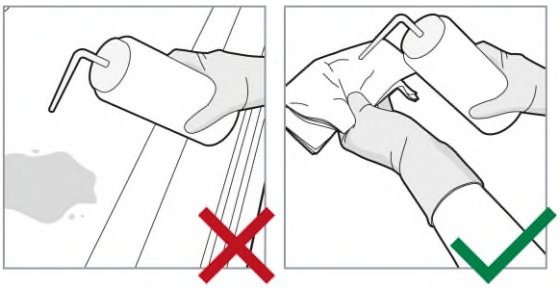
Spraying the instrument or carriers surfaces will lead to damage of equipment.

- *Do not spray the disinfectant agent onto the instrument or carrier surfaces directly. Spray the liquid onto a paper towel to clean the surfaces and devices.*

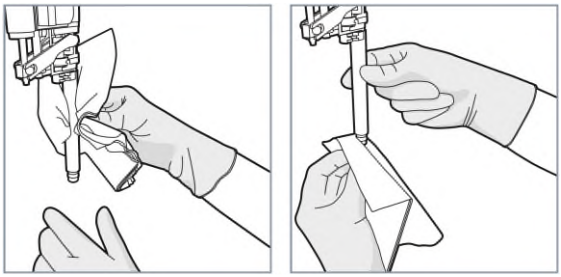
In general, for decontamination GLP must be observed. The recommended procedure for decontaminating the ML STAR instrument:

1. Spray the front and side covers with an appropriate cleaning agent (see (see Section 4.4.1 Cleaning Agents) and wipe with clean paper towels.

2. Spray lint-free paper towels with an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
3. Open the front cover and wipe the deck with the paper towels.



4. Remove the tip eject plate of the tip waste station and clean it.
5. Spray an appropriate cleaning agent directly onto the surface of the tip waste station (see Section 4.4.1 Cleaning Agents).
6. Remove the frame that holds the plastic bag in place, and discard the plastic bag in the laboratory's contaminated waste. Put the tip eject plate back in place.
7. Clean the tip eject sleeve (outer part of the pipetting channels) with a lint-free paper towel soaked in an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents).
8. Lift up the tip eject sleeve and wipe the O-ring and stop disk.
9. Repeat this procedure for all pipetting channels.



10. The use of disinfectant liquids is recommended for decontamination of Carriers. Clean all carriers with an appropriate cleaning agent (see Section 4.4.1 Cleaning Agents) and leave them to dry. If they are heavily soiled, soak them afterwards in the appropriate cleaning agent.
11. Perform needle maintenance (see Section 4.8 Needle Maintenance).
12. Perform washer maintenance (see Section 4 Maintenance and the following sections).

ATTENTION

Autoclaving (i.e. using superheated steam under pressure) may only be used for decontamination followed by final disposal of disposables and racks; they will be destroyed by the process. Autoclaving cannot be used for instrument components or accessories (pipetting channels, transport tools, wash station components, heaters, shakers, carriers, etc.).

ATTENTION

In some cases, other decontamination procedures may be desirable (e.g. for reliably destruction of infectious materials or DNA/RNA). Many of these decontamination procedures are very aggressive and can cause damage to the ML STAR instrument. Please follow the guideline given below. If using other decontamination procedures not listed here, be aware that they may increase service and maintenance requirements and may make shorter maintenance intervals necessary.

6.1 Surface Decontamination using Liquids

ATTENTION

For surface decontamination using liquids, perform a risk analysis regarding increased service and maintenance requirements.

Preferably, use the procedure described using the appropriate cleaning agent. Other procedures have not been tested by HAMILTON. Use the appropriate cleaning agent at a maximum temperature of 50 °C.

If using other liquids or sprays for surface decontamination, follow the manufacturer´s instructions. Pay particular attention to potential corrosiveness (e.g. acidic or alkaline solutions and oxidizing agents).

NOTE

Use of such agents may increase service and maintenance requirements (O-rings exchange, greasing of spindles, etc.) and may make shorter maintenance intervals necessary.

NOTE

Do not use disinfecting materials which contain hypochlorite (Javel water, Chlorox) or bleaching fluids. If possible, use non corrosive, neutral liquids.

When the instrument deck and carriers are cleaned using enzyme solutions such as DNAse and RNAse, make sure to thoroughly remove any remaining residue afterwards by wiping deck and carriers with deionized water in order to avoid aggressive substances left on the surface.

6.2 Decontamination using Gases

ATTENTION

For decontamination using gases, perform a risk analysis regarding increased service and maintenance requirements.

Should decontamination by fumigation be necessary, HAMILTON recommends using ethylene oxide.

NOTE

Be aware that ethylene oxide fumigation may increase service and maintenance requirements (O-rings exchange, greasing of spindles, etc.) and may make shorter maintenance intervals necessary.

HAMILTON does not carry out such fumigation procedures, please use a 3rd party contractor for such service.

Fumigation using hydrogen peroxide (H_2O_2) is possible, but leads to bleaching or discoloration of many instrument materials.

NOTE

Be aware that hydrogen peroxide fumigation may increase service and maintenance requirements (O-rings exchange, greasing of spindles, etc.) and may make shorter maintenance intervals necessary.

HAMILTON does not carry out such fumigation procedures, please use a 3rd party contractor for such service.

Do not use formaldehyde fumigation or chlorine oxides (chemical compounds of chlorine and oxygen such as bleach). They are not suitable for the ML STAR instrument because of chemical reaction and corrosion.

6.3 Decontamination using UV Light

HAMILTON offers an UV Light option for ML STAR instruments. We recommend UV radiation for 15 minutes and moving the arm left and right while the UV-lamp is on.

If using other UV radiation options, use wavelength, intensity and duration according to manufacturer´s instructions.

ATTENTION

For decontamination using other UV radiation options, perform a risk analysis regarding increased service and maintenance requirements.

ATTENTION

Do not expose yourself to the UV light(s), do not stare into the UV light source.

NOTE

Be aware that UV irradiation causes many synthetic materials to become brittle. This may increase service and maintenance requirements and may make shorter maintenance intervals necessary.

NOTE

The instrument cover shall be made of UV resistant acrylic glass, not of standard acrylic glass, due to its better resistance to UV irradiation.

7. Technical Specifications

7.1 Basic ML STAR Specifications

(for detailed information on configurations, see Section 8.3 Appendix C: Instrument Dimensions)			
Instrument Dimensions:	Width (x):	Height (z):	Depth (y):
STARlet	1124 mm (1450 mm with Multi Probe Head)	903 mm	795 mm (Autoload: 1006 mm)
STAR	1664 mm (1450 mm with Multi Probe Head)	903 mm	795 mm (Autoload: 1006 mm)
STARPlus	2160 mm (2486 mm with Multi Probe Head)	903 mm	795 mm (Autoload: 1006 mm)
Work Area Dimensions:	Width (x):	Height (z)[1]:	Depth (y):
STARlet	675 mm	136 mm	465 mm
STAR	1215 mm	136 mm	465 mm
STARPlus	1705 mm	136 mm	465 mm
Weight:	8 Pipetting Channels	Multi Probe Head and 8 Individual Pipetting Channels	
STARlet	135 kg	150 kg	
STAR	145 kg	160 kg	
STARPlus	205 kg	220 kg	
Deck Capacity:			
STARlet	30 Tracks (T) allow combinations of: • Maximum of 30 Tube Carriers (1 T) holding 24 or 32 Tubes per Carrier • Maximum of 5 Carriers (6 T) holding 5 Tip Racks or 5 Plate Positions per Carrier		
STAR	54 Tracks (T) allow combinations of: • Maximum of 9 Carriers (6 T) holding 5 Tip Racks or 5 Plate Positions per Carrier		
STARPlus	71 Tracks (T) allow combinations of: • Maximum of 11 Carriers (6 T) holding 5 Tip Racks or 5 Plate Positions per Carrier		
Modal Precision:	x-y-z positional accuracy of 0.1 mm		
Tip Sizes:	Low Volume: 10 µl, Intermediate Volume 50 µl, Standard Volume: 300 µl, High Volume: 1000 µl		
Needle Sizes:	Low Volume: 10 µl, Standard Volume: 300 µl, High Volume: 1000 µl, Needles available only for 1000 µl Single Pipetting Channels		
Power Consumption:	Standby Power Consumption: 100 VA Maximum Power Consumption 600 – 1000 VA, depending on configuration		

[1] The maximum height for labware which can be used on the deck is 140 mm

7.2 Pipetting Specifications

7.2.1 Disposable Tips with 1000 µl Pipetting Channels

Pipetting Specifications for Disposable Tips*	Disposable Tip Size	Volume	Trueness |R| (%)	Precision CV (%)
Individual 1000 µl Pipetting Channels	10 µl	0.5 µl	10.0%	6.0%
	10 µl	1 µl	5.0%	4.0%
	10 µl	5 µl	2.5%	1.5%
	10 µl	10 µl	1.5%	1%
	50 µl	0.5 µl	10.0%	6.0%
	50 µl	1 µl	5.0%	4.0%
	50 µl	5 µl	2.5%	1.5%
	50 µl	50 µl	2.0%	0.75%
	300 µl	10 µl	5.0%	2.0%
	300 µl	50 µl	2.0%	0.75%
	300 µl	200 µl	1.0%	0.75%
	1000 µl	10 µl	7.5%	3.5%
*** Test Criteria Available Upon Request**	1000 µl	100 µl	2.0%	0.75%
	1000 µl	1000 µl	1.0%	0.75%
	For pipetting of less than 10 µl, HAMILTON recommends 10 µl/ 50 µl Volume Disposable Tips to achieve the highest pipetting precision.			

7.2.2 Needles with 1000 µl Pipetting Channels

	Needle Size	Volume	Trueness |R| (%)	Precision CV (%)
Individual 1000 µl Pipetting Channels	10 µl	1 µl	5.0%	8.0%
	10 µl	5 µl	2.5%	2.0%
	10 µl	10 µl	1.5%	1.0%
	300 µl	5 µl	8.0%	8.0%
	300 µl	50 µl	2.0%	2.0%
	300 µl	200 µl	1.0%	1.0%
	1000 µl	50 µl	5.0%	3.0%
	1000 µl	100 µl	3.0%	2.0%
*** Test Criteria Available Upon Request**	1000 µl	1000 µl	2.0%	1.0%

7.2.3 Disposable Tips with 5 ml Pipetting Channels

Pipetting Specifications for 5 ml Disposable Tips*	Disposable Tip Size	Volume	Trueness |R| (%)	Precision CV (%)
Individual 5 ml Pipetting Channels	5 ml	50 µl	5.0%	2.5%
	5 ml	500 µl	2.0%	1.5%
	5 ml	1000 µl	1.5%	1.0%
*** Test Criteria Available Upon Request**	5 ml	5000 µl	1.0%	0.5%

7.2.4 Disposable Tips with 1000 µl CO-RE 96 Probe Head

CO-RE 96 Probe Head Pipetting Specifications for Disposable Tips*	Disposable Tip Size	Volume	Trueness |R| (%)	Precision CV (%)
1000 µl CO-RE 96 Probe Head	10 µl	1 µl	5.0%	5.0%
	10 µl	5 µl	2.5%	2.0%
Maximum pipetting volume: 1000 µl	10 µl	10 µl	1.5%	1.5%
	50 µl	1 µl	5.0%	5.0%
	50 µl	5 µl	2.5%	2.0%
	50 µl	50 µl	1.5%	1.0%
	300 µl	10 µl	3.0%	2.0%
	300 µl	50 µl	1.5%	1.0%
	300 µl	300 µl	1.0%	1.0%
	1000 µl	10 µl	7.5%	3.5%
	1000 µl	100 µl	2.0%	1.0%
*** Test Criteria Available Upon Request**	1000 µl	1000 µl	1.0%	1.0%
	For pipetting of less than 10 µl, HAMILTON recommends 10 µl/ 50 µl Volume Disposable Tips to achieve the highest pipetting precision.			

7.2.5 Disposable Tips with 50 µl CO-RE 384 Probe Head

CO-RE 384 Probe Head Pipetting Specifications for Disposable Tips*	Disposable Tip Size	Mode: Volume	Precision CV (%)
50 µl CORE 384 Probe Head	50 µl	Surface: 0.1 µl	8.0%
Maximum Pipetting Volume: 50 µl	50 µl	Surface: 0.5 µl	6.0%
	50 µl	Surface: 1 µl	3.5%
NOTE *Trueness is corrected in the Liquid Classes.*	50 µl	Jet: 1 µl	15.0%
	50 µl	Surface: 5 µl	3.0%
	50 µl	Jet: 5 µl	4.0%
	50 µl	Surface: 10 µl	2.0%
	50 µl	Jet: 10 µl	3.0%
*** Test criteria available upon request**	50 µl	Surface/Jet: 50 µl	2.0%

7.2.6 Disposable Tips with 50 µl CO-RE 384 Probe Head as 96 Probe Head

Disposable Tips with 50 µl CO-RE 384 Probe Head as 96 Probe Head	Disposable Tip Size	Mode: Volume	Precision CV (%)
Maximum Pipetting Volume: 300 µl	300 µl Rocket	2 µl	4.0%
	300 µl Rocket	5 µl	2.0%
	300 µl Rocket	10 µl	2.0%
	300 µl Rocket	100 µl	2.0%
	300 µl Rocket	300 µl	2.0%

7.2.7 Operating Data for Tips

Operating Data for Tips	Temperature Range:	+15 °C – +25 °C

7.2.8 Storage Data for Tips

Storage Data for Tips	Temperature:	Max. +55 °C

7.3 Liquid Level Detection

Liquid Level Detection:	Individual Pipetting Channels: • Capacitive Liquid Level Detection (cLLD) • Pressure Liquid Level Detection (pLLD) on Aspiration, cLLD on Dispense • Minimum Volume 10 µl, depending on container type CO-RE 96 Probe Head: • Capacitive Liquid Level Detection (cLLD) CO-RE 384 Probe Head: • Capacitive Liquid Level Detection (cLLD)

7.4 Throughput

Throughput:	8-Pipetting Channels: • To fill one 96-Well Microplate with 100 µl Samples (new tip for each sample): 320 seconds • Aliquot Reagent to a 96-Well Microplate (< 90 µl per well): 60 seconds CO-RE 96 Probe Head: • Replication of one 96-Well Microplate: 100 µl, new tips, with cLLD on Aspiration: 35 seconds • Reformatting of 4 – 96-Well Microplates to one 384 Well Microplate: 50 µl, new tips, with cLLD on Aspiration: 140 seconds CO-RE 384 Probe Head: • Replication of one 384-Well Microplate: 20 µl, new tips, with cLLD on Aspiration: 35 seconds • Reformatting of 4 – 384-Well Microplates to one 1536-Well Microplate: 10 µl, new tips, with cLLD on Aspiraton: 140 seconds
Labware:	All SBS standard plate types up to 1536 wells and most commercially available tube types
Carriers:	For all standard labware formats and according to customer requirements.

7.5 Operating Data

The maximum power consumption depends on the configuration of the instrument

Maximum Power Consumption 600 VA

Operating Data:	Voltage:	100/115 VAC / 230 VAC (± 10%)
	Frequency:	50/60 Hz
	Delayed Action Fuse :	100/115 VAC: 6.3 A (T6.3AL250) 230 VAC: 3.15 A (T3.15AL250)
	Overvoltage Category:	II
	Protection Class:	Class I
	Pollution Degree:	2
	Temperature Range:	15 ºC – 35 ºC (indoor use only)
	Relative Humidity:	30% – 85% (non-condensing, indoors)
	Noise Level:	< 65 dBA (regarding EN27779) < 46 dBA in standby mode
	Altitude:	Maximum 2000 meters above sea level
	Heat: (The Power Consumed will be Transferred to Heat)	(e.g. 600 Watts of Heat = 600 Joules/ Second)
	Indoor Use Only	

Maximum Power Consumption 1000 VA

	Voltage:	115 VAC / 230 VAC (± 10%)
	Frequency:	50/60 Hz
	Delayed Action Fuse	115 VAC: 10 A (T10AL250) 230 VAC: 5 A (T5AL250)
	Overvoltage Category:	II
	Protection Class:	Class I
	Pollution Degree:	2
	Temperature Range:	15 ºC – 35 ºC (indoor use only)
	Relative Humidity:	30% – 85% (non-condensing, indoors)
	Noise Level:	< 65 dBA (regarding EN27779) < 46 dBA in standby mode
	Altitude:	Maximum 2000 meters above sea level
	Heat: (The Power Consumed will be Transferred to Heat)	(e.g. 1000 Watts of Heat = 1000 Joules/ Second)
	Indoor Use Only	

7.6 Storage and Transportation

Transportation and Storage:	Temperature Range:	-25 ºC – +70 ºC
	Relative Humidity:	10% – 90% (non-condensing, indoors)

7.7 Lifetime

Lifetime	7 Years based on 8 hours/day, 5 days/week

7.8 Materials Used in ML STAR Line Instruments and Accessories

Materials used in the ML STAR Line Instruments and Accessories	Steel: 1.4301, 1.4305, 1.4310, 1.4435 Aluminum: AA 5083.0, various anodizing Bronze Brass Brazing solder: Zinc Plastics: PE, PP, PTFE, PEEK, FFPM, EPT, EPDM, FPM, NBR, POM, PCDF, FFKM, Hypalon, SI, Kapton, Polyamide PCBs

7.9 Communication

Communication	USB with Dual Processor Board Ethernet or USB with LAN Dual Processor Board

ATTENTION

The RS232 is no longer supported for communication. The connector is still available, however, without communication.

7.10 Computer Requirements (Computer not included in shipment)

PC Minimum Requirements	≥ 4GHz Processor, ≥ 4GB RAM, ≥ 250GB HD, ≥ 16x DVD DirectX 9 Graphics device Windows 7 Professional or Ultimate (32-bit or 64-bit) or Windows 10 Professional or Ultimate (32-bit or 64-bit)
Communication	≥ 5 USB ports, ≥ 2 RS232 ports

7.11 HAMILTON Standard PC Specifications

7.11.1 Windows 10

Compatible with: VENUS four	**Desktop Computer** **Windows 10, 64 Bit**
Configuration Overview	**OptiPlex XE2 Minitower** • 4th Gen Intel Core I7-4770S Processor (Quad Core HT, 3.10GHz Turbo, 8MB, w/ HD Graphics 4600) • 8GB (2x4GB) 1600Mhz DDR3 Non-ECC • 2x 500GB 3.5inch Serial ATA (7,200 rpm) Hard Drive – RAID1 • 16X Half Height DVD +/- RW Drive • 1GB AMD Radeon R6 240, 1GB Full Height (PP & DVI-I) • Broadcom NetXtreme 10/100/1000 PCIe Gigabit Networking Card, Full Height • 2 Ethernet Network Connectors • 4x USB 2.0 Connector x2 • 4x USB 3.0 Connector x2 • 4-Port, I/O Serial Card; in total 5 Serial Connectors • Wide Screen MONITOR LED 21.5" • Country specific keyboard and Mouse • Mousepad HAMILTON • Webcam 1.3MP USB 2.0 • Soundbar Stereo USB • Windows embedded standard 7/64 bit runtime (WS7P) "P" or Windows 10 • Microsoft Excel 2013 • 5 Yr. ProSupport and Next Business Day On-Site Service • 5 Yr. Data Protection – Keep Your Hard Drive • For additional RS232 Ports, PN 396247 (+2) or PN 396250 (+8) can be ordered • Country-specific Mains Power Cables (for computer and monitor) are not included and must be ordered separately • Additional programs must be obtained locally

7.12 Touch Screen Monitor Specifications for the ML STAR Line

Screen Size:	21.5"	**Technology:**	AccuTouch Five-Wire Resistive
Resolution:	1920 x 1080	**Colors:**	Up to 16.2 million
Interfaces:	1x VGA HD D-Sub	**Voltage:**	115 VAC / 230 VAC (± 10%)
	1x USB Type A	**Frequency:**	50/60 Hz
	1x serial RS-232	**Consumption:**	50 W
Brightness:	211 cd/m^2	**# of Touches:**	35 Million
Contrast:	550:1	**Certification:**	FCC Class B, CE, IC Class B, UL, TUV GS, VCCI Class B ITE, cUL, C-Tick Class B, RoHS
Viewing Angle:	± 70º Horizontal +75º/-60º Vertical	**Weight:**	7.9 kg

7.13 CR Needle Wash Station Specifications

Specification	Needle Capacity:	24 per Wash Station 8 per Chamber
	Wash Time (typical):	45 seconds / cycle
	Liquid Consumption (typical):	100 ml / cycle
	Carryover (typical):	1000 µl Needle, 100 µl: <3*10^{-6} 300 µl Needle, 100 µl: <2*10^{-6} 10 µl Needle, 10 µl: <5*10^{-6}
	Deck Capacity:	6 Tracks
	Maximum Units:	2 per Instrument
Operating Data	Maximum Power Consumption:	140 VA
	Voltage:	115 VAC / 230 VAC (± 10%)
	Frequency:	50/60 Hz
	Delayed Action Fuse:	115 VAC: 3.15A (T3.15AL250) 230 VAC: 1.6A (T1.6AL250)
	Installation Category:	II
	Pollution Degree:	2
	Temperature Range:	15 ºC – 35 ºC
	Relative Humidity:	30% – 85% (non-condensing, indoors)
	Altitude:	Maximum 2000 meters above sea level
	Indoor Use Only	
Storage and Transportation	Temperature Range:	-25 ºC – +70 ºC
	Relative Humidity:	10% – 90% (non-condensing, indoors)

7.14 CO-RE 96/384 Dual Wash Station Specifications

Specification	Wash Time (typical):	67 seconds / 2 cycles (96x10 tip) 71 seconds / 2 cycles (96x300 tip) 73 seconds / 2 cycles (384x30 tip)
	Liquid Consumption (typical):	880 ml / 2 cycles
	Carryover (typical):	
	2 Cycles:	10 µl Tips, 10 µl: < $3*10^{-6}$
		300 µl Tips, 300 µl: < $5*10^{-6}$
		50 µl Tips: < $1.8*10^{-5}$
	3 Cycles:	50 µl Tips: < $1*10^{-6}$
	Deck Capacity:	6 Tracks
	Maximum Units:	3 per Instrument
Operating Data	Power Consumption:	41 V / 100 VA (Maximum) Supplied by the ML STAR
	Temperature Range:	15 ºC – 35 ºC
	Relative Humidity:	30% – 85% (non-condensing, indoors)
	Altitude:	Maximum 2000 meters above sea level
	Indoor Use Only	
Storage and Transportation	Temperature Range:	-25 ºC to +70 ºC
	Relative Humidity:	10% – 90% (non-condensing, indoors)

7.15 iSWAP Specifications

Plate Format:	Microplate Footprint Plate Height 5 mm – 43 mm		
Positioning:	Reproducibility: X/Y/Z = ± 0.5 mm		
Movement Range: (on a STAR 8/iSWAP Instrument)	**Minimum Absolute Position**	**Maximum Absolute Position**	**Remarks**
X	-206 mm	+1578 mm	at x_{min} 58 mm space between microplate and deck at x_{max} half plate on the deck
Y	-185 mm	+605 mm	
Z	+100 mm +0 mm	+282 mm +282 mm	iSWAP PN 182600 rev. 00 – 02 iSWAP PN 182600 rev. 03 and iSWAP Landscape PN 190220
Gripper Opening:	72 mm 72 mm	108 mm 132 mm	iSWAP PN 182600 rev. 00 – 03 iSWAP Landscape PN 190220
Gripping Force:	5 N – 16 N (default 9 N)		iSWAP PN 182600 rev. 00 – 03 iSWAP Landscape PN 190220
Transport Mass:	300g filled Deep Well Plate		
No restriction of random access range for 4, 8, 12, and 16 pipetting channel on ML STAR.			
Maximum Modules per System	1 iSWAP per System		

7.16 Tube Gripper Specifications

Tube Sizes:	Tube diameter from 8 mm to 20 mm Tube height ≤ 120 mm		
Modal Precision:	X-, Y-, Z- positional accuracy of 0.1 mm (measured on tube-gripper)		
Movement Range:			
X	Reaches the same X-coordinate as the single pipetting channels (e.g. on a STAR 8x 1000µl-pipetting channel instruments)		
Y	Reaches all tube positions of the sample carriers: SMP-CAR-32 and SMP-CAR-24 (on a ML STAR instrument with 6x 1000µl-pipetting channels)		
Z	**Minimum Absolute Position**	**Maximum Absolute Position**	**Remarks**
	+54.2 mm	+254.2 mm	Measured from the deck work surface to the gripping point on a tube
Gripper Opening:	**Minimum Absolute Position**	**Maximum Absolute Position**	
	5.5 mm	22 mm	
Transport Mass:	200 g		
Maximum Modules per System	1 Tube Gripper per System		

7.17 CO-RE Gripper 1000 µl Specifications

Labware Format:	Microplate Footprint Plate Height 5 mm – 43 mm		
Positioning:	Reproducibility: X/Y/Z = ± 0.5 mm		
Movement Range:			
x	Track 1 – n (depending on instrument type)		
y	Depending on # of pipetting channels and used front channel		
z	Lowest position = 15 mm over the deck work surface		
Gripper Opening:	**Minimum Opening**	**Maximum Opening**	**Arm Type**
	9 mm	Dependent upon the travel range on the pipetting arm	Modular arm Modular arm MPH
	18 mm	Dependent upon the travel range on the pipetting arm	MPH/iSWAP arm iSWAP arm
Gripping Force:	5 N – 16 N (default 9 N)		
Transport Mass:	300 g filled Deep Well Plate		
Maximum Modules per System	2 Channels per System		

7.18 CO-RE Gripper 5 ml Specifications

Labware Format:	Microplate Footprint Plate Height 5 mm – 43 mm		
Positioning:	Reproducibility X/Y/Z = ± 0.5 mm		
Movement Range:			
X	Track 1 – n (depending on instrument type)		
Y	Depending on # of pipetting channels and used front channel		
Z	Lowest position = 46 mm over the deck work surface		
Gripper Opening:	**Minimum Opening**	**Maximum Opening**	**Arm Type**
	18 mm	Dependent upon the travel range on the pipetting arm	Modular arm Modular arm MPH
	36 mm	Dependent upon the travel range on the pipetting arm	MPH/iSWAP arm iSWAP arm
Gripping Force:	5 N – 16 N (default 9 N)		
Transport Mass:	300 g filled Deep Well Plate		
Maximum Modules per System	2 Channels per System		

7.19 HAMILTON SBS Centrifuge Specifications

For detailed information about the HAMILTON SBS Centrifuge, refer to the HAMILTON SBS Centrifuge Operator's Manual.

7.20 48VDC Power Box Specifications

Features	**Specifications**
Dimensions	370 mm x 100 mm x 210 mm
Weight	600 g
Input Voltage	115 VAC / 230 VAC (± 10%)
Frequency	50/60 Hz
Maximum Power Consumption	480 VA
Primary Fuse	240 VAC: 4AT 250 V 5x20 120 VAC: 10AT 250 V 5x20
Secondary Fuse	10 AT 250 V 5x20
Pollution Degree	2
Noise Level	<62dBA
Lifetime	6 years
Over-Voltage Category Specification	II

7.21 Crystal Vacuum System (CVS) Specifications

CVS Carrier Dimensions:	Width (x)	Height (z)	Depth (y)
	155 mm	112 mm	555 mm
Vacuum Pump Dimensions:	Width (x)	Height (z)	Depth (y)
	242 mm	245 mm	257 mm
Waste Container:	Diameter: 155 mm	Height: 380 mm	
Weight of CVS:	4 kg		
Weight of Vacuum Pump:	13.5 kg		
Operating Data:	Maximum Power Consumption:	530 W (230 VAC)	
	Voltage:	115 VAC / 230 VAC (± 10%)	
	Frequency:	50/60 Hz	
	Temperature Range:	10 ºC – 40 ºC	
	Relative Humidity:	30% - 85%	
	Maximum HHS Shaker Speed:	2000 rpm	

For detailed information about the Multiflex Cooling-Heating Module, refer to the Multiflex Cooling-Heating Module Operator's Manual.

7.22 Multiflex Heating Module Specifications

Dimensions (without carrier base):	Width (x)	Height (z)	Depth (y)
	135 mm	120 mm	190 mm
Weight:	0.5 kg		
Operating Data:	Maximum Power Consumption:	50 W	
	Voltage:	41 VDC	
	Temperature Range:	Ambient plus 5 ºC to 65 ºC	
	Temperature Accuracy at 30 ºC – 65 ºC	< -0.5/+1.5 ºC	
	Temperature Accuracy	± 1 ºC	

7.23 Multiflex Cooling Module Specifications

Dimensions (without carrier base):	Width (x)	Height (z)	Depth (y)
	135 mm	120 mm	190 mm
Weight:	1.2 kg		
Operating Data:	Maximum Power Consumption:	50 W	
	Voltage:	41 VDC	
	Temperature Range:	15 ºC to 0 ºC	
	Temperature Accuracy:	± 1 ºC	

7.24 Tube Twister Channel and Decapper Module Specifications

Tube Twister Channel	Supported tube sizes	Vial diameters: from 15 mm to 38 mm Vial height: from 50 mm to 120 mm
	Transport area and positioning	The complete instrument work area, with limited access depending on instrument configuration. Maximal 4 channels. Minimal grip height 50 mm above deck Positioning X/Y/Z: ≤ ±0.35 mm
	Mixing (twisting)	From 0 to 1500 rpm ± 5%
	Barcode reading	As defined in Section 7.27.5 Sample Barcodes Additional feature: Align barcode
	Transport Mass 50 ml Falcon tubes	50 g contents (+tube weight)

7.24.1 Barcode Reader Specifications

Type:	Class II Laser Diode
Wavelength:	655 nm
Pulse Duration:	<420 µs
Maximum Output (peak):	1.7 mW
Nominal Ocular Hazard Distance (NOHD):	≤ 0 mm
Applied Standard:	IEC/EN-60825-1:2014

7.24.2 Additional features for the Tube Twister Channel in combination with a Decapper Module:

Tube Twister Channel / Decapper Module	De-capping / Recapping	De-capping / recapping of screw top vials
	Decapper Module Small: Supported Tube Sizes with:	Vial diameters: from 15 mm to 27 mm Cap diameters: from 18 mm to 32 mm Vial height: from 50 mm to 120 mm (including cap)
	Decapper Module Large: Supported Tube Sizes with:	Vial diameters: from 27 mm to 38 mm Cap diameters: from 32 mm to 44 mm Vial height: from 50 mm to 120 mm (including cap)
Maximum Modules per System	2 or 4 Twister Channels per System No more and no less, all 2 or 4 channels are on separate iSWAP Arm, no combination with other components on the same arm.	

7.25 Autoload Option: Barcode and Reader Specifications

Carriers, containers, racks and tip racks can be identified by a barcode, which a reader, mounted on the Autoload slide, scans. The system must allow specification of ranges (barcode mask) for plausibility checking of barcode information.

7.25.1 Barcode Symbologies

The following barcode symbologies can be read by the Autoload:

ISBT Standard	Code 128 (Subset B and C)
Code 39	Codabar
Code 2 of 5 Interleaved	UPC A/E
JAN/EAN 8	

For the highest reading safety HAMILTON recommends to:

1. Use the barcode type Code128 (subset B and C).
2. Disable the unused barcode types in the configuration editor of the User Software (refer to the VENUS Software Programmer's Manual).
3. Define a barcode mask via the Labware Editor of the User Software (refer to the VENUS Software Programmer's Manual).

7.25.2 Reading Accuracy

The rate of inaccurate readings of sample plates and container bar codes is less than 1 ppm.

The above mentioned specification are valid under the following conditions:

- Barcode Symbology Module: ISBT standard
- Code Density: 0.0065 inches (0.1651 mm)
- Print Quality see Section 7.27.4 Barcode Specifications
- Recognized errors are defined as an accurate reading

7.25.3 Barcode Scanner Specifications

Type:	Class I Laser Diode
Wavelength:	655 nm
Pulse Duration:	continuously
Applied Standard:	IEC/EN 60825-1:2014

7.25.4 Barcode Specifications

Type:	Black bars and white background	
Length of String:	Maximum 20 characters excluding start, stop and check characters, depending on the barcode length (see label dimensions).	
Code Density, Tolerance:	Minimum module width (x dimension) including a print tolerance: ≥ 0.0065 inches (0.1651 mm) Maximum module width (x dimension) including a print tolerance: ≤ 0.02 inches (0.508 mm) Best reading performance with x dimension ≥ 0.01 inches (0.254 mm)	
Check Character:	ISBT Standard	One character
	Code 128	One character
	Code 39	None
	Codabar	None
	Code 2 of 5 Interleaved	None
	UPC A/E	One character
Quiet Zone:	≥ 10 Times the x dimension, but at least 3 mm	
Print Quality:	The barcode print must be of a high quality. A printed barcode with an ANSI/ CEN/ ISO grade A or B is required. Offset, typographic, intaglio and flexographic printing are suitable. Mechanical dot matrix and thermos matrix printing are not suitable. The surface may be treated, sealed or plastic-coated.	

7.25.5 Sample Barcodes

Barcode Specifications:

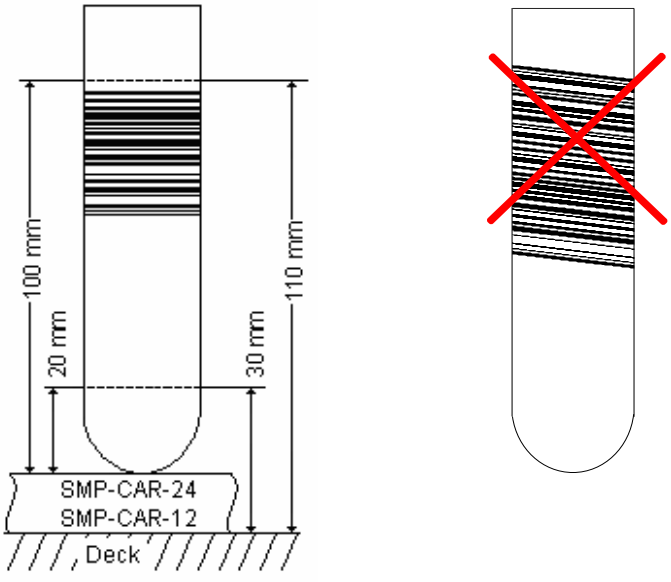
For General Barcode Specifications, see Section 7.27.4 Barcode Specifications.

Dimension		Minimum	Maximum
A	Label length	-	80 mm
B	Code length	-	74 mm
C	Quiet zone	3 mm	
D	Label width	12 mm	-
E	Code width	12 mm	-
F	Distance from the barcode to the label edge	-	1 mm

Positioning Barcode Labels:

The label must be glued within a range of between 20 mm to 100 mm from the bottom of the tube.

The label must fit tightly at an angle of 90° to the tube.



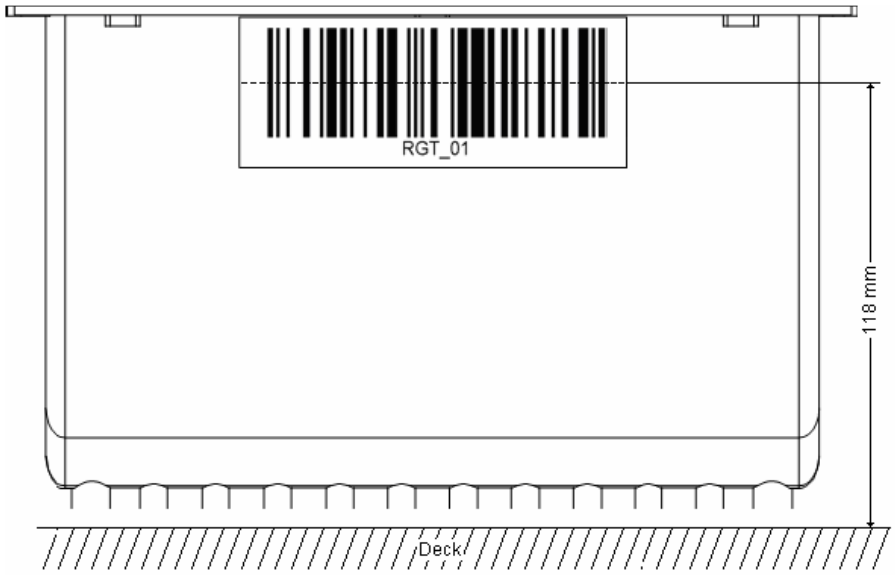
The label must fit tightly over its entire length.

7.25.6 Reagent Barcodes

Proposed Barcode Mask: **RGT_mm**	**RGT**: Reagent _: Separator (underline) **mm**: Reagent number 1.....99
Label Specification: For General Barcode Specifications, see Section 7.27.4 Barcode Specifications.	

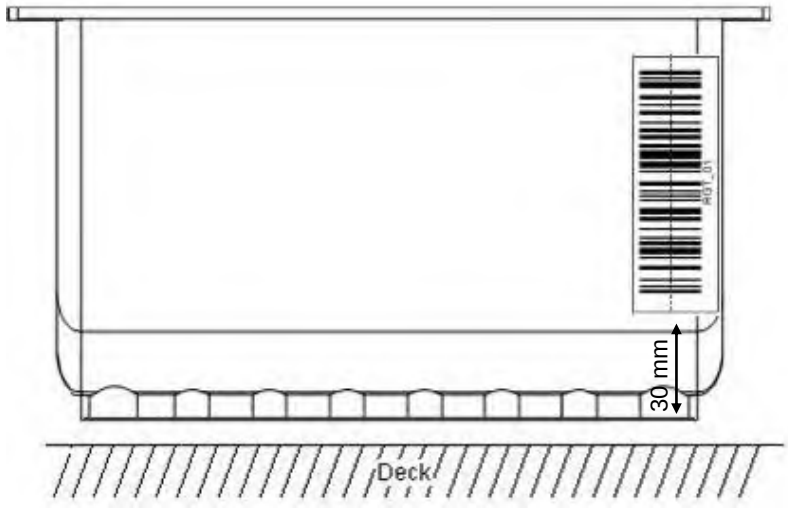
Dimension		Minimum	Maximum
A	Label length	-	66 mm
B	Code length	-	60 mm
C	Quiet zone	3 mm	-
D	Label width	15 mm	-
E	Code width	12 mm	-
F	Distance from barcode to the edge of the label	-	1 mm

Positioning of the Barcode Labels:



The label must be positioned on the upper edge, in the middle of the container.
The label must fit tightly over its entire length.

Positioning of the Barcode Label on a 50 ml Reagent Trough:

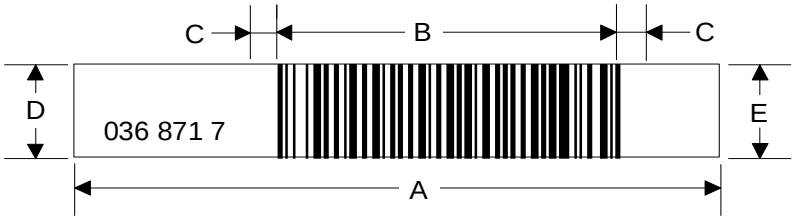


The label must be glued within a range between 30 mm to 110 mm from the deck.

7.25.7 Plate Barcodes

Label Specifications:

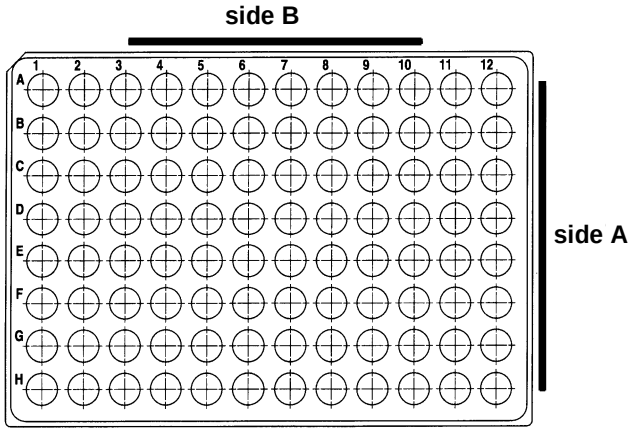
For General Barcode Specifications, see Section 7.27.4 Barcode Specifications.



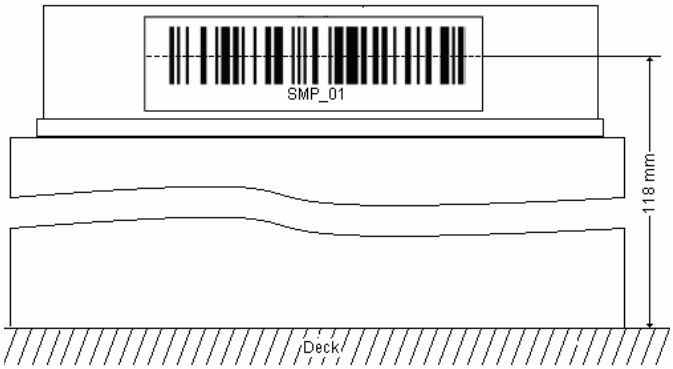
Dimension		Minimum	Maximum
A	Label length	-	66 mm
B	Code length	-	30 mm
C	Quiet zone	3 mm	-
D	Label width	10 mm	-
E	Code width	7 mm	-
	Distance from the barcode to the label edge (if necessary)	-	1 mm

Positioning Barcode Labels:

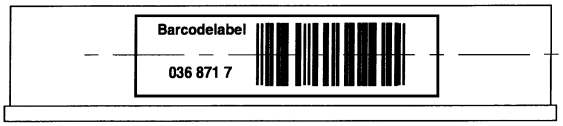
The plate barcode must fit on side A or side B of the plate.



The barcode labels must be positioned in the middle of the plate.



The barcode label must be centered and parallel to the edge of the plate.



The barcode label must not protrude above or below the edge of the plate.

8. Appendices

8.1 Appendix A: Ordering Information

NOTE

This list does not claim to be complete. Please contact your HAMILTON representative for more information.

8.1.1 Standard Tips

8.1.1.1 50 µl Standard Tips

50 µl Standard Tips (0.5 µl – 50 µl)		
Part Number	**Description**	**Image**
235966	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Conductive Case of 5760 tips (blister pack)	
235948	**50 µl CO-RE tips** with filter 0.5 µl – 50 µl Conductive Case of 5760 tips (blister pack)	
235978	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235979	**50 µl CO-RE tips** with filter 0.5 µl – 50 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235829	**50 µl CO-RE tips** with filter 0.5 µl – 50 µl Clear tip Case of 5760 tips (blister pack)	
235831	**50 µl CO-RE tips** with filter 0.5 µl – 50 µl Clear tip, sterilized Case of 5760 tips (blister pack)	
235836	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Clear tip Case of 5760 tips (blister pack)	
235837	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Clear tip, sterilized Case of 5760 tips (blister pack)	

50 µl Standard Tips (0.5 µl – 50 µl)		
Part Number	**Description**	**Image**
235964	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Clear tip Case of 11520 tips (NTR)	
235947	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Conductive Case of 11520 tips (NTR)	
235987	**50 µl CO-RE tips** without filter 0.5 µl – 50 µl Conductive, sterilized Case of 11520 tips (NTR)	

8.1.1.2 300 µl Standard Tips

300 µl Standard Tips (10 µl – 300 µl)		
Part Number	**Description**	**Image**
235902	**Standard CO-RE tips** without filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	
235937	**Standard CO-RE tips** without filter 10 µl – 300 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235985	**Standard CO-RE tips** without filter 10 µl – 300 µl Conductive, sterilized Case of 11520 tips (NTR)	
235950	**Standard CO-RE tips** without filter 10 µl – 300 µl Conductive Case of 11520 tips (NTR)	
235830	**Standard CO-RE tips** without filter 10 µl – 300 µl Clear tip Case of 5760 tips (blister pack)	
235832	**Standard CO-RE tips** without filter 10 µl – 300 µl Clear tip, sterilized Case of 5760 tips (blister pack)	
235834	**Standard CO-RE tips** without filter 10 µl – 300 µl Clear tip Case of 5760 tips (blister pack)	
235835	**Standard CO-RE tips** without filter 10 µl – 300 µl Clear tip, sterilized Case of 5760 tips (blister pack)	

300 µl Standard Tips (10 µl – 300 µl)		
Part Number	**Description**	**Image**
235965	**Standard CO-RE tips** without filter 10 µl – 300 µl Clear tip Case of 11520 tips (NTR)	
235938	**Standard CO-RE tips** with filter 10 µl – 300 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235903	**Standard CO-RE tips** with filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	

8.1.1.3 1000 µl Standard Tips

1000 µl Standard Tips (10 µl – 1000 µl)		
Part Number	**Description**	**Image**
235904	**High CO-RE tips** without filter 10 µl – 1000 µl Conductive Case of 3840 tips (blister pack)	
235939	**High CO-RE tips** without filter 10 µl – 1000 µl Conductive, sterilized Case of 3840 tips (blister pack)	
235905	**High CO-RE tips** with filter 10 µl – 1000 µl Conductive Case of 3840 tips (blister pack)	
235940	**High CO-RE tips** with filter 10 µl – 1000 µl Conductive, sterilized Case of 3840 tips (blister pack)	
235820	**High CO-RE tip** without filter 10 µl – 1000 µl Clear Case of 3840 tips (blister pack)	
235821	**High CO-RE tip** without filter 10 µl – 1000 µl Clear, sterilized Case of 3840 tips (blister pack)	
235822	**High CO-RE tip** without filter 10 µl – 1000 µl Clear Case of 3840 tips (blister pack)	
235823	**High CO-RE tip** without filter 10 µl – 1000 µl Clear, sterilized Case of 3840 tips (blister pack)	

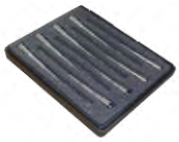
8.1.1.4 5 ml Standard Tips

5 ml Standard Tips (50 µl – 5000 µl / (4000 µl with filter))		
Part Number	**Description**	**Image**
184020	**5 ml CO-RE tips** without filter 50 µl – 5000 µl Conductive Case of 720 tips (blister pack)	
184022	**5 ml CO-RE tips** without filter 50 µl – 5000 µl Conductive, sterilized, Case of 720 tips (blister pack)	
194050	**5 ml CO-RE tips** without filter 50 µl – 5000 µl Conductive Case of 96 tips, 4 tips individually wrapped (blister pack)	
184021	**4 ml CO-RE tip** with filter 50 µl – 4000 µl Conductive Case of 720 tips (blister pack)	
184023	**4 ml CO-RE tip** with filter 50 µl – 4000 µl Conductive, sterilized Case of 720 tips (blister pack)	
194053	**4 ml CO-RE tip** with filter 50 µl – 4000 µl Conductive Case of 96 tips, 4 tips individually wrapped (blister pack)	

8.1.2 Steel Needles for 1000 µl Single Channels

Steel Needles for 1000 µl Single Channels		
Part Number	**Description**	**Image**
235930	**1000 µl NEEDLE SET CR** 50 µl – 1000 µl Set of 8x 1000 µl needles, CR Wash Station	
235931	**300 µl NEEDLE SET CR** 5 µl – 300 µl Set of 8x 300 µl needles, CR Wash Station	
235932	**10 µl NEEDLE SET CR** 1 µl – 10 µl Set of 8x 10 µl needles, CR Wash Station	
187290	**NEEDLE SERVICE KIT** Contains a Needle pick-up tool to remove needles individually from the wash station.	

8.1.3 Teaching Needles

Teaching Needles for Single Channels		
Part Number	**Description**	**Image**
182136	**SET OF 8 TEACHING NEEDLES** Used for the maintenance to check the pressure tightness of the 1000 µl pipetting-channels	
182176	**TEACHING NEEDLE** 1 needle for 1000 µl pipetting channels, used for labware teaching (Set of 8 PN 182136)	
184184	**TEACHING NEEDLE** 1 needle for 5 ml pipetting channels, used for labware teaching	
187290	**NEEDLE SERVICE KIT** Contains a Needle pick-up tool to remove needles individually from the wash station.	

8.1.4 10 µl Low Volume Tips

10 µl Low Volume Tips (0.5 µl – 10 µl)		
Part Number	**Description**	**Image**
235900	**10 µl CO-RE tips** without filter 0.5 µl – 10 µl Conductive Case of 5760 tips (blister pack)	
235901	**10 µl CO-RE tips** with filter 0.5 µl – 10 µl Conductive Case of 5760 tips (blister pack)	
235935	**10 µl CO-RE tips** without filter 0.5 µl – 10 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235936	**10 µl CO-RE tips** with filter 0.5 µl – 10 µl Conductive, sterilized Case of 5760 tips (blister pack)	
235949	**10 µl CO-RE tips** without filter 0.5 µl – 10 µl Conductive, in a Nested Tip Box Case of 11520 tips (NTR)	

10 µl Low Volume Tips (0.5 µl – 10 µl)		
Part Number	**Description**	**Image**
235983	**10 µl CO-RE tips** without filter 0.5 µl – 10 µl Conductive, sterilized Case of 11520 tips (NTR)	
235971	**10 µl CO-RE tips** without filter 0.5 µl – 10 µl Clear tip Case of 11520 tips (NTR)	

8.1.5 50 µl Standard Tips for CO-RE 384 Probe Head

50 µl Standard Tips for CO-RE 384 Probe Head (0.5 µl – 50 µl)		
Part Number	**Description**	**Image**
235989	**50 µl 384 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Conductive Case of 7680 tips, 384 tips per rack (NTR)	
235694	**50 µl 384 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Conductive, sterilized Case of 7680 tips, 384 tips per rack (NTR)	
235446	**50 µl 384 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Clear tips Case of 7680 tips, 384 tips per rack (NTR)	
235824	**50 µl 384 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Clear tips, sterilized Case of 7680 tips, 384 tips per rack (NTR)	
235993	**50 µl 96 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Conductive Case of 1920 tips, 96 tips per rack (NTR)	
235695	**50 µl 96 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Conductive, sterilized Case of 1920 tips, 96 tips per rack (NTR)	
235447	**50 µl 96 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Clear tips Case of 1920 tips, 96 tips per rack (NTR)	
235825	**50 µl 96 Probe Head CO-RE tips** without filter 0.5 µl – 50 µl Clear tips, sterilized Case of 1920 tips, 96 tips per rack (NTR)	

8.1.6 250 µl Robotic Piercing Tips for CO-RE 96-Probe Heads

250 µl Robotic Piercing Tips for CO-RE 96-Probe Heads (2.0 µl – 250 µl (150 µl with filter)		
Part Number	**Description**	**Image**
235805	**250 µl Robotic Piercing Tips** without filter 2.0 µl – 250 µl Conductive Case of 3840 tips (blister pack)	
235659	**250 µl Robotic Piercing Tips** without filter 2.0 µl – 250 µl Conductive, sterilized Case of 3840 tips (blister pack)	
235658	**250 µl Robotic Piercing Tips** with filter 2.0 µl – 150 µl Conductive Case of 3840 tips (blister pack)	
235649	**250 µl Robotic Piercing Tips** with filter 2.0 µl – 150 µl Conductive, sterilized Case of 3840 tips (blister pack)	

8.1.7 300 µl Special Tips Extra-Long and Slim Tips

300 µl SPECIAL TIPS Extra-Long and Slim Tips (10 µl – 300 µl)		
Part Number	**Description**	**Image**
235806	**300 µl Extra Long and Slim Tips** without filter 10 µl – 300 µl Conductive Case of 3840 tips (blister pack)	
235648	**300 µl Extra Long and Slim Tips** without filter 10 µl – 300 µl Conductive, sterilized Case of 3840 tips (blister pack)	
235647	**300 µl Extra Long and Slim Tips** with filter 10 µl – 300 µl Conductive Case of 3840 tips (blister pack)	
235646	**300 µl Extra Long and Slim Tips** with filter 10 µl – 300 µl Conductive, sterilized Case of 3840 tips (blister pack)	

8.1.8 300 µl Rocket Tips for CO-RE 384 Probe Head

Rocket Tips for CO-RE 384 Probe Head (2 µl – 300 µl)		
Part Number	**Description**	**Image**
235974	**300 µl Rocket Tips** without filter 2 µl – 300 µl Conductive, rack, containing 96 tips Case of 4800 tips (blister pack)	

8.1.9 300 µl Wide Bore Tips

300 µl Wide Bore Tips (10 µl – 300 µl)		
Part Number	**Description**	**Image**
235452	**300 µl Wide bore tips, 0.71 mm orifice** with filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	
235688	**300 µl Wide bore tips, 0.71 mm orifice** without filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	
235449	**300 µl Wide bore tips, 1.55 mm orifice** with filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	
235451	**300 µl Wide bore tips, 1.55 mm orifice** without filter 10 µl – 300 µl Conductive Case of 5760 tips (blister pack)	

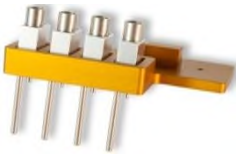
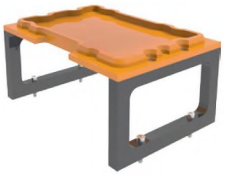
8.1.10 1000 µl Wide Bore HIGH Tips

1000 µl Wide Bore HIGH Tips for CO-RE 96 Probe Head (10 µl – 1000 µl)		
Part Number	**Description**	**Image**
235678	**1000 µl Wide Bore Tips, 1.20 mm orifice** with filter 10 µl – 1000 µl Conductive Case of 3840 tips (blister pack)	
235679	**1000 µl Wide Bore Tips, 1.20 mm orifice** without filter 10 µl – 1000 µl Conductive Case of 3840 tips (blister pack)	

235441	**1000 µl Wide Bore Tips, 3.20 mm orifice** with filter 10 µl – 1000 µl Conductive, Sterilized Case of 3840 tips (blister pack)	
235544	**1000 µl Wide Bore Tips, 3.20 mm orifice** without filter 10 µl – 1000 µl Conductive Case of 3840 tips (blister pack)	

8.1.11 FlipTube System

Reagent Tubes		
Part Number	**Description**	**Image**
235454	**FlipTubes** 1.5 ml reagent tubes with a lid attached which can close the tube tightly.	
235692	**FlipTubes Purity Tested** 1.5 ml reagent tubes with a lid attached which can close the tube tightly.	
235693	**FlipTubes Biological Purity Plus** 1.5 ml reagent tubes with a lid attached which can close the tube tightly.	
809032	**FlipTube Tool**	

809306	**FlipTube Tool Park Station on Waste**	
199085	**HAMILTON HEATER SHAKER 3 for FlipTube Rack** This HHS is equipped with a built-in adapter for a FlipTube Rack. Shaker orbit: 3 mm Shaking speed: 100 – 1800/2400 rpm Temperature control: RT+5 ºC – 105 ºC	
814269	**FlipTube H/C Block** 24 position block for cooling/heating devices. It must be used with FlipTube Racks. This block is not compatible with the HAMILTON Heater Shaker.	
814275	**MULTIFLEX FlipTube Rack Park Position** The Multiflex Rack Park Position is used for a correct alignment after the transport 24 position block for cooling/heating devices. It must be used with FlipTube Racks.	
814270	**FlipTube Rack** The FlipTube Rack is a 24 position SBS format rack for handling and transport of HAMILTON FlipTube A (PN 235454, PN 235692, PN 235693) between different functional devices such as heaters, coolers, shakers, centrifuges and others. **Set of 5 racks. Material: Polycarbonate.**	
809030	**SMP_CAR_32FT** Sample Carrier for 32 FlipTubes (1T)	

8.1.12 Disposable Waste Bags

Disposable Waste Bags		
Part Number	**Description**	**Image**
185319	**Plastic Waste Chute without Biohazard Labeling** (roll of 10 pcs), 460 mm x 700 mm, PE This chute is used with Waste Container (PN 281520)	
199201	**Plastic Waste Chute "Biohazard" with Biohazard Labeling** (roll of 10 pcs), 460 mm x 700 mm, PE This chute is used with Waste Container (PN 281520)	
199202	**Waste Bag without Biohazard Labeling** (roll of 25 pcs), 460 mm x 500 mm, PE	
199203	**Waste Bags "Biohazard" with Biohazard Labeling** (roll of 25 pcs), 460 mm x 500 mm, PE	
281520	**Waste Container "Biohazard"** (Pack of 1 box), combustible This container is used with Waste Chute (PN 199201 or PN 185319)	

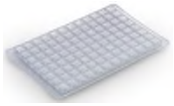
8.1.13 Reagent Containers

Reagent Containers		
Part Number	**Description**	**Image**
56694-01	**REAGENT CONTAINER 60 ml** Set of 28 transparent, self-standing with lid, for carrier RGT_CAR_5R60 (PN 194057 1T)	
56694-02	**REAGENT CONTAINER 60 ml** Set of 28 transparent, self-standing with lid, for carrier RGT_CAR_5R60 (PN 194057 1T)	
194051	**REAGENT CONTAINER 60 ml** Set of 28 transparent, self-standing, barcodes included in box, w/o lid, individually wrapped for carrier RGT_CAR_5R60 (PN 194057 1T)	

Reagent Containers		
Part Number	**Description**	**Image**
182703	**REAGENT CONTAINER 120 ml** Set of 12 transparent for carrier RGT_CAR_3R (PN 185290 1T)	
194052	**REAGENT CONTAINER 120 ml** Set of 12 transparent, self-standing, barcodes included in box, 120 ml for carrier RGT_CAR_3R (PN 194058 1T)	
56695-01	**REAGENT CONTAINER 200 ml** Set of 10 transparent, self-standing with lid, for carrier RGT_CAR_4R200 (PN 185436)	
56695-02	**REAGENT CONTAINER 200 ml** Set of 10, black, self-standing with lid, for carrier RGT_CAR_4R200 (PN 185436)	

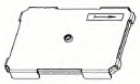
8.1.14 Deep Well Plates

Deep Well Plates		
Part Number	**Description**	**Image**
6471-01	**DEEP WELL BLOCK 1.2 ml, PP** 96 well block, polypropylene, 1.2 ml volume per well, round wells. W: 126.3 mm; D: 84.7 mm; H: 40.6 mm 32 pcs/case, 4 plates are wrapped	
6472-01	**DEEP WELL BLOCK 1.2 ml, PS** 96 well block, polystyrene, transparent, 1.2 ml volume per well, round wells. W: 127.5 mm; D: 85.5 mm; H: 41 mm 32 pcs/case, 4 plates are wrapped	
235655	**DEEP WELL BLOCK 1.2 ml, PS** 96 well block, polystyrene, transparent, with barcode 1.2 ml volume per well, round wells. W: 127.5 mm; D: 85.5 mm; H: 41 mm 32 pcs/case, 4 plates are wrapped	
6473-01	**DEEP WELL BLOCK 2.2 ml, PP** 96 well block, polypropylene, 2.2 ml volume per well, square wells, round bottom. W: 126.3 mm; D: 84.7 mm; H: 40.6 mm 32 pcs/case, 4 plates are wrapped	

Deep Well Plates		
Part Number	**Description**	**Image**
235656	**DEEP WELL BLOCK 2.2 ml, PP** 96 well block, polypropylene, with barcode, 2.2 ml volume per well, square wells, round bottom. W: 126.3 mm; D: 84.7 mm; H: 40.6 mm 32 pcs/case, 4 plates are wrapped	
6474-01	**SEALING MAT** Sealing mat for 2.2 ml blocks, ethylenevinylacetate W: 121.5 mm; D: 78.8 mm; H: 4.68 mm 50 pcs/case, 10 pcs/bag	

8.1.15 Plate Carrier

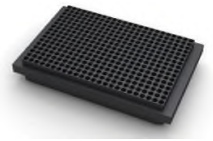
Plate Carrier		
Part Number	**Description**	**Image**
182035	**PLT_CAR_L5PCR384** Carrier for 5x 384 PCR plates	
182065	**PLT_CAR_P3AC** Carrier for 3x Deep Well Plates, portrait orientation (6T)	
182070	**PLT_CAR_L5PCR** Carrier for 5x 96-well PCR plates (6T)	
182075	**PLT_CAR_P3MD** Carrier for 3x 96/384 well plates, portrait orientation (6T)	
182090	**PLT_CAR-L5AC** Carrier for 5x 96 Deep Well Plates or for 5x 384 tip racks (e.g. 384HEAD_384TIPS_50 µl) (6T)	
191287	**PLT-CAR-L4HD** Carrier for 4x 1536-well plates (6T)	
182190	**PLT-CAR-P3HD** Carrier for 3 x 1536-well plates, portrait orientation (6T)	
182365	**PLT_CAR_L5MD** Carrier for 5x 96/384-well plates (6T)	
185295	**PLT_CAR_L5AC PINNED** Carrier for 5x HAMILTON DWP (6T)	

Plate Carrier		
Part Number	**Description**	**Image**
185330	**PLT_CAR_L4ST (4x8 MTP)** Stacker carrier for 4 x 8 MTP (7T) iSWAP loadable on one dedicated side. The stacks may be 5 plates high, or 8 plates high if the Lever Plate is removed.	
185340	**PLT_CAR_L4ST (4x5 MTP)** Stacker carrier for 4 x 5 MTP (7T) iSWAP loadable on one dedicated side.	
182735	**PLT_CORE_COVER** Lid to cover MTP on standard carriers	
187223	**ANTI-EVAPORATION LID** Lid for minimizing evaporation of liquid in MTP. Typically used for protein crystallization plate preparation. Can be handled by the iSWAP or CO-RE Gripper. Requires PN 188054APE PLT-CAR-L4-Crystal.	
182712	**FRAME FOR FILTER PLATE** To place filter plates on archive carriers	

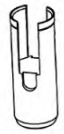
8.1.16 Reagent Carrier

Reagent Carrier		
Part Number	**Description**	**Image**
194057	**RGT_CAR_5R60** Reagent Carrier for 5x reagent troughs (1T) For reagent container PN 194051, 60 ml	
187239	**RGT_CAR_4R100** Reagent Carrier for 4x reagent troughs 100 ml (1T) For reagent container with lids (PN 187236) or reagent container without lids (PN 187244), 100 ml	
182080	**RGT_CAR_12R** Reagent Carrier for 12x reagent troughs 100 ml (1T) For reagent container PN 137257, 100 ml	
185290	**RGT_CAR_3R** Reagent Carrier for 3x reagent troughs 120 ml (1T) For reagent container PN 182703, 120 ml	
194058	**RGT_CAR_3R120** Reagent Carrier for 5x reagent troughs 120 ml (1T) For reagent container PN 194052, 120 ml	
185436	**RGT_CAR_4R200** Reagent Carrier for 4x reagent troughs 200 ml (1T) For reagent container PN 56695-01/PN 56695-02 2T, 200 ml	

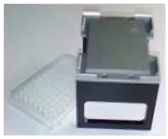
8.1.17 Tip Carrier

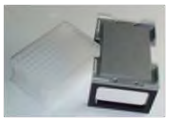
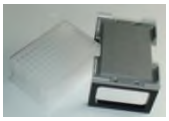
Tip Carrier		
Part Number	**Description**	**Image**
182060	**TIP_CAR_288** Carrier for 3x 96 tip racks portrait for 12-pipetting channel (1000 µl) ML STAR or 3x 24 tip racks portrait for 6-pipetting channel (5 ml) ML STAR (4T)	
182085	**TIP_CAR_480** Carrier for 5x 96 tip (10 µl, 50 µl, 300 µl, 1000 µl) racks or 5x 24 tip (5 ml) racks (6T)	
182390	**TIP_CAR_L384_A00** Carrier for 4x 96 tip racks for 16-channel (1000 µl) ML STAR or 3x 24 tip racks portrait for 8-channel (5 ml) ML STAR (6T)	
182074	**TIP_CAR_NTR_A00** Carrier for Nested Tip Racks (NTR) 6T; for 1 pack tray containing 1920x 10 µl, 50 µl, 300 µl CO-RE tips (5 stacks x 4 tip racks x 96 tips) or 7680x 50 µl CO-RE tips for 384 Probe Head (5 stacks x 4 tip racks x 384 tips)	
182040	**Adapter for TIP_CAR_480** Intermediate storage position for tips allows pick-up of a single tip/row/column using the CO-RE 96 Probe Head	
191055	**384 Tip Support** Intermediate storage position for tips allows pick-up of a single tip/row/column using the CO-RE 384 Probe Head The tip support fits into Multiflex tip module (PN 188160) or tip carrier (PN 182085)	
182041	**Drip Pan for Tip Carrier for 1000 µl and 5 ml Tips** Fits into TIP_CAR_480 (PN 182085), Multiflex Tip Module (PN 188160)	

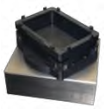
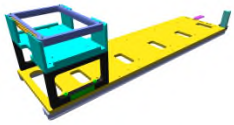
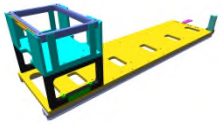
8.1.18 Sample Tube Carrier and Inserts

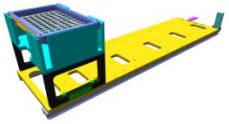
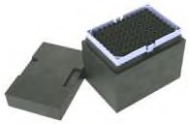
Sample Tube Carrier and Inserts		
Part Number	**Description**	**Image**
173400	**4 SMP-CAR-24** Set of 4x sample carrier for 24 tubes sizes 14.5 mm x60 mm – 18 mm x 120 mm (1T)	
173410	**3 SMP-CAR-32** Set of 3x sample carrier for 32 tubes sizes 11 mm x60 mm – 14 mm x 120 mm (1T)	
182045	**SMP-CAR-12** Sample Carrier for 12 Falcon tubes 50 ml (2T)	
185346	**Tube Carrier Filling Rack** Holds up to four 1T sample carriers for loading and filling of tubes in the carriers. Compatible with 1T sample carriers 24, 32.	
190070	**Rack Holder** Holds up to four 1T sample carriers for loading and filling tubes in carriers and transporting up to four sample carriers to and from a ML STAR workstation. Compatible with 1T sample carriers 24, 32.	
182238	**EPI-INS-32L** Set of 32 inserts for 1.5 ml Eppendorf cups in SMP-CAR-32	
182239	**EPI-INS-32S** Set of 32 inserts for 0.5 ml Eppendorf cups in SMP-CAR-32	
187142	**24-FALCON-INS-15ML** Set of 24 inserts for 15 ml Falcon tubes in SMP-CAR-24	
187350	**8 inserts for 1.5 ml Eppendorf cups in SMP-CAR-32** Set of 8 inserts for 1.5 ml Eppendorf cups in SMP-CAR-32	
185393	**SET OF INSERTS 24/16.5 mm 24** Set of 24 inserts for tubes with outer diameter < 16.5 mm in SMP-CAR-24	
There are more inserts available. Please consult a local HAMILTON representative.		

8.1.19 Multiflex Carrier

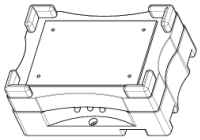
Multiflex Carrier		
Part Number	**Description**	**Image**
188039	**Multiflex Carrier Base (Landscape Orientation)** Labware carrier base for up to 5 Multiflex Modules	
188160	**Multiflex Tip Module** Module to position a high-, standard-, low volume or 5 ml tip rack (but not a 384 tip rack) The Multiflex carrier base is not included.	
191420	**Multiflex NTR 96 Module** Module to position a Nested Tip Rack (NTR) with standard (300 µl), low volume (10 µl) or 50 µl tips The Multiflex carrier base is not included.	
196371	**Multiflex NTR 384 Module** Module to position a Nested Tip Rack (NTR) with 384 50 µl tips The Multiflex carrier base is not included.	
191425	**Multiflex NTR4 Module** Module to position a stack of 4 Nested Tip Racks (NTR) with standard (300 µl), low volume (10 µl) or 50 µl tips The Multiflex carrier base is not included.	
188041	**Multiflex MTP Module** Module to position 96-/384-well plates in SBS format / or flat reagent troughs The Multiflex carrier base is not included.	
188042	**Multiflex DWP Module** Module to position a Deep Well Plate / tube racks (MATRIX or MICRONICS) / NUNC reagent trough The Multiflex carrier base is not included.	
188094APE	**MULTIFLEXMTPNESTCONTBASED** Module to hold a microplate by supporting the wells on a Multiflex Carrier. The Multiflex carrier base is not included. Alternatively, the module can be mounted on a PLT_CAR_L4_SHAKER base plate, PN 187001. This requires PN 188133APE MULTIFLEXMODULEBRACKET7T for fixation.	

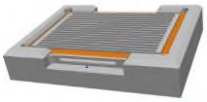
Multiflex Carrier		
Part Number	**Description**	**Image**
188228APE	**MULTIFLEXMTPNESTRACKBASED** Module to hold the frame of a microplate on a Multiflex Carrier. The Multiflex carrier base is not included. Alternatively, the module can be mounted on a PLT_CAR_L4_SHAKER base plate, PN 187001. This requires PN 188133APE MULTIFLEXMODULEBRACKET7T for fixation.	
188293	**MULTIFLEXXLDWPNESTRACKBASED** Module to position any SBS format plate, including Deep Well Plates up to H=90 mm / or flat reagent containers. Plates are held on their frame. The Multiflex carrier base is not included. Alternatively, the module can be mounted on a PLT_CAR_L4_SHAKER base plate, PN 187001. This requires PN 188133APE MULTIFLEXMODULEBRACKET7T for fixation.	
188180APE	**MULTIFLEXDWPNESTCONTBASED** Module to hold a Deep Well Plate by supporting the wells on a Multiflex Carrier. The Multiflex carrier base is not included. Alternatively, the module can be mounted on a PLT_CAR_L4_SHAKER base plate, PN 187001. This requires PN 188133APE MULTIFLEXMODULEBRACKET7T for fixation.	
188043	**CO-RE Gripper on Multiflex Module** Plate handling tool for plate transfer on the deck using two pipetting channels. Includes module with parking position for CO-RE gripper. The Multiflex carrier base is not included.	
188044	**Multiflex Stacker Module (Landscape)** Module to use as passive plate hotel. Depending on plate height, up to 10 plates can be stacked on one position. The Multiflex carrier base is not included.	
188045	**Multiflex Heating Module** Heating module (up to 60 ºC) including one adapter for one labware type. Temperature: Room temperature up to 60 ºC Temperature gradient: ± 1 ºC The Multiflex carrier base is not included.	
188046	**Multiflex Cooling Module** Cooling module (4 ºC – 15 ºC) including one adapter for one labware type. Temperature: 4 ºC – 15 ºC Temperature gradient: ± 1 ºC The Multiflex carrier base is not included.	
808440	**MULTIFLEX COOLING-HEATING MODULE** Module to cool or heat one labware type. Temperature: 4 ºC – 95 ºC Dry mode feature The Multiflex carrier base is not included.	

Multiflex Carrier		
Part Number	**Description**	**Image**
188047	**Multiflex Reagent Trough Module** Module to hold six 50 ml troughs The Multiflex carrier base is not included.	
188048	**Multiflex Tube / Cup Module** Module to hold EPPENDORF, SARSTEDT, NUNC tubes 0.5 ml / 1.5 ml / 2.0 ml with or without snap-lid in a passively cooled adapter The Multiflex carrier base is not included.	
188049	**MULTIFLEX PCR Plate Module 96** Module to position a 96-well PCR plate. The Multiflex carrier base is not included.	
188052	**Multiflex PCR Plate Module 384** Module to position a 384-well PCR plate The Multiflex carrier base is not included.	
188053	**Multiflex Carrier Base (Portrait Orientation)** Labware carrier base for up to 3 Multiflex modules	
188055APE	**Plate Turntable** Automated device to turn plates from landscape into portrait orientation or vice-versa The Multiflex carrier base is not included.	
188313	**Multiflex Matrix Fixation Frame for 1.4 ml Matrix Tubes 47.1 mm** Module to hold down sealed Matrix tubes in a 96well tube rack for piercing using needles or tips. Supports one pierceable SBS format 96well tube rack. Requires a Multiflex MTP Module (PN 188041, not included) and a Multiflex carrier base (not included). Piercing needles not included.	
188095	**Multiflex MTP Fixation Frame 16.6 mm** Module to hold down sealed plates for piercing using needles or tips. Supports one pierceable SBS format plate. The Multiflex carrier base and piercing needles are not included.	
188295	**Multiflex MTP Fixation Frame for agenTCOURT 384 26.2 mm** Module to hold down sealed plates for piercing using needles or tips. Supports one pierceable SBS format plate. The Multiflex carrier base and piercing needles are not included.	
188395	**Multiflex MTP Fixation Frame for agenTCOURT 96 38.6 mm** Module to hold down sealed plates for piercing using needles or tips. Supports one pierceable SBS format plate. The Multiflex carrier base and piercing needles are not included.	

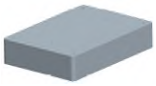
Multiflex Carrier		
Part Number	**Description**	**Image**
188495	**Multiflex MTP Fixation Frame for qiagen 31 mm** Module to hold down sealed plates for piercing using needles or tips. Supports one pierceable SBS format plate. The Multiflex carrier base and piercing needles are not included.	
188058APE	**Multiflex Lid Parking Module** Module to park the lid of the cooling or heating module. The Multiflex carrier base is not included.	
188059	**Multiflex Plate Stacker Module (Portrait)** Module to use as passive plate hotel. Depending on plate height, up to 10 plates can be stacked on one position. The Multiflex carrier base is not included.	
188114APE	**Multiflex Liquid Dispenser Trough 8** Module to automatically refill a trough on the deck with fresh reagent. Compatible with 8-channel instruments. The Multiflex carrier base is not included.	
188061APE	**Multiflex Tilt Module** Module to tilt plates on the y-axis (in landscape orientation) The Multiflex carrier base is not included.	
188062	**Multiflex Tip Stacker Module** Module to hold 4 standard volume or 6 low volume tip racks on one position The Multiflex carrier base is not included.	
188063APE	**Multiflex Seesaw Module** Module to shake bead-, cell-, or reagent solutions at a predefined speed in troughs or plates. Angle is adjustable up to 15°. 16 rpm – 69 rpm adjustable in 4 steps. The Multiflex carrier base is not included.	
188115APE	**Multiflex Liquid Dispenser Trough 96** Module to automatically refill a trough on the deck with fresh reagent. Compatible with 96 channel instruments. The Multiflex carrier base is not included.	
188078APE	**Multiflex Sterile Tip Box Module** Module to store sterile tips on the deck.	
182774	**STAR Shelf 4MTP/2AC Plates** Shelving Unit for 4 MTP or 2 archive plates	

8.1.20 CVS Vacuum System

CVS Vacuum System		
Part Number	**Description**	**Image**
199020 **190021**	**CVS Vacuum System including pump (230V)** **CVS Vacuum System including pump (115V)** Carrier with a vacuum box, one park position for manifold top and two plate positions including Vacuubrand Pump ME 4C VARIO with controller, pressure sensor and air-bleed valve (Waste bottles are not included. Please select PN 281540)	 
187143 **187149**	**TELESHAKER 220V FOR DWP** **TELESHAKER 115V FOR DWP** Maximum one shaker per CVS carrier.	
187788	**SHAKER HEATER CAT SH10 FOR DWP (110/220V)** Shaker heater (Shaking: 200 - 1200 1/min, orbital: 2 mm, temperature: from RT+5 ºC – 90 ºC) for standard 96 and Deep Well Plates Maximal one shaker heater per CVS carrier.	
281540	**BVS 4-Liter Waste Bottle** PP waste bottle for the BVS / CVS with confectioned cap, including connectors and LDPE Bucket 11 L Instruction sheet	
190034	**DWP Kit for BVS** Manifold Top for BVS / CVS and insert to adapt the BVS / CVS to the used collecting plate	 
186320	**BVS Insert Kit** Adjustable insert to adapt the BVS / CVS to the used collecting plate Contains two different spacers: H = 30 mm and H = 45 mm Adapter: H = 15 mm Range: 45 mm to 60 mm. Bottom of Elution Plate will be 27.4 mm to 42.4 mm below the Gasket of the Manifold top (contact surface filter plate). Round up/down to 28 mm to 42 mm (nominal values).	

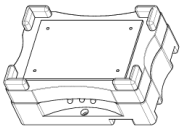
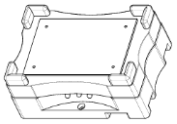
CVS Vacuum System		
Part Number	**Description**	**Image**
190035	**BVS Insert Kit for MTP** Insert to adapt the BVS / CVS to the used collecting plate Contains two different spacers: H = 15 mm and H = 30 mm Adapter: H = 17 mm Range: 32 mm to 47 mm. Bottom of Elution Plate will be 40.4 mm to 55.4 mm below the Gasket of the Manifold top (contact surface filter plate). Round up/down to 41 mm to 55 mm (nominal values).	
190036	**BVS Insert Kit for DWP** Insert to adapt the BVS / CVS to the used collecting plate Contains no spacers Adapter: H = 5 mm No Range: Bottom of Elution Plate will be 82.4 mm below the Gasket of the Manifold top (contact surface filter plate). Round down to 82 mm (nominal values).	
190037	**BVS Grid I for Millipore** Manifold Top for BVS / CVS incl. Grid I for Millipore Kits "Montage Plasmid Miniprep96 Kit" and "Montage PCR96 Cleanup Kit"	
186321	**BVS Grid II for Millipore** Manifold Top for BVS / CVS incl. Grid II for Millipore Kits with 96/384 SBS plates with one filter	
186303	**BVS Adapter for MN DNA KIT** Insert to adapt the BVS / CVS to the used collecting plate	
182712	**Frame for Filter Plate** To place filter plates on archive carriers	

8.1.21 HAMILTON Heater Shaker

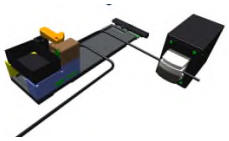
HAMILTON Heater Shaker		
Part Number	**Description**	**Image**
187001	**PLT_CAR_L4_Shaker** Template carrier with 4 positions for HAMILTON Heater Shaker (optional: Shaker H+P, Shaker Heater CAT) and plate bases (7T)	
190755	**Heater Shaker Box** Needed if more than two HAMILTON Heater Shakers are integrated (or the connectors "TCC1", "TCC2" on the ML STAR are occupied by other devices). One heater shaker module is connected to a USB port of the PC and serves as master module for up to seven additional HHSs. The modules are connected to the external heater shaker box (HSB), which serves as power supply and as signal distributor.	
199013	**Heater Shaker Support Block for MTP** HAMILTON Heater Shaker support block for MTP. Used to raise the heater shaker if only MTPs are processed.	
199027	**Heater Shaker 1.5 mm SARSTEDT 48 ml x 1.5 ml** HAMILTON Heater Shaker with 1.5 mm shaking orbit and fitted adapter for plates (shaking speed: 100 rpm – 1800 rpm, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm) Sarstedt tubes 1.5 ml: sterile Sarstedt Cat. No. 72.687.772 non-sterile Sarstedt Cat. No. 72.687 http://www.Sarstedt.com	
199033	**Heater Shaker 2.0 mm MTP Flat Bottom** HAMILTON Heater Shaker with 2.0 mm shaking orbit and flat bottom adapter (shaking speed: 100 rpm – 2500 rpm, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm)	
199034	**Heater Shaker 3.0 mm Flat Bottom** HAMILTON Heater Shaker with 3.0 mm shaking orbit and flat bottom adapter (shaking speed: 100 rpm – 2400 rpm, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm)	
199037	**Heater Shaker 1.5 mm NUNC DWP 96 2 ml** HAMILTON Heater Shaker with 1.5 mm shaking orbit and fitted adapter for plates (shaking speed: 100 rpm – 2000 rpm, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm)	
199038	**Heater Shaker 2.0 mm NUNC DWP 96 2 ml** HAMILTON Heater Shaker with 2.0 mm shaking orbit and fitted adapter for plates (shaking speed: 100 rpm – 2000 rpm, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm)	

HAMILTON Heater Shaker		
Part Number	**Description**	**Image**
199039	**Heater Shaker 3.0 mm NUNC DWP 96 2 ml** HAMILTON Heater Shaker with 3.0 mm shaking orbit and fitted adapter for plates (Shaking speed: 100 rpm – 1800 rpm, Temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm)	
188318	**Heater Shaker 2.0 mm MTP Flat Bottom APE** HAMILTON Heater Shaker with 2.0 mm shaking orbit and the option to be equipped with a customized adapter. The labware has to be sent to HAMILTON Bonaduz AG (Shaking speed depends on used adapter, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm). **NOTE** *Do not remove the Flat Bottom heating plate. Customized adapters are designed to mount onto the Flat Bottom heating plate.*	
188319	**Heater Shaker 3.0 mm MTP Flat Bottom APE** HAMILTON Heater Shaker with 3.0 mm shaking orbit and the option to be equipped with a customized adapter. The labware has to be sent to HAMILTON Bonaduz AG (shaking speed depends on used adapter, temperature control: RT+5 ºC – 105 ºC, max. loading: 300 mm). **NOTE** *Do not remove the Flat Bottom heating plate. Customized adapters are designed to mount onto the Flat Bottom heating plate.*	

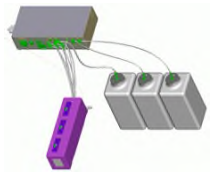
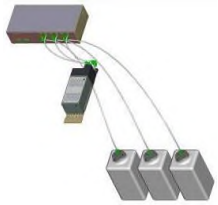
8.1.22 3rd Party Shakers

Shaker		
Part Number	**Description**	**Image**
187143 **187149**	**TELESHAKER 220V FOR DWP** **TELESHAKER 115V FOR DWP** Shaker (shaking: 100 - 2000 1/min, orbital: 2 mm) for standard 96 and Deep Well Plates on the PLT_CAR_L4_shaker (7T)	
187295 **187296**	**TELESHAKER 220V FOR MTP** **TELESHAKER 115V FOR MTP** Shaker (Shaking: 100 - 2000 1/min, Orbital: 2 mm) for standard 96-well plates (no DWP) on the PLT_CAR_L4_shaker (7T)	
187144	**DWP BASE FOR SHAKER CAR** Position for standard and Deep Well Plates on the PLT_CAR_L4_shaker (7T)	
187292	**MTP BASE FOR SHAKER CAR** Positions for standard well plates (no DWP) on the PLT_CAR_L4_shaker (7T)	
187788	**SHAKER HEATER CAT SH10 FOR DWP (110/220V)** Shaker heater (shaking: 200 - 1200 1/min, orbital: 2 mm, temperature: from RT+5 ºC – 90 ºC) for standard 96 and Deep Well Plates on the PLT_CAR_L4_shaker (7T)	
187789	**SHAKER HEATER CAT SH10 FOR MTP (110/220V)** Shaker heater (shaking: 200 - 1200 1/min, orbital: 2 mm, temperature: from RT+5 ºC – 90 ºC) for standard 96-well plates (no DWP) on the PLT_CAR_L4_shaker (7T)	

8.1.23 Multiflex Media Line

Multiflex Media Line		
Part Number	**Description**	**Image**
188156	**MULTIFLEX MEDIA LINE**	

8.1.24 Wash Stations

Wash Stations		
Part Number	**Description**	**Image**
186360	**CR Needle Wash Station** Wash Station for all needle types, two wash liquids, needle wash in parallel to the pipetting	
190248	**96/384 Wash Station Dual** Wash Station for 96/384 disposable tips, CO-RE 384 Probe Head, 2 wash chambers	
281107	**Wash / Waste Container 12 Liter** Wash and waste container for wash stations	

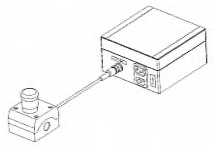
8.1.25 Housing Extension

Housing Extension		
Part Number	**Description**	**Image**
188122	**Housing Extension MPH** This housing extension can be mounted on either side of ML STAR instruments. If used on the left hand side on instruments with modular arm and MPH (96 or 384), the following waste has to be ordered separately; either: PN 188124APE Waste MPH L Rear or, PN 188125APE Waste MPH L Front If used on the left hand side on instruments with 'thin-' arm and MPH (96 or 384) the following waste has to be ordered separately; either: PN 188124APE Waste MPH L Rear or, PN 188125APE Waste MPH L Front **NOTE** *Side waste for MPH's (96 or 384) will not fit.* In conjunction with one of the following SCAP, a seal kit (PN 188235) is required when ordering this item. It has to be ordered separately. UV Light requires UV resistant acrylic covers which have to be ordered separately. Please consult a HAMILTON representative.	
188235	**Housing Extension MPH HEPA-Hood Kit** Seal kit required for the SCAP Filter Hood of Housing Extension MPH, PN 188122	

8.1.26 CO-RE Tools

CO-RE Tools		
Part Number	**Description**	**Image**
188066	**CO-RE Gripper with Attachment for Waste Block** Plate handling tool for plate transfer on the deck using 2 1000 µl pipetting channels. Includes parking position for attachment to waste block (waste block not included). **NOTE** *Order this gripper if the ML STAR instrument is equipped with 1000 µl pipetting channels only.*	
184089	**CO-RE Gripper with Attachment for Waste Block** **(For Instruments with 1000 µl and 5 ml Pipetting Channels)** Plate handling tool for plate transfer on the deck using 2 1000 µl pipetting channels. Includes parking position for attachment to waste block (waste block not included). **NOTE** *Order this gripper if the ML STAR instrument is equipped with 1000 µl pipetting channels and 5 ml pipetting channels.*	
184099	**CO-RE Gripper with Attachment for Waste Block** **(For Instruments with 1000 µl and 5 ml Pipetting Channels)** Plate handling tool for plate transfer on the deck using 2 5 ml pipetting channels. Includes parking position for attachment to waste block (waste block not included). **NOTE** *Order this gripper if the ML STAR instrument is equipped with 5 ml pipetting channels.*	
186100	**CO-RE Gripper** Gripper tool for plate transport with pipetting channels, including parking position. **NOTE** *This gripper can be installed by removing a plate position of the plate carrier PLT_CAR_L5MD, PN 182365*	
188227APE	**CO-RE Lid Tool (CLT)** Suction cup that can be picked up by a CO-RE pipetting channel to move lids.	
188082APE	**Replacement CO-RE Lid Tool (CLT)** Replacement suction cup.	

8.1.27 Emergency Stop Box

Emergency Stop Box		
Part Number	**Description**	**Image**
186060APE	**Emergency Stop Box** Category 0 Emergency Stop Button	

8.1.28 Mains Power Cables

Mains Power Cables		
Part Number	**Description**	**Image**
355234	**POWER CABLE EU, 2.50 m C13**	
355235	**POWER CABLE CH, 2.50 m C13**	
355236	**POWER CABLE US, 2.50 m C13**	
355237	**POWER CABLE UK, 2.50 m C13**	

8.1.29 HAMILTON Standard Computers

HAMILTON Standard Computers			
Part Number	**Description**		**Image**
	WINDOWS 10 COMPUTERS		
396341	**STANDARD PC WIN10 CH**	**Swiss German**	
396342	**STANDARD PC WIN10 FR**	**French**	
396343	**STANDARD PC WIN10 US**	**US English**	
396344	**STANDARD PC WIN10 ES**	**Spanish**	
396345	**STANDARD PC WIN10 IT**	**Italian**	
396346	**STANDARD PC WIN10 GE**	**German**	
396347	**STANDARD PC WIN10 GB**	**UK English**	
	(The mains power cable is not included)		
Refer to Section 7.11 HAMILTON Standard PC Specifications			

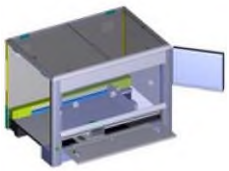
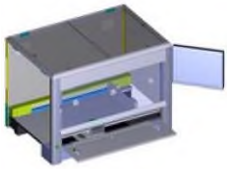
8.1.30 VENUS Software

VENUS Software	
Part Number	**Description**
911264	**VENUS four V4.5**
911095	**VENUS one Dynamic Scheduler** (This package is compatible to VENUS four)
911099	**Venus one TADM feature** (This package is compatible to VENUS four)
911122	**VENUS one Database PLUS** (This package is compatible to VENUS four)

8.1.31 ML STAR Manuals

VENUS Software	
Part Number	**Description**
627043	**ML STAR Line Operator´s Manual**
627044	**VENUS Software Programmer's Manual**

8.1.32 Touch Screen Monitor for the ML STAR

Touch Screen Monitor for the ML STAR		
Part Number	**Description**	**Image**
188250	**19" Touch Screen Kit Shaded** (Mains power cable not included)	
188260	**19" Touch Screen Kit Clear** (Mains power cable not included)	
355010	**Monitor Power Cable DE**	
355020	**Monitor Power Cable CH**	
355009	**Monitor Power Cable US**	
235062	**Monitor Power Cable UK**	

8.2 Appendix B: Chemical Compatibility

8.2.1 Abbreviations of Metals and Polymers Used in the Following Tables

1.4310	X10CrNi18-8 steel	PE	Polyethylene
1.4435	X2CrNiMo18-14-3 steel	PEEK	Polyetheretherketone
AA 5083 0	Aluminum	PMMA	Polymethyl-metharcylate
EPDM	Ethylene-propylene-elastomer	POM	Polyoxymethylene
FPM	Fluoroelastomer	PP	Polypropylene
FFKM	Kalrez	PTFE	Polytetrafluorethylene
FFPM	Per-Fluor-elastomer	PVC	Polyvinylchloride
FKM	Viton	PVDF	Polyvinylidenefluoride
NBR	Acrylnitril-butadiene-rubber	SI	Silicone

The tables for chemical compatibility are based on information from different manufacturers. The results refer to laboratory tests with raw materials. The results with these materials are often associated with effects that cannot be observed under laboratory conditions (e.g. temperature, pressure, tension, chemical influences of substances, design features, etc.). The results listed may be considered only as a guideline. In case of doubt, we recommend significant tests. The chemical resistance is not sufficient for an evaluation of a particular material for a product. Particular regulations (e.g. explosion prevention in the case of flammable liquids) have to be taken into account.

8.2.2 Chemical Resistance of the Multi-Probe Heads

	Materials													Overall resistance		
Chemical	1.4034	1.4301	1.4305	1.4404	1.4435	PE	PP	PTFE	PEEK	FKM	FFKM	EPT	ZrO_2	CO-RE	96	384
Acetic acid, 20%	2	1	1	1	1	1	1	1	1	2	1	1	0	1	1	2
Acetic acid, glacial	2	1	1	1	1	1	1	1	1	4	1	1	0	1	1	4
Acetone	1	1	1	1	1	2	1	1	1	4	1	1	0	1	1	4
Acetonitrile	1	1	1	1	1	1	3	1	0	2	0	3	0	3	3	2
Ammonium hydroxide, 5%	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1
Chloroform	1	1	1	1	1	3	3	1	1	1	1	4	0	4	4	1
Deionized water	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1
Dimethyl formamide	1	1	1	1	1	1	1	1	1	3	1	1	0	1	1	3
Dimethyl sulfoxide	1	1	1	1	1	1	1	1	0	0	0	1	0	1	1	1
Ethyl acetate	1	1	1	1	1	2	1	1	1	4	1	1	0	1	1	4
Hexane	1	1	1	1	1	3	2	1	1	1	1	4	0	4	4	1
Hydrochloric acid, 5%	4L	2L	3L	2L	2L	1	1	1	1	1	1	1	1	1	1	1
Hydrochloric acid, 20%	4L	3L	3L	2L	2L	1	1	1	1	1	1	1	1	1	1	1
Hydrogen peroxide, 10%	1	1	1	1	1	2	2	1	1	2	2	2	1	2	2	2
Isopropyl alcohol	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1
Methanol	1	1	1	1	1	1	1	1	1	2	1	1	0	1	1	2
Methylene chloride	1	1	1	1	1	4	3	1	2	2	1	4	0	4	4	2
Nitric acid, 5-10%	1	1	1	1	1	1	1	1	1	1	1	3	1	3	3	1
Nitric acid, 70%	1	1	1	1	1	3	4	1	1	2	1	3	1	3	3	2
Phosphate buffer	1	1	1	1	1	1	1	1	0	1	1	1	0	1	1	1
Phosphoric acid, 85%	3	2	3	2	2	1	1	1	0	1	1	1	1	3	3	2
Potassium hydroxide conc.	3	1	2	1	1	1	1	1	1	3	1	1	1	2	2	3
Sodium acetate	1	1	1	1	1	1	1	1	0	4	1	1	0	1	1	4
Sodium borate	1	1	1	1	1	1	1	1	0	1	1	1	0	1	1	1
Sulfuric acid, 1-75%	4	2	3	2	2	1	1	1	2	1	1	1	1	3	3	2
Urine	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1
Triethylamine	1	1	1	1	1	0	4	1	0	4	0	4	0	4	4	4
Toluene	1	1	1	1	1	3	3	1	1	1	1	4	0	4	4	1
Sodium hydroxide 5%	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Formic acid 5%	3	1	2	1	1	1	1	1	1	2	1	1	0	2	2	2
Sodium hypochloride 10%	3L	2L	2L	1L	1L	1	1	1	0	1	1	1	0	1	1	1
Ethanol	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1

Effects (key to codes in the table above):

1 = No effect, little or no noticeable change
2 = Slight corrosion or discoloration
3 = Moderate corrosion or other change in physical properties or dimensions; not recommended for continuous contact
4 = Severe corrosion or physical change; prolonged contact not recommended
0 = No data
L = Danger of pitting / corrosion (a localized form of corrosion that leads to the creation of small holes in the metal)

Assigned Materials

CO-RE Head consist of 1.4305, EPDM, PEEK, ZrO_2 and PTFE
96 Head consist of 1.4305, EPDM, PEEK, ZrO_2 and PTFE
384 Head consist of 1.4435, Viton and ZrO_2

8.2.3 Chemical Resistance of the CR Needle Wash Station

Chemical	1.4435	1.4310	AA 5083 0	FFKM	PE	PTFE	PEEK	PP	FFPM	Overall resistance
Acetic acid, 20%	1	1	2	1	1	1	1	1	1	1
Acetic acid, glacial	1	1	2	4	1	1	1	1	1	1
Acetone	1	1	1	1	2	1	1	1	1	2
Acetonitrile	1	1	0	0	1	1	0	3	0	(3)
Ammonium hydroxide, 5%	1	1	0	0	1	1	1	1	1	1
Chloroform	1	1	1	1	3	1	1	3	1	3
Deionized water[2]	1	1	1	1	1	1	1	1	1	1
Dimethyl formamide	1	1	0	1	1	1	1	1	1	1
Dimethyl sulfoxide max. 30%	1	1	0	1	1	1	0	1	0	1
Ethyl acetate	1	1	0	1	2	1	1	1	1	2
Hexane	1	1	1	1	3	1	1	2	1	3
Hydrochloric acid, 20%	2L	2L	2	1	1	1	1	1	1	4
Isopropyl alcohol	1	1	1	1	1	1	1	1	1	1
Methanol	1	1	1	1	1	1	1	1	1	1
Methylene chloride	1	1	1	1	4	1	2	3	1	4
Nitric acid, 5-10%	1	1	2	1	1	1	1	1	1	1
Nitric acid, 70%	1	1	2	1	3	1	1	4	1	4
Phosphate buffer	1	1	0	1	1	1	0	1	1	(1)
Phosphoric acid, 85%	2	2	0	1	1	1	0	1	1	(2)
Potassium hydroxide conc.	1	1	4	1	1	1	1	1	1	1
Sodium acetate	1	1	0	1	1	1	0	1	1	(1)
Sodium borate	1	1	0	1	1	1	0	1	1	(1)
Sulfuric acid, 1-75%	1L	2L	0	1	1	1	0	1	0	2
Urine	2	2	3	1	1	1	2	1	1	1
Triethylamine	1	1	0	1	0	1	1	1	1	4
Toluene	1	1	0	0	3	1	0	4	0	3

Effects (key to codes in above table):

1 = No effect, little or no noticeable change
2 = Slight corrosion or discoloration
3 = Moderate corrosion or other change in physical properties or dimensions; not recommended for continuous contact
4 = Severe corrosion or physical change; prolonged contact not recommended
0 = No data
L = Danger of pitting / corrosion (a localized form of corrosion that leads to the creation of small holes in the metal)

8.2.4 Chemical Resistance of the Wash Station

Chemical	1.4310	PE	PTFE	PEEK	PP	PVDF	FPM	SI	PVC	EPDM	NBR	POM
Acetic acid, 20%	1	1	1	1	1	1	3	2	1	2	3	1
Acetic acid, glacial	1	1	1	1	1	1	4	2	4	4	4	4
Acetone	1	2	1	1	1	3	4	3	0	1	4	1
Acetonitrile	1	1	1	0	3	1	3	0	0	3	4	3
Ammonium hydroxide, 5%	1	1	1	1	1	2	2	1	0	1	2	1
Chloroform	1	3	1	1	3	1	3	4	4	4	4	4
Deionized water²)	1	1	1	1	1	1	1	1	1	1	1	1
Dimethyl formamide	1	1	1	1	1	4	4	2	4	2	4	1
Dimethyl sulfoxide max. 30%	1	1	1	0	1	3	3	0	4	3	4	1
Ethyl acetate	1	2	1	1	1	3	4	2	4	3	4	1
Hexane	1	3	1	1	2	1	1	4	4	4	1	1
Hydrochloric acid, 20%	4	1	1	1	1	1	1	3	1	1	4	4
Isopropyl alcohol	1	1	1	1	1	1	1	1	4	1	3	1
Methanol	1	1	1	1	1	1	3	1	3	1	3	1
Methylene chloride	1	4	1	2	3	1	3	4	4	4	4	3
Nitric acid, 5-10%	1	1	1	1	1	1	1	2	1	2	4	4
Nitric acid, 70%	1	3	1	1	4	1	2	4	4	4	4	4
Phosphate buffer	1	1	1	0	1	1	1	4	0	1	1	1
Phosphoric acid, 85%	2	1	1	0	1	1	1	3	1	3	4	4
Potassium hydroxide conc.	1	1	1	1	1	2	4	3	0	1	3	3
Sodium acetate	1	1	1	0	1	1	3	4	3	1	3	1
Sodium borate	1	1	1	0	1	1	1	1	1	1	3	1
Sulfuric acid, 1-75%	2	1	1	2	1	1	1	3	1	4	4	4
Urine	1	1	1	1	1	1	1	1	1	1	1	1
Triethylamine	1	0	1	0	4	3	3	4	0	4	3	1
Toluene	1	3	1	1	3	1	1	4	4	4	4	1

Effects (key to codes in above table):

1 = No effect, little or no noticeable change
2 = Slight corrosion or discoloration
3 = Moderate corrosion or other change in physical properties or dimensions; not recommended for continuous contact
4 = Severe corrosion or physical change; prolonged contact not recommended
0 = No data

8.2.5 Chemical Resistance of the 96/384 Wash Station

Chemical	1.4310	PE	PP	PTFE	PEEK	FFPM	Hypalon	Overall resistance
Acetic acid, 20%	1	1	1	1	1	1	2	2
Acetic acid, glacial	1	1	1	1	1	1	3	3
Acetone	1	2	1	1	1	1	3	3
Acetonitrile	1	1	3	1	0	0	0	(3)
Ammonium hydroxide, 5%	1	1	1	1	1	1	3	3
Chloroform	1	3	3	1	1	1	4	4
Deionized water	1	1	1	1	1	1	1	1
Dimethyl formamide	1	1	1	1	1	1	3	3
Dimethyl sulfoxide, max. 30%	1	1	1	1	0	0	0	(1)
Ethyl acetate	1	2	1	1	1	1	4	4
Hexane	1	3	2	1	1	1	2	3
Hydrochloric acid, 20%	4	1	1	1	1	1	2	4
Isopropyl alcohol	1	1	1	1	1	1	1	1
Methanol	1	1	1	1	1	1	1	1
Methylene chloride	1	4	3	1	2	1	4	4
Nitric acid, 5-10%	1	1	1	1	1	1	2	2
Nitric acid, 70%	1	3	4	1	1	1	4	4
Phosphate buffer	1	1	1	1	0	1	1	(1)
Phosphoric acid, 85%	2	1	1	1	0	1	2	(2)
Potassium hydroxide conc.	1	1	1	1	1	1	2	2
Sodium acetate	1	1	1	1	0	1	2	(2)
Sodium borate	1	1	1	1	0	1	1	(1)
Sulfuric acid, 1-75%	2	1	1	1	2	1	2	2
Urine	1	1	1	1	1	1	1	1
Triethylamine	1	0	4	1	0	0	3	4
Toluene	1	3	3	1	1	1	1	3

Effects (key to codes in above table):

1 = No effect, little or no noticeable change
2 = Slight corrosion or discoloration
3 = Moderate corrosion or other change in physical properties or dimensions; not recommended for continuous contact
4 = Severe corrosion or physical change; prolonged contact not recommended
0 = No data

8.2.6 Chemical Resistance of the CVS

Chemical	PE	PP	PVDF	PMMA	AA6023 TTG	Remarks
N-Hexan	2/3	2/3	1/1	(3)	1/1	flammable
Ethyl-Acetate	1/3	1/3	3/3	0	1/1	flammable
Methanol	1/1	1/1	1/1	(3)	1/0	flammable
Acetonitrile (100%)	1/1	3/4	1/1	(3/4)	(1)	flammable
Methylene Chloride	0/0	(3)	(2)	(3)	1/1	flammable
Formic Acid (0.1%)	1/1	1/2	1/1	(3)	(3)	
Ammonia (0.1%)	1/1	1/1	(2)	2/2	1/1	

*PMMA may cause stains to the labware.

Resistance:

Two values are given for each medium:

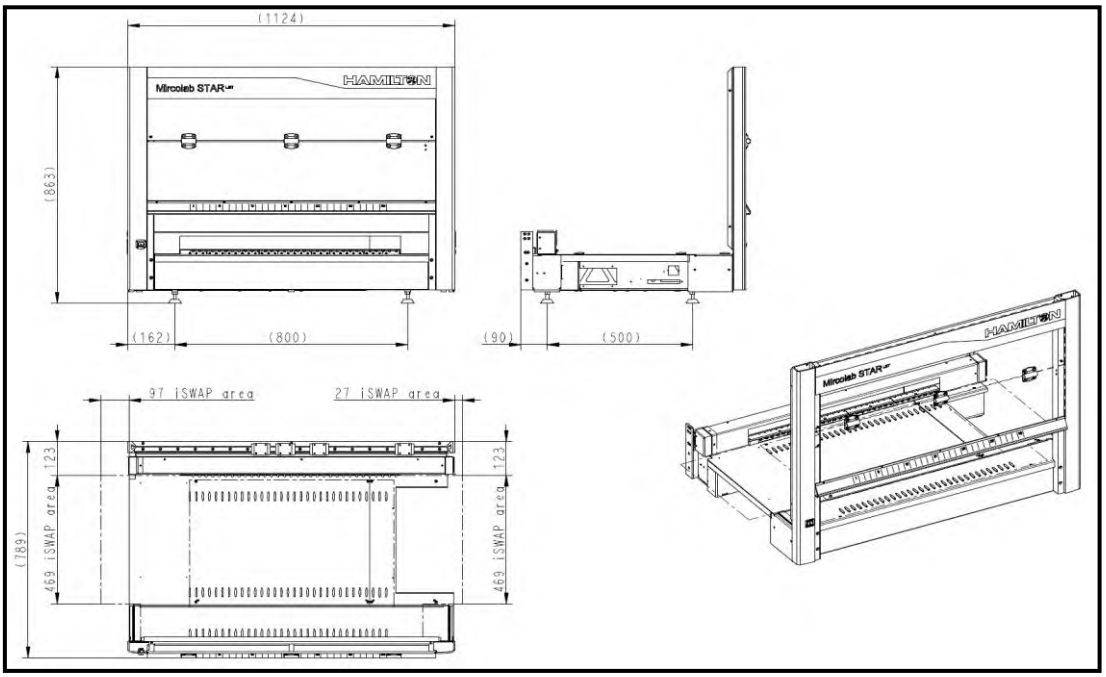
left number = value at +20 ºC / right number = value at +50 ºC.

Effects (key to codes in above table):

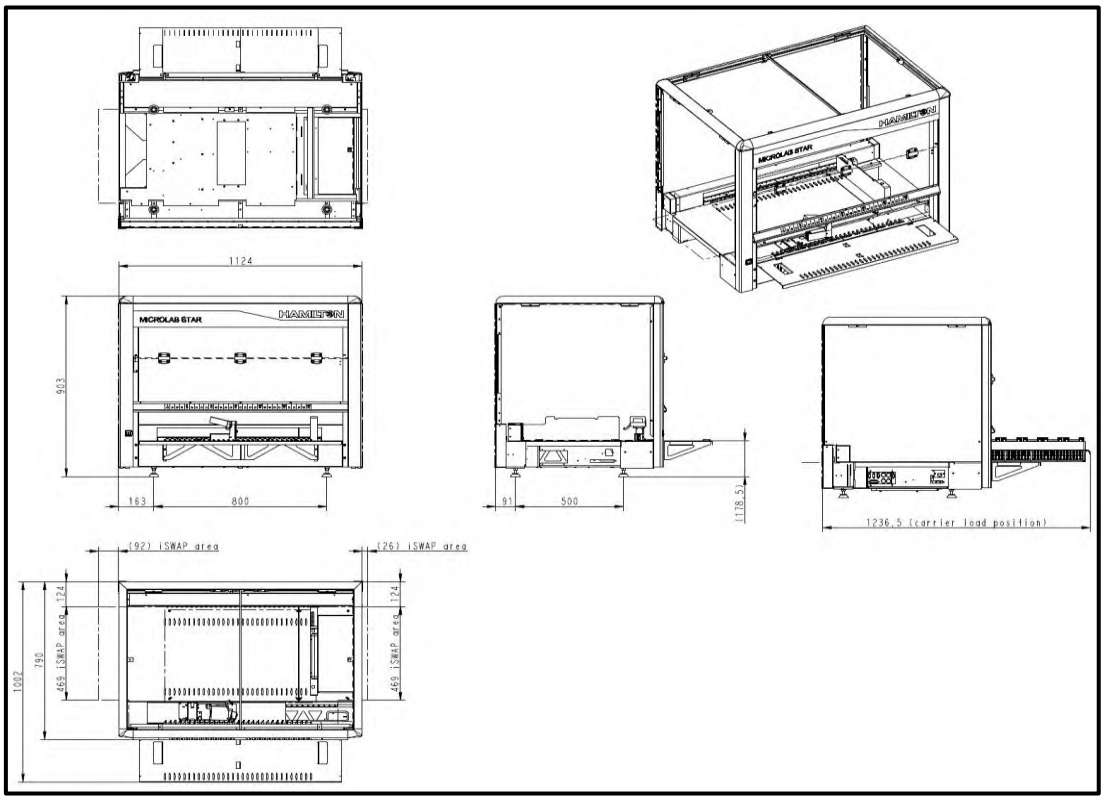
0 = No data available / no statement possible
1 = Very stable / suitable
2 = Good resistance / suitable
3 = Partially resistant
4 = Not resistant
K = No general specifications possible
L = Danger of pitting / corrosion
() = Estimated

8.3 Appendix C: Instrument Dimensions

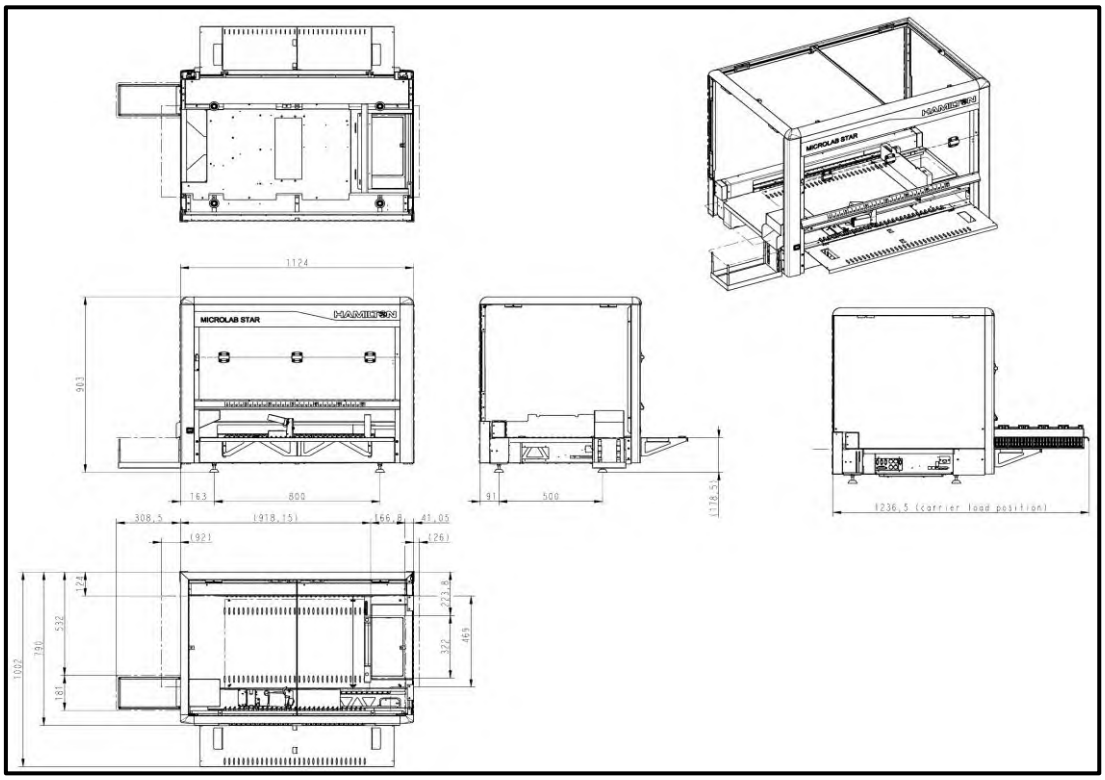
8.3.1 ML STARlet, Manual Load, Shown Without Housing



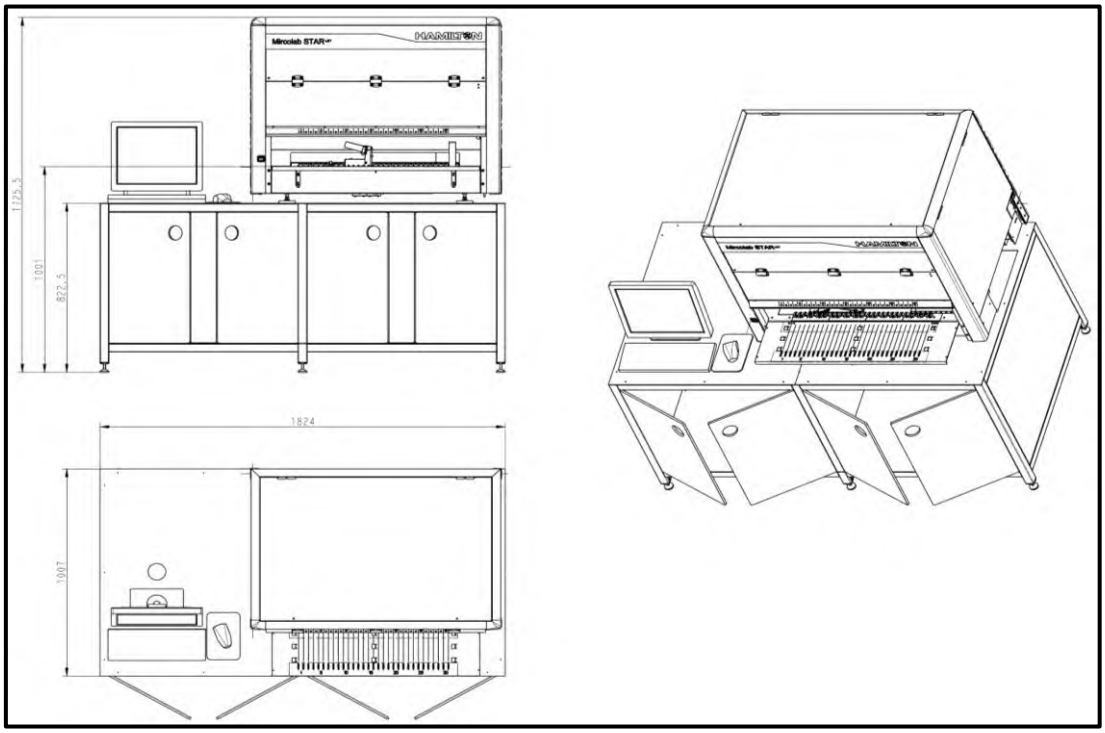
8.3.2 ML STARlet, Autoload



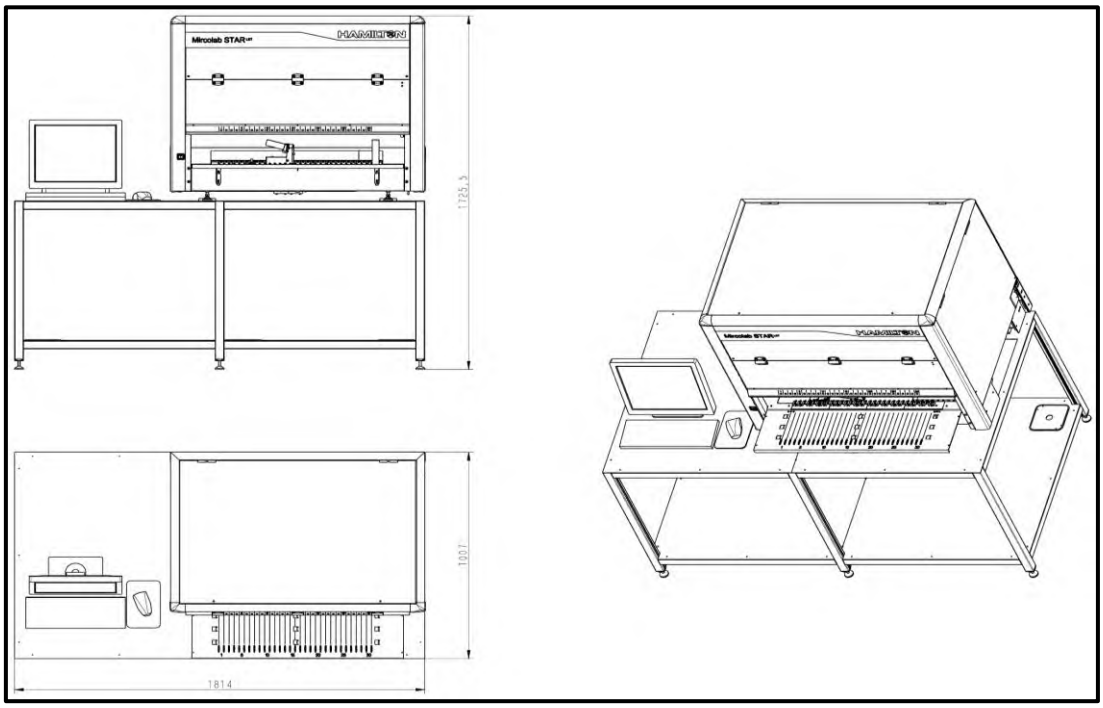
8.3.3 ML STARlet, Autoload, MPH



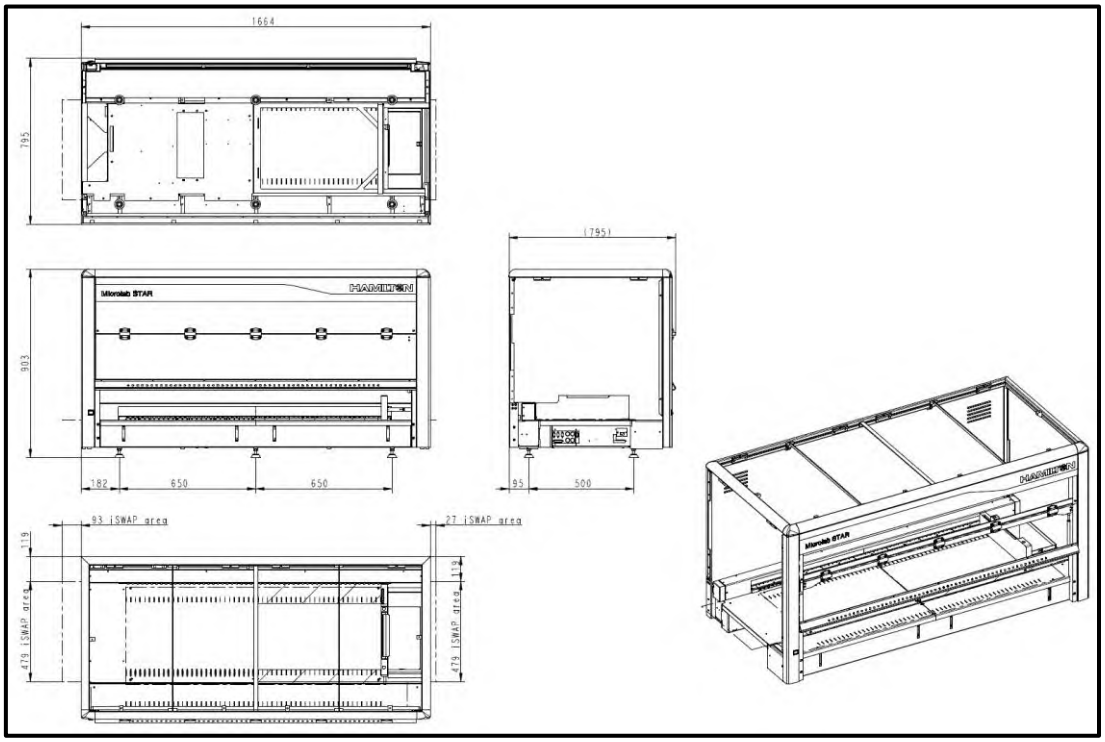
8.3.4 ML STARlet, on Bench with Doors and PC



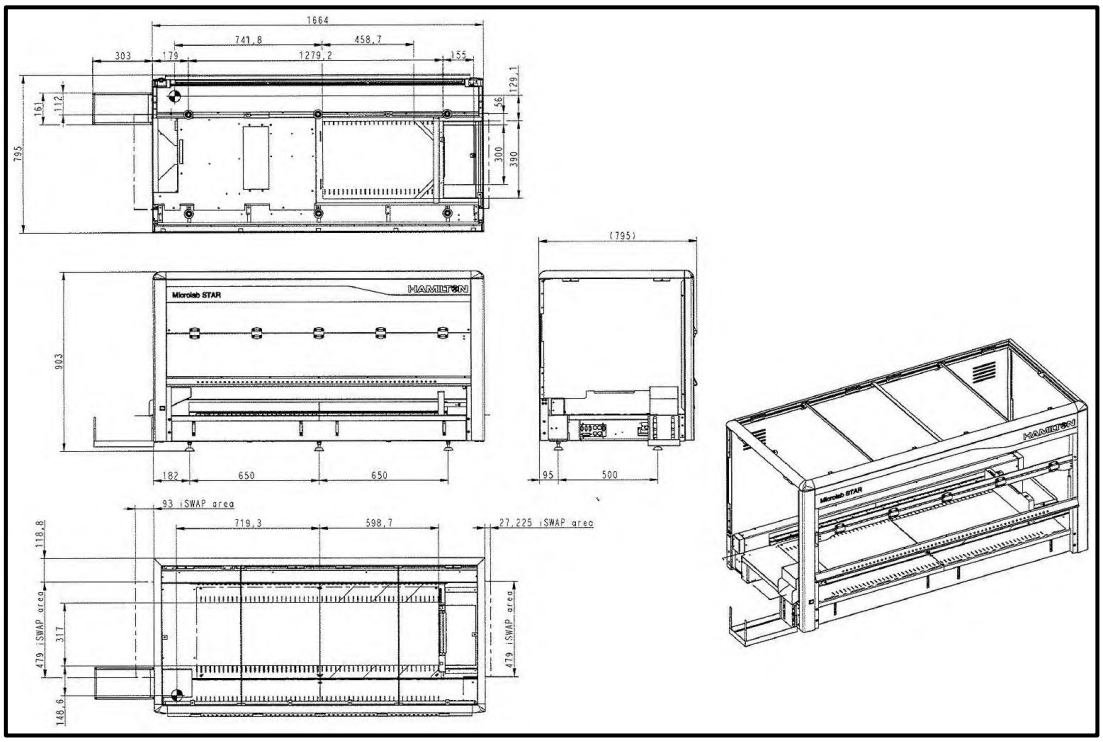
8.3.5 ML STARlet, on Table with PC



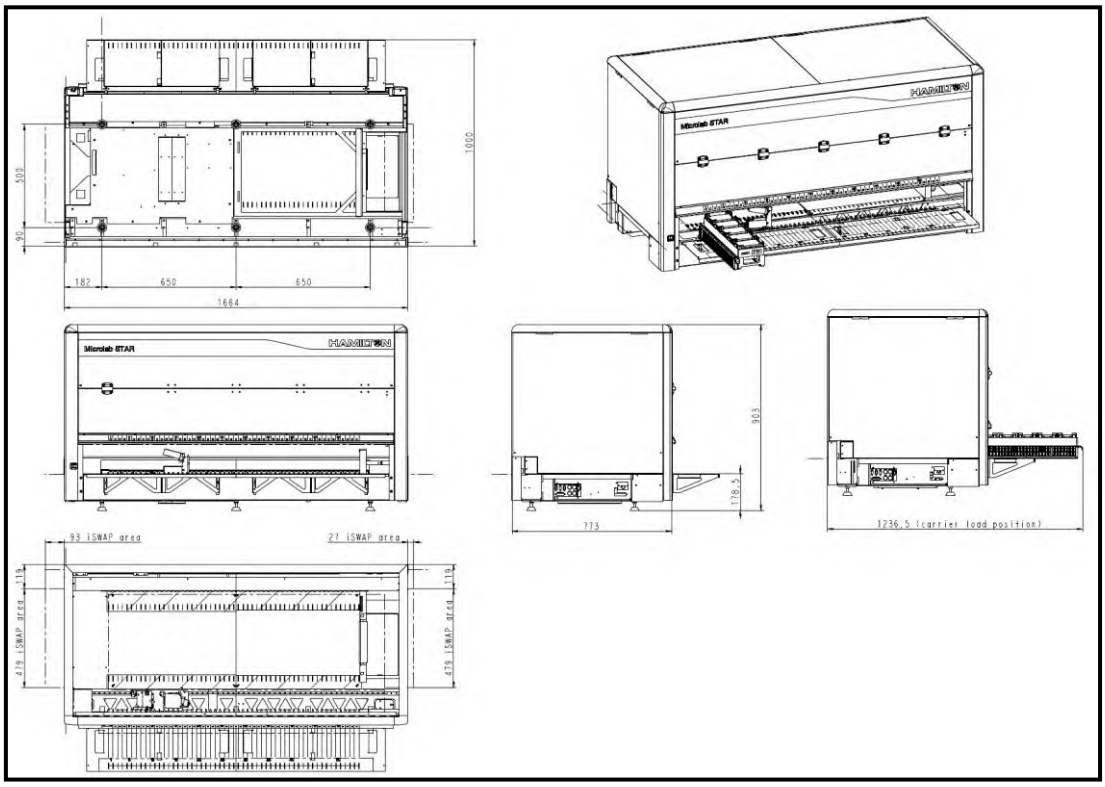
8.3.6 ML STAR, Manual Load



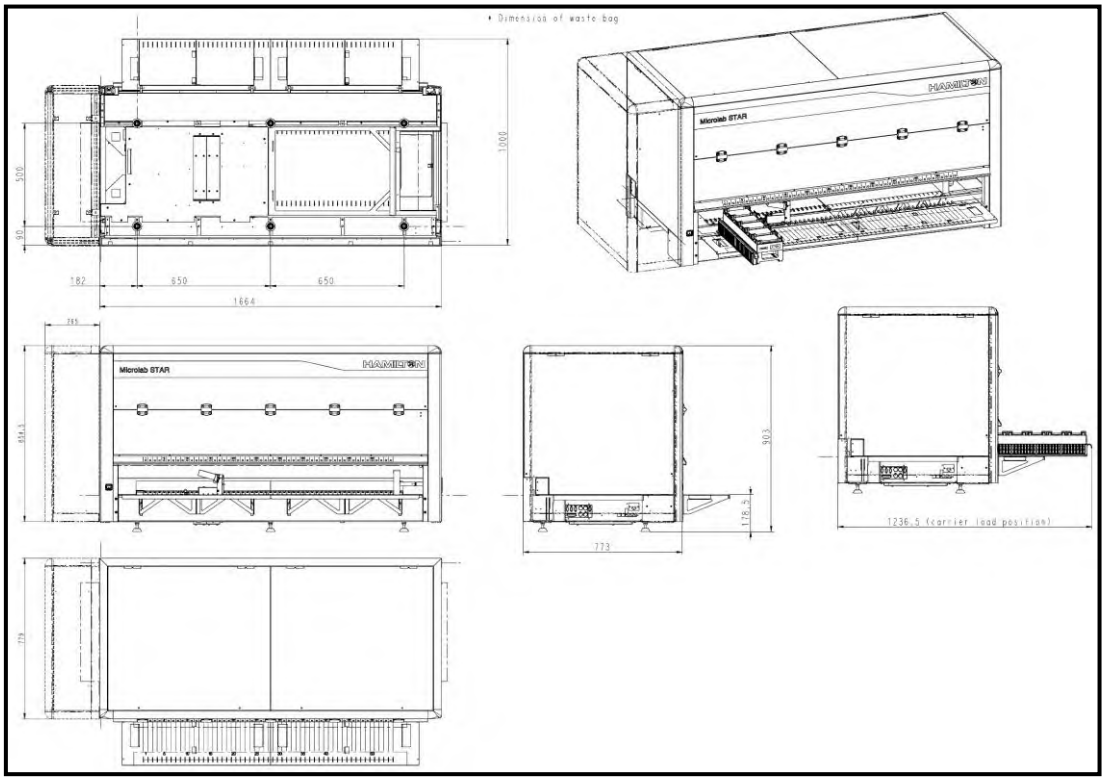
8.3.7 ML STAR, Manual Load, MPH



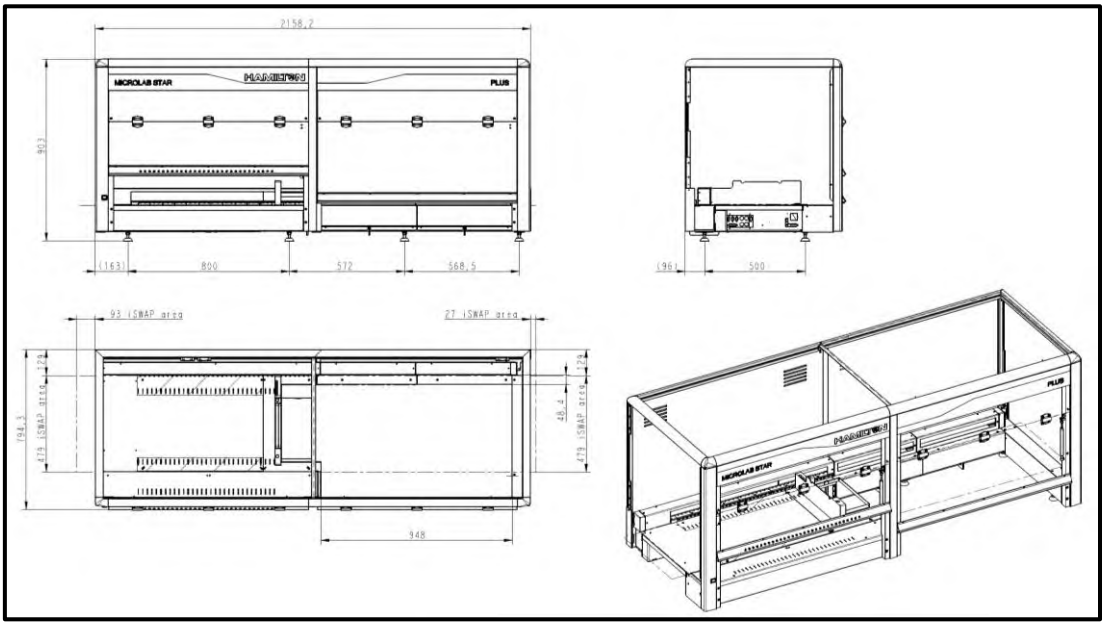
8.3.8 ML STAR, Autoload



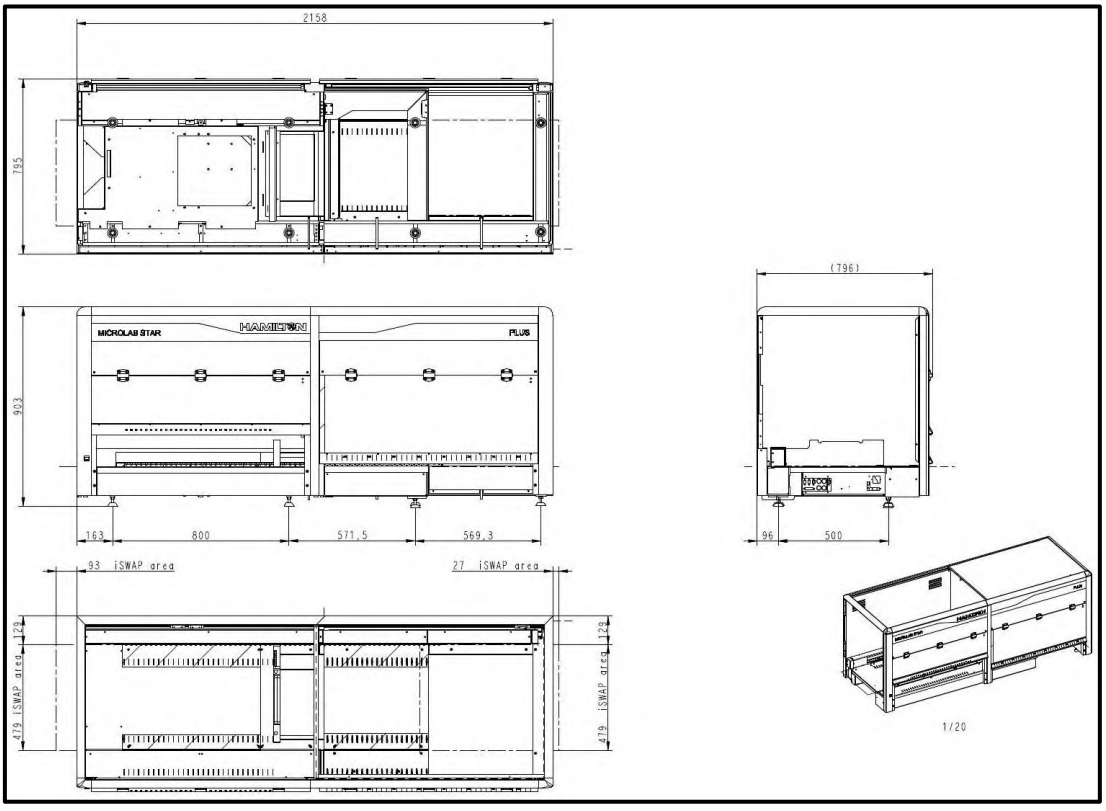
8.3.9 ML STAR, Autoload, Extension Housing Left



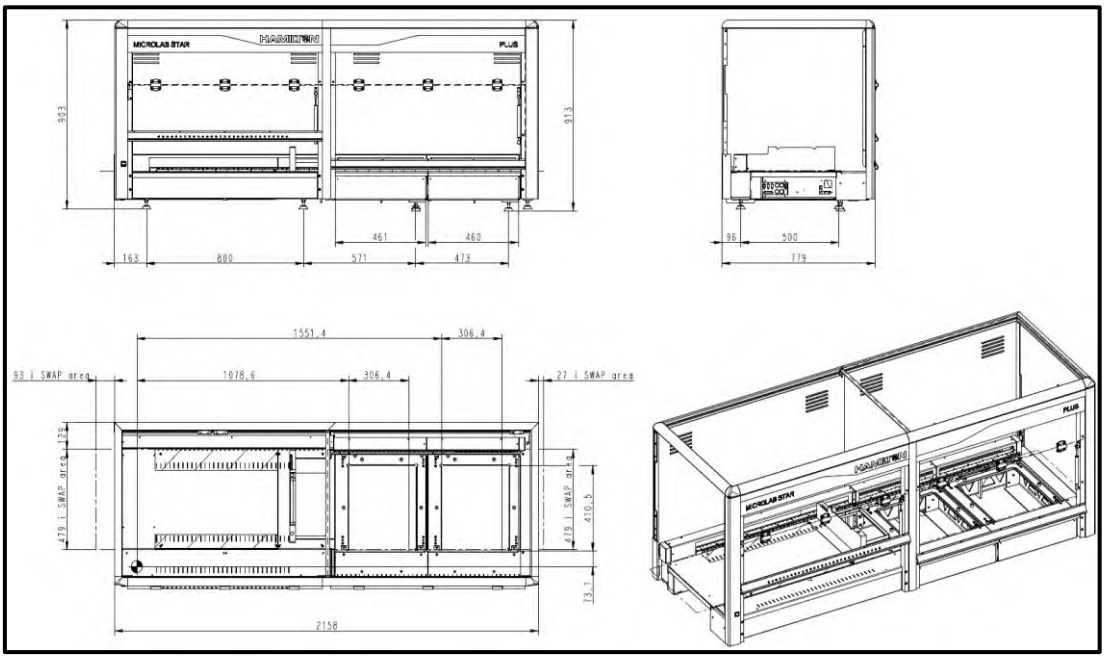
8.3.10 ML STARplus, Manual Load, 30T



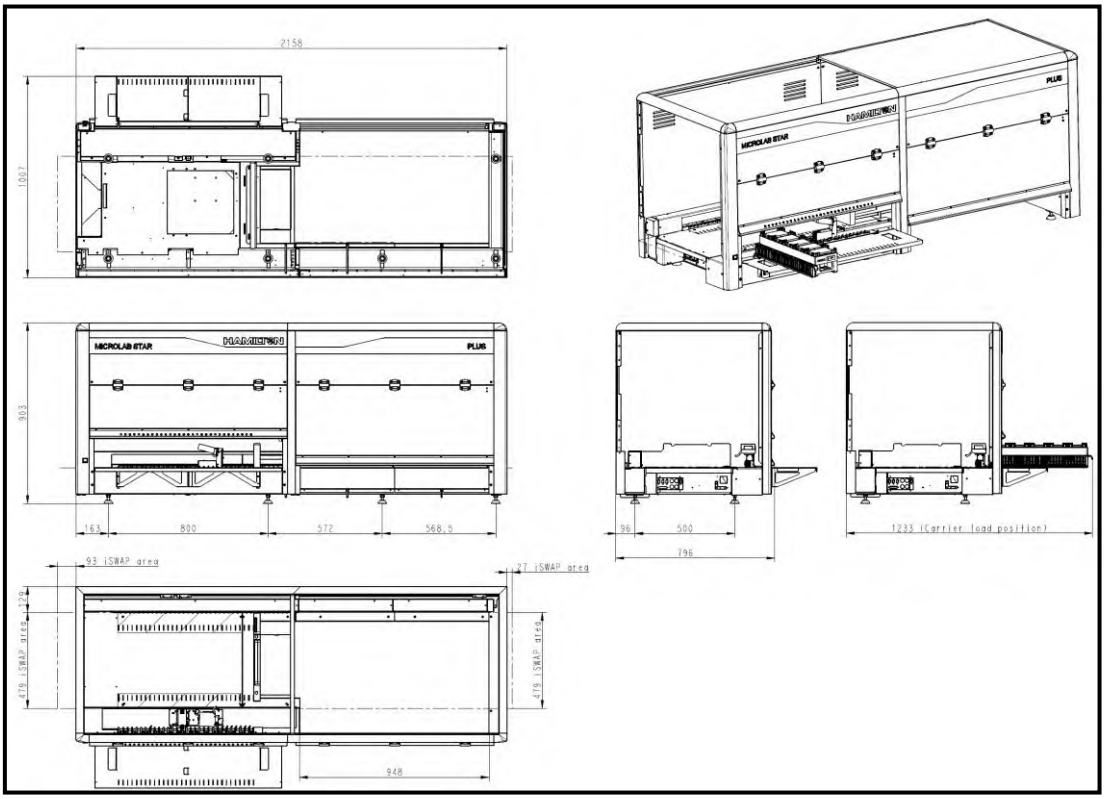
8.3.11 ML STARplus, Manual Load, 50T



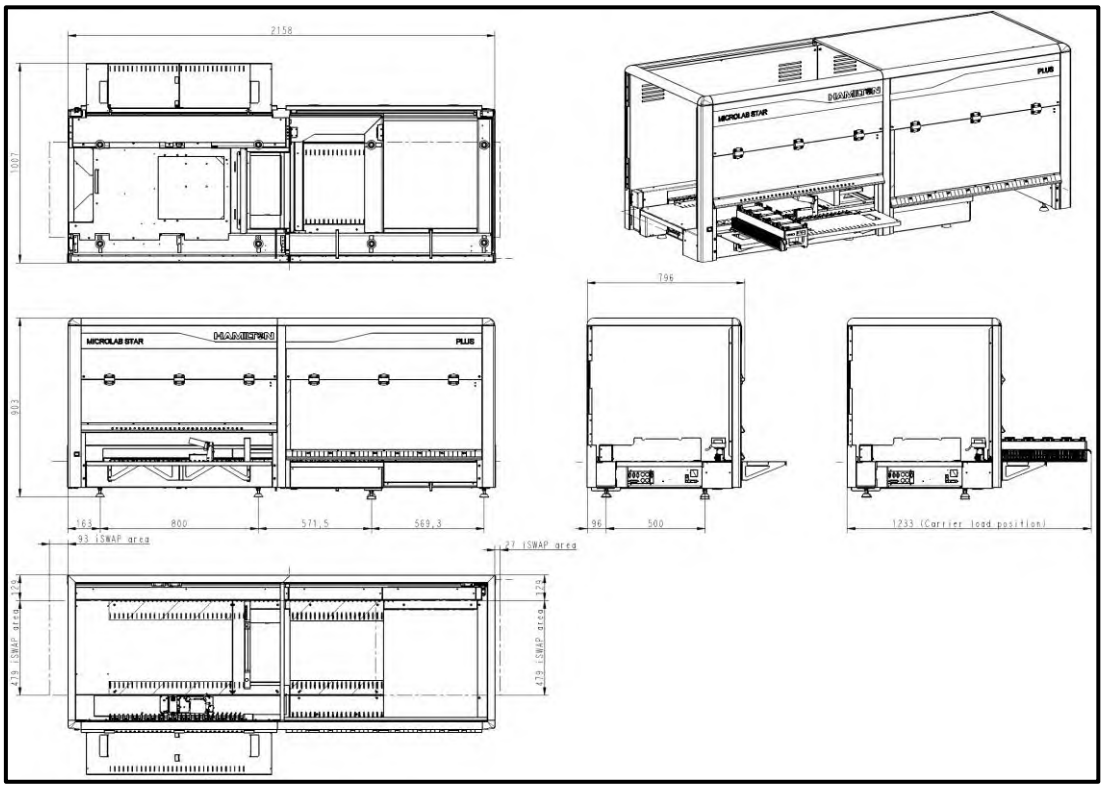
8.3.12 ML STARplus, Manual Load, 71T



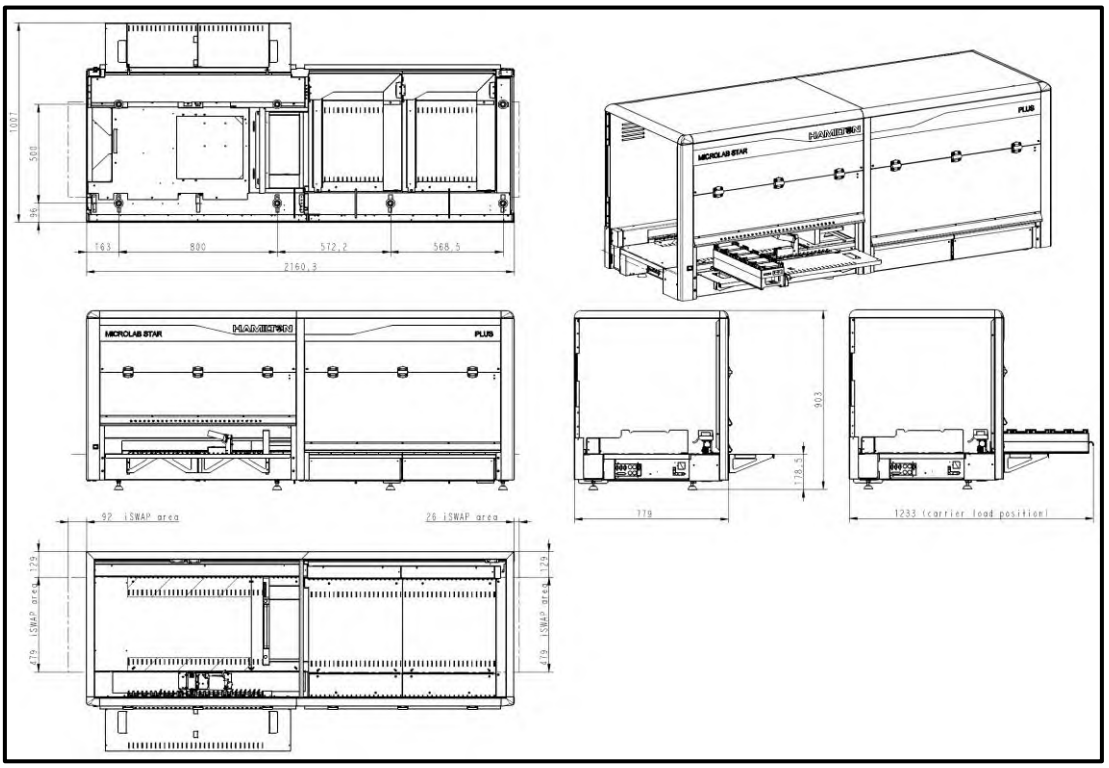
8.3.13 ML STARplus, Autoload, 30T



8.3.14 ML STARplus, Autoload, 50T



8.3.15 ML STARplus, Autoload, 71T



8.4 Appendix D: Regulatory Affairs

CE conformity is issued and maintained for the ML STAR. See the Declaration of Conformity provided with the instrument as well as the information in the following sections.

8.4.1 Radio Interference (USA and Canada)

This equipment has been tested and found to comply with the limits for a Class "A" digital device, pursuant to both Part 15 of the FCC Rules and the radio interference regulations of the Canadian Department of Communications. These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. This equipment generates, uses and can radiate radio frequency energy and, if not installed and used in accordance with the present user manual, may cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause harmful interference, in which case the user will be required to correct the interference at his own expense.

Pursuant to the Canadian Radio Interference Regulations, ICES-001 Notice for Industrial, Scientific and Medical Radio Frequency Generators, this ISM apparatus meets all requirements of the Canadian Interference-Causing Equipment Regulations. Please note that this requirement is only for generators which operate at over 10,000 Hz.

8.4.2 *In vitro* Diagnostics

The ML STAR is classified as a general laboratory instrument and not as an *in vitro* Diagnostic device.

NOTE

Products for general laboratory use are not in vitro Diagnostic medical devices unless such products in combination with an assay, in view of their characteristics and methods, are specifically intended by the assay manufacturer to be used for in vitro Diagnostic examinations.

There are ML STAR based instruments for *in vitro* Diagnostic use.

8.4.3 Applied Company Quality Management Systems

Applied company quality management systems EN ISO 9001 and EN ISO 13485.

Legal Manufacturer	HAMILTON Bonaduz AG Via Crusch 8 CH-7402 Bonaduz Switzerland (CHE)	HAMILTON Company Inc. 4970 Energy Way Reno, NV 89502 United States of America (USA)
Certification Body ISO 13485	TÜV Rheinland LGA Products GmbH Am Grauen Stein 29 D-51105 Köln-Poll Germany	TÜV Rheinland LGA Products GmbH Am Grauen Stein 29 D-51105 Köln-Poll Germany
Certification Body ISO 9001	TÜV Rheinland LGA Products GmbH Am Grauen Stein 29 D-51105 Köln-Poll Germany	TUV Rheinland of North America, Inc. 295 Foster Street, Suite 100, Littleton, MA 01460 USA

8.4.4 Declaration of Conformity

Each individual instrument includes a Declaration of Conformity showing the directives, regulations and additional information with which it conforms.

8.4.5 Declaration of Quality

Each individual instrument includes a Declaration of Quality. It will be filled out according to the results of the final inspection and is valid for the specific instrument serial number entered.

Each individual instrument includes a printed Declaration of Quality including the results of the final inspections and the instrument identification. It proves the pipetting performance achieved during final inspection.

8.4.6 WEEE Declaration

8.4.6.1 Recycling of a HAMILTON ML STAR Line Instrument

This section is mandatory for all instruments to be recycled within the European Union, United Kingdom, Norway, Island, Switzerland, Principality of Liechtenstein, Turkey.

In other countries, please follow the local regulations and codes.

The Directive 2012/19/EU EU legislation requires in the from manufacturers to organize the disposal and waste of electrical and electronic equipment (WEEE).

For this reason, HAMILTON Bonaduz AG took part in an initiative to organize the disposal of IVD instruments through a European disposal network called RENE. RENE is the largest recycling network for the disposal of electronic equipment in Europe. The mission of RENE is a European-wide, WEEE-compliant, high quality recycling for electrical and electronic equipment through a dense network of competent companies. As a result, HAMILTON Bonaduz AG gets a turn-key-solution that includes all processes from treatment of incoming orders over collection, logistics and recycling down to reporting and management of material flows.

HAMILTON offers a WEEE process in collaboration with Toolpoint and RENE AG.,

Responsibilities	
Ordering party	Decontamination Preparation for transport Note: The cost for decontamination and preparation for shipment is paid by the ordering party. On request HAMILTON offers to take care for that part of the recycling process.
RENE	Transport Disposal
Toolpoint	Registration Invoice the disposal to HAMILTON
HAMILTON	Organize the disposal in accordance of the WEEE directive

Recycling Process Workflow

a. Request for the collection of the HAMILTON instrument via Toolpoint (www.toolpoint.ch).

b. The responsibility for decontamination remains with the ordering party.

- It is mandatory to sign the decontamination form and send an electronic copy to Toolpoint.
- Toolpoint forwards the documentation to RENE who is in charge with the disposal of the instrument.

c. Completion of the decontamination confirmation form.

d. Preparation for transport: packing.

e. Activation of the recycling order.

f. Archiving of the decontamination confirmation.

g. Disposal of the instrument by RENE.

8.4.7 RoHS Compliance

Since June 2011 ML STAR Line instruments are in compliance with RoHS Directive 2002/95/EC of the European Parliament and of the Council of 27 January 2003, on the restriction of the use of certain hazardous substances in electrical and electronic equipment (RoHS) and its amendments.

Since July 2016 ML STAR Line instruments are in compliance with RoHS II Directive 2011/65/EU of the European Parliament and of the Council of 8 June 2011, on the restriction of the use of certain hazardous substances in electrical and electronic equipment (RoHS II) and its amendments.

Since July 2019 ML STAR Line instruments are in compliance with amendment ROHS directive 2015/863/EU. This amendment directive bans 4 phthalates in electric / electronic devices.

8.4.8 RoHS Reduction of Hazardous Substances

As indicated in Section 8.4.4 Declaration of Conformity, the European RoHS compliance is stated on the Declaration of Conformity.

8.4.9 21 CFR Part 11 Compliance

The FDA issued regulations covering agency criteria for maintenance, security and submittal of electronic records by publication of Final Rule 21 CFR Part 11 Electronic Records; Electronic Signatures in the Federal Register of March 20, 1997. Laboratory implementation and compliance with the 21 CFR Part 11 regulation requires a program combining GLP (Good Laboratory Practice) with compliant instrument software and secure LIMS database management. HAMILTON's Vector/VENUS software contains the tools necessary for 21 CFR Part 11 compliant operation of the Microlab robotic instruments (shown on the back of this document). This section describes the specific features of the Microlab Vector/VENUS software enabling compliant instrument operation along with areas of laboratory responsibility.

Requirements	Vector/VENUS Software	21 CFR Part 11.10 Section
Controlled system access	Microlab Vector/VENUS uses the security tools provided in Windows 7 or Windows 10 for five defined user groups	a, d
	Vector/VENUS functional protections	d, g
Files accessible and printable in a human readable form	Files can either be printed as text or from the correct viewer	b
Electronic records must be protected and maintained throughout the records retention period	User id and date/time stamps are applied to every electronic record when it is saved	g
	Changes to records are monitored by checksum	g
	HAMILTON maintains backward compatibility when reasonable	b, c
	Vector/VENUS is compatible with database software programs for version control for complete audit trails	e
Documentation	HAMILTON documentation for robotic instruments is controlled through an ISO-9001 compliant change control process consistent with 21 CFR Part 11 regulation	k (1)
Training	HAMILTON provides ISO-9001-compliant training with certification of training for users	i

NOTE

All other sections of 21 CFR part 11 are out of responsibility of Microlab Vector/VENUS Software and shall be covered by the customer.

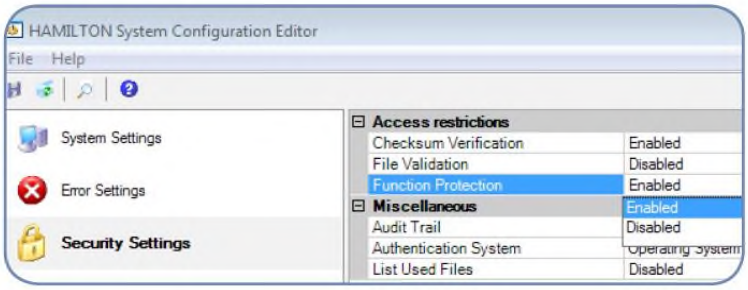
A. What do I need to do to be 21 CFR Part 11 compliant with my HAMILTON ML STAR instrument and Vector/VENUS software?

1. Define Windows user groups and assign users to the groups with unique user id/passwords. Define accessible directories for the user groups along with privileges in those directories, such as read-only or full access.

Windows environment for defining user groups and adding users to the groups.

The Windows operating systems have security tools to restrict user access to desktops and directories by user id/password protection and user group definitions. Each user group is given different access and privilege to HAMILTON files as is appropriate for their functions. Methods can be protected by Windows tools by saving all validated methods in limited-access directories, while saving non-validated methods in other restricted directories. Low level users can be allowed to access the directory with validated methods while blocking these user groups from the non-validated methods directory. Barcode and worklist files are not protected by Vector/VENUS software. These files may be created and used by programs outside of HAMILTON's control. Further protection of these files for full compliance must be accomplished by laboratory practices.

2. Enable the function protections in Vector/VENUS software. When these protections are enabled, Vector/VENUS checks the user group of the logged in user and allows functional privileges based on the user group.



Environment for enabling Vector/VENUS function protection.

The function protection in Vector/VENUS, along with the user groups established in Windows, provide authority-based access to the software for user level authentication and access. Additionally, all files are protected by checksum and are date/time stamped with the user id when the files are saved or created.

3. Define the SOPs and practices for complete laboratory compliance. Laboratories are responsible for defining SOPs for reaching full compliance when using Vector/VENUS software with HAMILTON instrumentation. These SOPs include:

   - Maintaining archives of old methods and software revisions throughout the records maintenance period
   - Implementing practices or additional software sufficient for a complete audit trail of records
   - Maintaining internal documentation on any HAMILTON instrumentation

   Recommendations for necessary SOPs are addressed inside.

B. HAMILTON Electronic Records

Operation of a HAMILTON instrument involves multiple file types or electronic records, which need to be protected under 21 CFR Part 11. These files are either input to be used during the method development and runtime or output files, which are created by the software during runtime.

Input Files:

- Labware Definitions
- Liquid Class Definitions
- Method Files
- System configuration files
- Output Files:
- Trace files

Barcode and worklist files are not protected by Vector/VENUS software. These files may be created and used by programs outside of HAMILTON's control. Further protection of these files for full compliance must be accomplished by laboratory practices.

C. Controls for Closed Systems

HAMILTON robotic instruments are controlled by closed computer systems. A closed system is one to which access is controlled by persons responsible for the content of the electronic records on that system. Below each section of 21 CFR Part 11.10 is followed by an explanation of HAMILTON's Vector/VENUS software compliance approach and the implementation responsibilities of the end user.

Controls for Closed Systems - 21 CFR Part 11.10

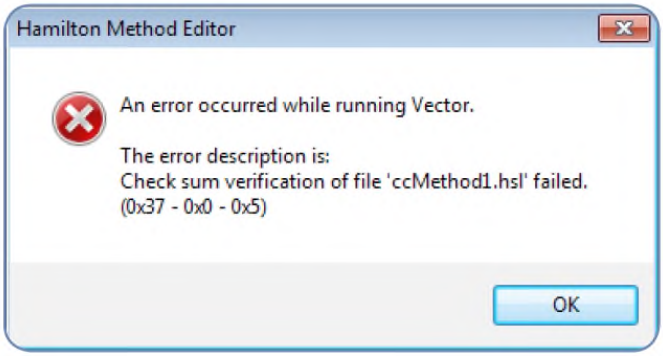
Persons who use closed systems to create, modify, maintain, or transmit electronic records shall employ procedures and controls designed to ensure the authenticity, integrity, and when appropriate, the confidentiality of electronic records, and to ensure that the signer cannot readily repudiate the signed record as not genuine. Such procedures and controls shall include the following:

This section describes the requirements to define an electronic record as protected for closed systems. This applies to all records created on a closed system, whether or not it is validated or formally recognized by the organization. The following sections address more specific requirements.

Validation - 21 CFR Part 11.10 (a)

Validation of systems to ensure accuracy, reliability, consistent intended performance, and the ability to discern invalid or altered records.

This refers to the validation of the electronic signature system to be used. The electronic signature system must be able to restrict access to various levels of functionality, identify when a change is made to an electronic record, by whom, and whether these changes were validated. Microlab Vector/VENUS software uses the security tools of the Windows 10 operating system. This system allows for different user groups within an organization to have different levels of access to the software. The files are user id and date/time stamped whenever they are saved. In addition, these files are checksum protected which protects files from inappropriate modification.

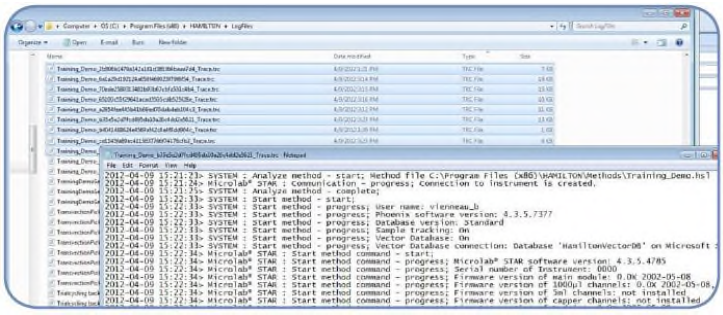


Error message seen when a checksum-protected file has been inappropriately modified. This message will appear when the user attempts to reopen the file

Readability - 21 CFR Part 11.10 (b)

The ability to generate accurate and complete copies of records in both human readable and electronic form suitable for inspection, review, and copying by the agency.

All records must be readily generated in either electronic or human readable form, along with the procedures used to create the records. This means all trace files must be retrievable along with the method files used to create these trace files (Figure 5). Organizations need to maintain electronic records as defined by agency-defined retention periods. Obsolete hardware and software versions do not need to be maintained, as long as all the records are retrievable with the change in technologies.



A trace file (.trc) documenting run time events for a particular method. A unique trace file is generated for each run time event with date/time and user id information located at the end of the file.

Vector/VENUS software files are either binary or ASCII text. ASCII is easily printed in human readable form with a text program. Other file types can be viewed and printed with the correct viewer or converted to ASCII and printed. HAMILTON maintains backward compatibility of method files between software revisions when reasonable. In the event a software revision no longer supports an archived method, the appropriate software version must be maintained to support the archived record.

Archived Record Protection - 21 CFR Part 11.10 (c)

Protection of records to enable their accurate and ready retrieval throughout the records retention period.

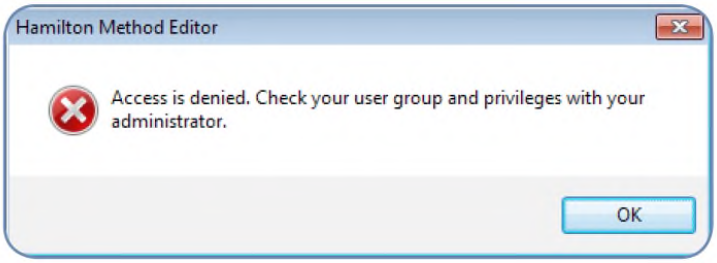
This is similar to the above section requiring maintained electronic files throughout the data retention period. However, this rule is not limited to retrieval of data. The method must be able to be processed as originally done. HAMILTON maintains backward compatibility of method files between software revisions when reasonable. Even if the current software version on the instrument does not support the archived method, records associated with the archived methods can still be viewed and are protected.

System Security - 21 CFR Part 11.10 (d)

Limiting system access to authorized individuals.

System access can be limited by current bilateral identification systems, such as user id and passwords, and/or by a single biometric identification. User access must be defined and limited to various levels of the software. This security can be applied through Windows 10 security tools.

System access for Vector/VENUS software is controlled through the Windows 10 operating system. Software allows access to various levels of the software based on the user group membership of the logged in user. If a user with an insufficient access level attempts to enter restricted sections of the software, an error message is displayed, and the user is blocked from inappropriate access.



Error message shown when a user with insufficient access attempts to open a restricted file or directory.

Audit Trail - 21 CFR Part 11.10 (e)

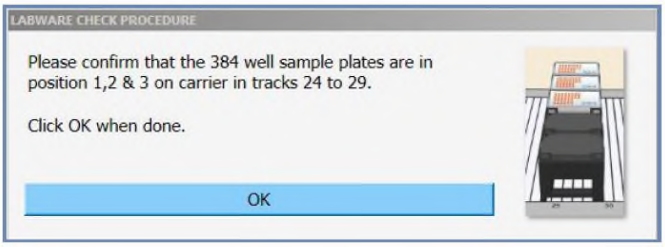
Use of secure, computer-generated time-stamped audit trails to independently record the date and time of operator entries and actions that create, modify or delete electronic records. Record changes shall not obscure previously recorded information. Such audit trail documentation shall be retained for a period at least as long as that required for the subject electronic records shall be available for agency review and copying.

Complete audit trail documentation is outside the scope of Vector/VENUS software and must be maintained by laboratory practices.

Sequencing - 21 CFR Part 11.10 (f)

Use of operational system checks to enforce permitted sequencing of steps and events, as appropriate.

The purpose of operational system checks is to verify that operations are not performed out of sequence as defined by the method. It is the agency's intent that such checks be performed by the computer system. Method checkpoints can be written into the program to direct the user and ensure that certain events have taken place before continuing with the method, such as all labware being in position. However, fulfilling this is a laboratory procedure and is not enforced within the Vector/VENUS software.

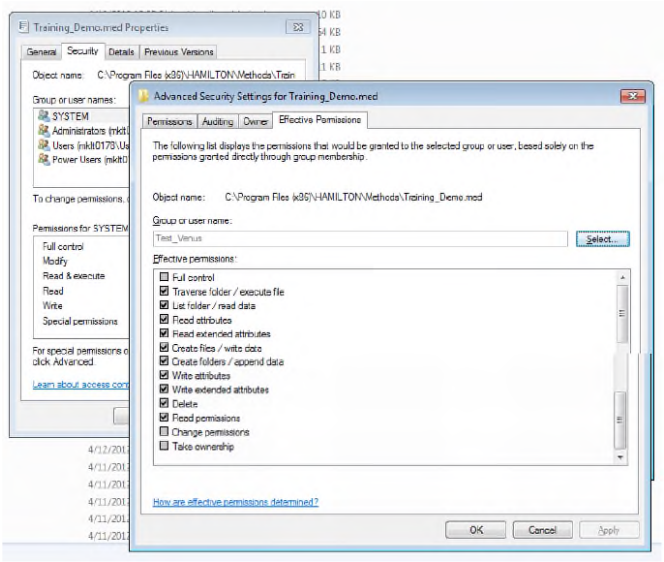


Messages to the operator can be programmed into the method to guide the operator in the correct sequence of events.

Authority - 21 CFR Part 11.10 (g)

Use of authority checks to ensure that only authorized individuals can use the system, electronically sign a record, access the operation or computer system input or output device, alter a record, or perform the operation at hand.

Five user groups have been defined and each group has a distinct level of access to the Vector/VENUS software (see table below). Methods can be controlled by saving validated methods in an access-controlled directory. Lab Operator and Lab Operator 2 user groups shall be denied access to directories containing non-validated methods while having read-only access to validated method directories (see screenshot below). Additional protection is supplied by the checksum system for all records, which protects files from inappropriate modification. The user id and time/date stamp are applied to every electronic record when it is saved.



User groups can be given different levels of access to files and directories with Windows 10 security. For example, the Lab Method Programmer will have full control to modify, execute, read and write method files, while the Lab Operator will have read-only access to method files.

User Group	Allowed Function
Lab Operator	Allowed to run validated methods
Lab Operator 2	Allowed to run validated methods and move labware using the Layout Editor
Lab Method Programmer	Allowed to change methods, labware and sequences
Lab Service	HAMILTON Trained Field Service Engineers with full access in order to install and service the system
Lab Remote Service	Allowed to only read data

These are the user groups recognized by Vector/VENUS software for the function protection features. Vector/VENUS software checks the user's user group identity to appropriately restrict their functional access to the software.

Location Checks - 21 CFR Part 11.10 (h)

Use of device (e.g. terminal checks) to determine, as appropriate, the validity of the source of data input or operational instruction.

This requirement is applicable to a source of data (e.g. plate reader) that can receive commands from more than one system, such as on a network. The source of data instrument would have to question the source of the command to ensure that only the authorized workstation is the actual source of the commands. HAMILTON workstations have hardwired connections to the computer. For unique instruments, location checks are unnecessary. In the event of identical instruments under control by the same computer, the node identity of the second instrument must be changed from the default setting during installation, creating a unique address for each of the identical instruments. This differentiates the systems to the computer. Correct instrument identification is verified during instrument installation.

Education/Training - 21 CFR Part 11.10 (i)

Determination that persons who develop, maintain, or use electronic record/electronic signature systems have the education, training, and experience to perform their assigned tasks.

HAMILTON supplies training and application support as necessary to assist in developing and maintaining application performance. After training with a HAMILTON representative, each trainee receives a certificate of training completion. This certificate can be used to document adequate training for use of a HAMILTON instrument. Internal training programs and documentation need to be developed and documented by the end user laboratory.


HAMILTON

This is to certify that

Full Name
of Company Name

Has been trained on the operation, programming, and maintenance of the

Instrument

Date

Trainer, Hamilton Company

Certificate for user training. Training certificates must be maintained by laboratory SOPs.

Written Policies - 21 CFR Part 11.10 (j)

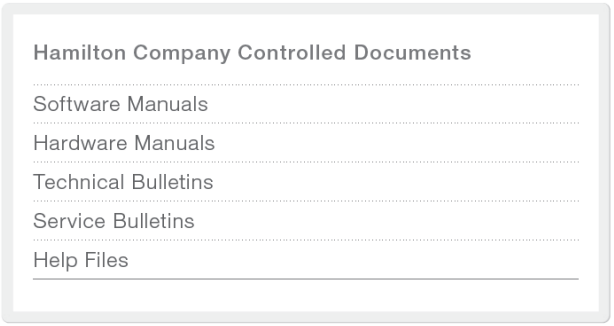
The establishment of, and adherence to, written policies that hold individuals accountable and responsible for actions, initiated under their electronic signatures, in order to deter record and signature falsification. This is a laboratory procedure, separate from HAMILTON instrumentation.

Document Controls/Audit Trails - 21 CFR Part 11.10 (k (1))

Adequate controls over the distribution of, access to, and use of documentation for system operation and maintenance.

This rule applies to the system documentation, which describes how a system operates and is maintained. System documentation includes, but is not limited to, operation manuals, help files, SOPs, access information, operating system manuals and privilege logs. This rule only pertains to documentation that can be changed by individuals within an organization. HAMILTON instrumentation documentation such as operation manuals or help files can only be changed by HAMILTON. Laboratories do not need to control these documents. Even though laboratories do not need a change control procedure for these documents, they are responsible for archiving and making them accessible to the appropriate users. Laboratories are also responsible for protecting their own documents

regarding HAMILTON instrumentation. HAMILTON regulates the distribution of access to and use of system documentation by ISO-9001 compliant procedures.



These documents are controlled by HAMILTON's change-control process. Laboratories do not need to maintain change documentation on these documents. However, the laboratory is responsible for archiving and making these documents available to the appropriate users.

Document Controls/Audit Trails - 21 CFR Part 11.10 (k (2))

Revision and change control procedures to maintain an audit trail that documents time-sequenced development and modification of systems documentation.

This rule is similar to the above in that it refers to the same documents mentioned above. HAMILTON maintains an internal change control process for documentation by procedures that are consistent with the 21 CFR Part 11 guidelines. Laboratories are responsible for the change control process of their internal documents.

8.5 Appendix E: Glossary

Term	Definition
ADC	Anti-Droplet Control to prevent drops while pipetting highly volatile solvents.
Adjustment	Detailed positional setting for the hardware.
Agency	Food and Drug Administration
Air Displacement Tip	HAMILTON CO-RE disposable tip or commercial pipetting tip.
Aliquot	Aliquots are identical small volumes of liquid.
APE	Application Engineering.
Aspirate	To draw up liquid into a pipetting device.
Autoload	Hardware assembly that enables automatic loading of the ML STAR. It consists of a loading head movable in Y direction, which draws the carriers into the ML STAR and reads the barcodes on them.
Barcode Mask	The barcode mask defines the basic structure of a barcode. It is a pattern to which a barcode must conform. The assignment of a specific labware item can be done in this manner. The barcode mask can require a barcode to contain specific strings at fixed positions. It can also contain wildcards.
Barcode Reader	Device for reading sample/plate Barcodes. Part of the Autoload.
Basic Microlab STAR	Basic parts of the ML STAR with pipetting arm and deck, to which the loading unit and the options can be added on.
Bilateral Identification	A method of verifying an individual's identity based on a user id and password system.
Biometric Identification	A method of verifying an individual's identity based on measurement of the individual's physical feature(s) or repeatable actions where those features and/or actions are both unique to that individual and measurable.
BVS	Basic Vacuum System for automation of vacuum based filtration kits. Predecessor of the CVS.
Carrier	Unit for loading plates, tubes and tips on the ML STAR deck. Can be used manually or, if possible, by the Autoload option.
Checksum	A system by which the authenticity of a file can be checked, based on the binary code. This protects files from being improperly modified. The checksum value is updated every time a file is legitimately saved. If the checksum doesn't match when the file is reopened, the file has been improperly modified and the file is blocked from use.
cLLD	Capacitive Liquid Level Detection.

Term	Definition
Closed System	An environment in which system access is controlled by persons who are responsible for the content of electronic records that are on the system.
CLT	MULTIFLEX CORE-LID TOOL; Suction cup that can be picked up by a CO-RE pipetting channel to move around lids.
Container	A container defines a tube, vessel or a single well of a plate.
Container Identification	Barcode for the identification of a container. Serves for a unique identification of a vessel (e.g. a sample test tube).
Continuous Loading	Refers to the loading of elements that can be manipulated onto the ML STAR after processing has been started.
CVS	Crystal Vacuum System for automation of vacuum based filtration kits.
Decapper Module	Hardware assembly that enables opening and closing of tubes with screw-top caps. To be used in combination with the Tube Twister Channel.
Deck	The work surface (work area) of the ML STAR. The area where the pipetting channels perform liquid handling or transport steps. The deck is divided into equal tracks, which are occupied by labware.
Deck Layout	A collection of labware placed upon a deck.
Dispense	To distribute quantities of liquid from a pipetting device.
Docking Station	The long bar at the back of the ML STAR instrument for guiding the cables and the tubing for accessories, such as Wash Stations, TCC, etc..
DWP (Deep Well Plate)	Microplate with large well volume, used when higher volumes of sample need to be stored or collected (e.g. chemistry libraries, for cell culture or filtration applications). In general we assume a plate with 96 wells (8 x 12) 9 mm wide, standard SBS format. There are also DWPs with higher or lower well number.
Electronic Records	Any combination of text, graphics, data, audio, pictorial, or other information represented in digital form that is created, modified, maintained, archived, retrieved, or distributed by a computer system.
Electronic Signature	A computer data compilation of any symbol or series of symbols executed, adopted, or authorized by an individual to be the legally binding equivalent of the individual's handwritten signature.
Firmware	Lower Level program code that is carried out on the processors of the ML STAR Instrument.

Term	Definition
Front Cover	Protective covering for the ML STAR Instrument, featuring a hinged front window made of transparent Plexiglas. With this option and assembly, the work surface of the ML STAR is covered in such a way that it is shielded from user intervention and other outside influences (such as dust). At the same time, it protects the user from the movements of the ML STAR.
Good Laboratory Practices	Abbreviated as GLP, are set of appropriate laboratory behaviors which shall be observed.
Hardware Error	Type of error that is caused by a technical problem with the hardware.
HHS	HAMILTON Heater Shaker. Unit to heat and / or shake microplates in SBS format.
HSB	Heater Shaker Box. Interface unit which is needed if more than two HHS are being used.
HSL	HAMILTON Standard Language
Instrument	Hardware of the ML STAR (mechanics, electronics, and firmware)
Instrument Commands	The commands made available by the firmware for controlling the ML STAR.
Instrument Steps	The commands made available by the firmware for controlling the ML STAR.
Labware	Refers to movable items to be placed on the ML STAR deck, such as carriers, containers, or racks.
LIMS	Higher level data processing system, generally known as Laboratory Information Management System, also LMS.
Liquid	Includes all kinds of liquids, among which are included reagents, controls, standards, wash fluids.
LLD	*Liquid Level Detection* – Detection of liquid surface which may be achieved either by pressure or capacitive signal detection.
Loading Tray	Hardware unit. The carriers can be placed on it and held outside the ML STAR. The loading tray is attached to the ML STAR, to support the automatic loading and unloading process.
Loading, Unloading	The process by which a plate, tube or tip carriers are brought on and off the ML STAR deck. This can happen automatically by means of the Autoload Option, or manually.
MAD	Monitored Air Displacement: aspiration monitoring feature. During the aspiration process, the pressure within the pipetting channel is measured in real time.
Method	The method contains all instruction that must be executed during a run.

Term	Definition
Method Files	A method in Vector/VENUS software is defined by the .med, .stp, .lay, .sub, .res and .hsl files. These six files are necessary for running a method in Vector/VENUS software.
MFX	Multiflex: A multiple-use carrier base concept.
Microplate	See MTP.
MPH	Multi-Probe Head, a pipetting device consisting of an array of pipetting channels (96 or 384).
MTP (Microtiter plate or microplate)	In general, a microtiter plate (or microplate) is assumed to have 96 wells (8 x 12) 9 mm wide. There are also plates with 384 wells (16 x 24 / 4.5 mm), or others with a different size
NTR	Nested Tip Rack
Orbit / Amplitude (of the HAMILTON Heater Shaker)	The orbit (rotation distance) is defined as peak to peak distance in one direction (e.g. distance between extreme positions in the Y-Direction of the plate measured in millimeters [mm]). The amplitude is defined as the distance from the center of the shaking movement and it is 50% of the peak to peak distance.
PN	Part Number
Pause	Interruption of processing. The current processing steps are completed.
Pipetting	Transfer of liquids from one container to another.
Pipetting Arm	Assembly equipped with the pipetting device and/or plate handler, as well as the common X-drive.
Pipetting Channel	Hardware assembly including the function of picking up a tip aspirating, dispensing, tip eject, liquid level detection and the Y/Z-movements
Pipetting Module	Firmware (-processor-program) which controls a pipetting channel, in which category are included the Y and Z pipetting movements, and the LLD.
pLLD	Pressure-based Liquid Level Detection.
Pooling	Pipetting of different liquids in one well; 1, 2, 3…to n and n to 1, 2, 3...
Processing Step	Defines what must be carried out on the ML STAR instrument, as well as the location it must be carried out and possible interaction with other system components or labware. The action is defined in accordance with the methods, the loading and the tasks.
Pump Station	Part of the needle/tip wash station. Its function is to pump wash liquid to and from the wash station.
Rack	Group of containers, as DWP, MTP, etc.
Rack Identification	Barcode for rack identification
Random Access	Means that every pipetting channel can access any position anywhere on the work area.

Term	Definition
Run	Execution of the processing steps defined in the method with the aim of processing one or more liquids and containers (e.g. MTP). The run is a series of timed commands, in order to carry out processing on the ML STAR according to the processing plan. The run can include a reloading of elements.
Run Abort	Cancelled run by the user or by the ML STAR.
Run Visualization	Visualization of the current run, reporting the status of the ML STAR.
Sample	Refers to a liquid in a unique identified container which is to be processed.
SBS Format	Standard format for microplates, defined by the Society for Biomolecular Screening.
SOP	Standard Operating Procedure.
Stacker	Storage unit for racks.
T	Abbreviation used for "track". The ML STAR instruments have equal partitions of 22.5 mm, equivalent to 1-T. Labware carriers are adapted to those partitions.
TADM	Total Aspiration and Dispense Monitoring. The pressure inside each individual pipetting channel is monitored, during aspirate and dispense.
TCC	Temperature controlled carrier.
Tip	Disposable tip for pipetting.
Tip Rack	Frame that holds the tips.
Tip Waste	Container for ejected tips.
Touch-Off	Type of dispensing where the tip approaches the bottom of the empty container so close as to allow the dispensed droplet to have simultaneous contact with the tip and the container bottom.
Trace	Record of the status during processing.
Trace Files	The file generated by Vector/VENUS software for every runtime event. The trace file contains all the events of the run, date, time and user id information.
Tube	A container for liquid, usually having a circular cross-section, and a cylindrical length section.
Tube Twister Channel	Hardware assembly including the function of picking up and transporting of a tube, spin the tube (mix the liquid within the tube) as well as identify the barcode of the tube.
User	User of the software. Access rights for different types of users can be defined, such as operators, laboratory managers, etc.
Validation	Confirmation by examination and provision of objective evidence that the system specifications conform to user needs and intended uses, and that all requirements can be consistently fulfilled.

Term	Definition
Vector/VENUS Software	Vector/VENUS software is the software designed for HAMILTON robotic instrumentation control. The software provisions for 21 CFR Part 11 applies to all instruments controlled by the software including the ML STAR instruments.
Verification Kit	Balance, liquid, disposable tips to verify the function (volume check) of the ML STAR pipetting heads.
Waste Container	A device on the ML STAR deck to collect used disposable tips.
Well	The individual container of a MTP or DWP.
Well Type	Geometrical shape of the well, such as U, V or flat.
Side Touch	Type of dispensing whereby the tip or the needle touches the side of a container and thus releases the droplet.
Work Area	The area of the ML STAR to which access is provided during the processing. Elements to be pipetted or handled can be placed in this area.
Worklist	Information according to which a method is to be executed on the ML STAR. A worklist may contain different parameters (e.g. Pipetting volume, heating temperature, shaking speed, etc.).

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